

Donor 7703

Genetic Testing Summary

Fairfax Cryobank recommends reviewing this genetic testing summary with your healthcare provider to determine suitability.

Last Updated: 05/21/2025

Donor Reported Ancestry: German, Polish, English Jewish Ancestry: No

Genetic Test*	Result	Comments/Donor's Residual Risk**
Chromosome analysis (karyotype)	Normal male karyotype	No evidence of clinically significant chromosome abnormalities
Hemoglobin evaluation	Normal hemoglobin fractionation and MCV/MCH results	Reduced risk to be a carrier for sickle cell anemia, beta thalassemia, alpha thalassemia trait (aa/ and a-/a-) and other hemoglobinopathies
Expanded Genetic Disease Carrier Screening Panel attached- 549 diseases by gene sequencing.	Carrier: Familial Hemophagocytic Lymphohistiocytosis, (UNC13D- Related)	Partner testing is recommended before using this donor.
	Carrier: Walker-Warburg Syndrome (POMT2-Related)	
Special Testing	Negative for other genes sequenced.	
Gene(s): IL36RN, TCTN1	Negative via gene sequencing with deletion and duplication analysis	

^{*}No single test can screen for all genetic disorders. A negative screening result significantly reduces, but cannot eliminate, the risk for these conditions in a pregnancy.

^{**}Donor residual risk is the chance the donor is still a carrier after testing negative.

Patient Name: **Donor 7703**

Date Of Birth:

Gender:

Male

N/A

Ethnicity:

Caucasian/Non-Hispanic

White

Patient ID:

Medical Record #: 7703

Collection Kit:

Accession ID: N/A

Case File ID:

Test Information

Ordering Physician:

Clinic Information: Fairfax Cryobank

Phone: N/A

Report Date: 07/23/2024 07/05/2024 Sample Collected:

Sample Received: 07/06/2024

Sample Type: Blood



CARRIER SCREENING REPORT

ABOUT THIS SCREEN: Horizon™ is a carrier screen for specific autosomal recessive and Xlinked diseases. This information can help patients learn their risk of having a child with specific genetic conditions.

ORDER SELECTED: The Horizon Custom panel was ordered for this patient. Males are not

screened for X-linked diseases

FINAL RESULTS SUMMARY:



CARRIER for Familial Hemophagocytic Lymphohistiocytosis, UNC13D-Related

Positive for the pathogenic variant c.177_178del (p.Y61Hfs*10) in the UNC13D gene. If this individual's partner is a carrier for FAMILIAL HEMOPHAGOCYTIC LYMPHOHISTIOCYTOSIS, UNC13D-RELATED, their chance to have a child with this condition is 1 in 4 (25%). Carrier screening for this individual's partner is suggested.

CARRIER for Walker-Warburg Syndrome, POMT2-Related

Positive for the likely pathogenic variant c.1238G>C (p.R413P) in the POMT2 gene. If this individual's partner is a carrier for WALKER-WARBURG SYNDROME, POMT2-RELATED, their chance to have a child with this condition may be as high as 1 in 4 (25%). Carrier screening for this individual's partner is suggested.

Negative for 547 out of 549 diseases

No other pathogenic variants were detected in the genes that were screened. The patient's remaining carrier risk after the negative screening results is listed for each disease/gene on the Horizon website at https://www.natera.com/panel-option/h-all/. Please see the following pages of this report for a comprehensive list of all conditions included on this individual's screen.

Carrier screening is not diagnostic and may not detect all possible pathogenic variants in a given gene.

RECOMMENDATIONS

Individuals who would like to review their Horizon report with a Natera Laboratory Genetic Counselor may schedule a telephone genetic information session by calling 650-249-9090 or visiting naterasession.com. Clinicians with questions may contact Natera at 650-249-9090 or email support@natera.com. Individuals with positive results may wish to discuss these results with family members to allow them the option to be screened. Comprehensive genetic counseling to discuss the implications of these test results and possible associated reproductive risk is recommended.

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Patient Name: **Donor 7703** **Test Information**

Ordering Physician:

Fairfax Cryobank

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WALKER-WARBURG SYNDROME, POMT2-RELATED

Understanding Your Horizon Carrier Screen Results

What is Walker-Warburg Syndrome, POMT2-Related?

Walker-Warburg Syndrome, POMT2-Related is an inherited disorder that affects many parts of the body, especially the brain, eyes, and muscles. Signs and symptoms are often present before birth but sometimes start in infancy and include weak muscle tone (hypotonia), excess fluid on the brain (hydrocephalus), severe brain abnormalities, and eye defects with vision problems. Infants and children with Walker-Warburg Syndrome, POMT2-Related have worsening muscle weakness, problems with movement and coordination, seizures, and severe developmental delay with intellectual disability. Although symptoms vary from person to person, lifespan is usually shortened with death often occurring in early childhood. Currently, there is no cure or specific treatment for this disorder. Clinical trials involving potential new treatments for this condition may be available (see www.clinicaltrials.gov). Rarely, mutations in the same pair of genes cause a related condition: either Muscle-Eye-Brain Disease; Congenital Muscular Dystrophy, POMT2-Related; or Limb-Girdle Muscular Dystrophy, Type C2. Muscle-Eye-Brain Disease has similar symptoms to Walker-Warburg Syndrome, POMT2-Related but is usually less severe with a longer life span. Congenital Muscular Dystrophy, POMT2-Related has symptoms that include brain abnormalities, muscle weakness, and intellectual disability with lack of speech, but vision is usually normal. Limb-Girdle Muscular Dystrophy, Type C2 symptoms include severe muscle weakness in the shoulder and hip areas that usually starts in childhood, with intelligence being normal in most affected individuals. The information below is about Walker-Warburg Syndrome, POMT2-Related, the most common condition. However, the inheritance pattern and reproductive options listed below apply to all of the conditions described above.

What causes Walker-Warburg Syndrome, POMT2-Related?

Walker-Warburg Syndrome, POMT2-Related is caused by a change, or mutation, in both copies of the POMT2 gene pair. These mutations cause the genes to not work properly or not work at all. When both copies of the POMT2 gene do not work correctly, it leads to the symptoms described above. It is sometimes, but not always, possible to determine which of the above conditions a specific mutation in the POMT2 gene will cause. Walker-Warburg Syndrome, POMT2-Related is inherited in an autosomal recessive manner. This means that, in most cases, both parents must be carriers of a mutation in one copy of the POMT2 gene to have a child with Walker-Warburg Syndrome, POMT2-Related. People who are carriers for Walker-Warburg Syndrome, POMT2-Related are usually healthy and do not have symptoms, nor do they have the disorder themselves. Usually a child inherits two copies of each gene, one copy from the mother and one copy from the father. If the mother and father are both carriers for Walker-Warburg Syndrome, POMT2-Related or related condition there is a 1 in 4, or 25%, chance in each pregnancy for both partners to pass on their POMT2 gene mutations to the child, who will then have the disorder. Individuals found to carry more than one mutation for Walker-Warburg Syndrome, POMT2-Related should discuss their risk for having an affected child, and any potential effects to their own health, with their health care provider. There are a number of other forms of Walker-Warburg Syndrome, Limb-Girdle Muscular Dystrophy, Congenital Muscular Dystrophy, and Muscle-Eye-Brain Disease, each caused by mutations in different genes. A person who carries a mutation in the POMT2 gene is not likely to be at increased risk for having children with the other forms of these disorders.

What can I do next?

You may wish to speak with a local genetic counselor about your carrier test results. A genetic counselor in your area can be located on the National Society of Genetic Counselors website (www.nsgc.org). Your siblings and other relatives are at increased risk to also have this mutation. You are encouraged to inform your family members of your test results as they may wish to consider being tested themselves. If you are pregnant, your partner can have carrier screening for Walker-Warburg Syndrome, POMT2-Related ordered by a health care professional. If your partner is not found to be a carrier for Walker-Warburg Syndrome, POMT2-Related, your risk of having an affected child is greatly reduced. Couples at risk of having a baby with Walker-Warburg Syndrome, POMT2-Related can opt to have prenatal diagnosis done through chorionic villus sampling (CVS) or amniocentesis during pregnancy or can choose to have the baby tested after birth for this condition. If you are not yet pregnant, your partner can have carrier screening for Walker-Warburg Syndrome, POMT2-Related ordered by a health care professional. If your partner is found to be a carrier for Walker-Warburg Syndrome, you have several reproductive options to consider:

- Natural pregnancy with or without prenatal diagnosis of the fetus or testing the baby after birth for Walker-Warburg Syndrome, POMT2-Related or related
- Preimplantation genetic diagnosis (PGD) with in vitro fertilization (IVF) to test embryos for Walker-Warburg Syndrome, POMT2-Related or related disorder
- Adoption or use of a sperm or egg donor who is not a carrier for Walker-Warburg Syndrome, POMT2-Related or related disorder

What resources are available?

- Genetics Home Reference: http://ghr.nlm.nih.gov/condition/walker-warburg-syndrome
- Prenatal diagnosis done through CVS: http://www.marchofdimes.org/chorionic-villus-sampling.aspx
- Prenatal diagnosis done through Amniocentesis: http://www.marchofdimes.org/amniocentesis.aspx
- PGD with IVF: http://www.natera.com/spectrum



Patient Information
Patient Name:





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VARIANT DETAILS

POMT2, c.1238G>C (p.R413P), heterozygous, likely pathogenic

- The c.1238G>C (p.R413P) variant in the POMT2 gene has been observed at a frequency of 0.0028% in the gnomAD v2.1.1 dataset.
- This variant has been reported in a homozygous state or in conjunction with another variant in individual(s) with muscular dystrophy-dystroglycanopathy, type A, 2 (PMID: 17878207, 26013959, 30060766).
- This variant has been reported in ClinVar [ID: 3225].

UNC13D, c.177_178del (p.Y61Hfs*10), heterozygous, pathogenic

- The c.177_178del (p.Y61Hfs*10) variant in the UNC13D gene has not been observed in the gnomAD v2.1.1 dataset.
- This variant has been reported in a homozygous state or in conjunction with another variant in individual(s) with Familial hemophagocytic lymphohistiocytosis (PMID: 16278825).
- This premature termination variant is predicted to cause nonsense-mediated decay (NMD) in a gene where loss-of-function is a known mechanism of disease.
- This variant has been described in ClinVar [ID: 1074706].



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DISEASES SCREENED

Below is a list of all diseases screened and the result. Certain conditions have unique patient-specific numerical values, therefore, results for those conditions are formatted differently.

Autosomal Recessive

17-BETA HYDROXYSTEROID DEHYDROGENASE 3 DEFICIENCY (HSD17B3) negative

3-BETA-HYDROXYSTEROID DEHYDROGENASE TYPE II DEFICIENCY (HSD3B2) negative 3-HYDROXY-3-METHYLGLUTARYL-COENZYME A LYASE DEFICIENCY (HMGCL) negative 3-HYDROXYACYL-COA DEHYDROGENASE DEFICIENCY (HADH) negative 3-METHYLCROTONYL-CoA CARBOXYLASE 2 DEFICIENCY (MCCC2) negative 3-PHOSPHOGLYCERATE DEHYDROGENASE DEFICIENCY (PHGDH) negative

5-ALPHA-REDUCTASE DEFICIENCY (SRD5A2) negative

6-PYRUVOYL-TETRAHYDROPTERIN SYNTHASE (PTPS) DEFICIENCY (PTS) negative

ABCA4-RELATED CONDITIONS (ABCA4) negative ABETALIPOPROTEINEMIA (MTTP) negative ACHONDROGENESIS, TYPE 1B (SLC26A2) negative ACHROMATOPSIA, CNGB3-RELATED (CNGB3) negative
ACRODERMATITIS ENTEROPATHICA (SLC39A4) negative
ACTION MYOCLONUS-RENAL FAILURE (AMRF) SYNDROME (SCARB2) negative ACUTE INFANTILE LIVER FAILURE, TRMU-RELATED (TRMU) negative ACYL-COA OXIDASE I DEFICIENCY (ACOX1) negative AICARDI-GOUTIÈRES SYNDROME (SAMHD1) negative AICARDI-GOUTIERES SYNDROME, RNASEH2A-RELATED (RNASEH2A) negative AICARDI-GOUTIERES SYNDROME, RNASEH/2B-RELATED (RNASEH/2B) negative AICARDI-GOUTIERES SYNDROME, RNASEH/2C-RELATED (RNASEH/2C) negative AICARDI-GOUTIÈRES SYNDROME, TREX1-RELATED (TREX1) negative

ALPHA-MANNOSIDOSIS (MAN2B1) negative ALPHA-THALASSEMIA (HBA1/HBA2) negative ALPORT SYNDROME, COL4A3-RELATED (COL4A3) negative ALPORT SYNDROME, COL4A4-RELATED (COL4A4) negative

ALSTROM SYNDROME (ALMS1) negative
AMISH INFANTILE EPILEPSY SYNDROME (573GAL5) negative
ANDERMANN SYNDROME (SLC12A6) negative

ARGININE:GLYCINE AMIDINOTRANSFERASE DEFICIENCY (AGAT DEFICIENCY)

(GATM) negative
ARGININEMIA (ARG1) negative
ARGININOSUCCINATE LYASE DEFICIENCY (ASL) negative

ARGINIOSOCCINATE L'IASE DEFICIENCY (ASL) negative AROMATASE DEFICIENCY (CYP19A1) negative ASPARAGINE SYNTHETASE DEFICIENCY (ASNS) negative ASPARTYLGLYCOSAMINURIA (AGA) negative ATAXIA WITH VITAMIN E DEFICIENCY (TTPA) negative

ATAXIA-TELANGIECTASIA (ATM) negative
ATAXIA-TELANGIECTASIA-LIKE DISORDER 1 (MRE11) negative

ATRANSFERRINEMIA (TF) negative

AUTISM SPECTRUM, EPILEPSY AND ARTHROGRYPOSIS (SLC35A3) negative AUTOIMMUNE POLYGLANDULAR SYNDROME, TYPE 1 (AIRE) negative AUTOSOMAL RECESSIVE CONGENITAL ICHTHYOSIS (ARCI), SLC27A4-RELATED

(SLC27A4) negative

AUTOSOMAL RECESSIVE SPASTIC ATAXIA OF CHARLEVOIX-SAGUENAY (SACS) negative

BARDET-BIEDL SYNDROME, ARL6-RELATED (ARL6) negative BARDET-BIEDL SYNDROME, BBS10-RELATED (BBS10) negative BARDET-BIEDL SYNDROME, BBS12-RELATED (BBS12) negative BARDET-BIEDL SYNDROME, BBS1-RELATED (BBS1) negative BARDET-BIEDL SYNDROME, BBS2-RELATED (BBS2) negative BARDET-BIEDL SYNDROME, BBS4-RELATED (BBS4) negative BARDET-BIEDL SYNDROME, BBS5-RELATED (BBS5) negative BARDET-BIEDL SYNDROME, BBS7-RELATED (BBS7) negative BARDET-BIEDL SYNDROME, BBS9-RELATED (BBS9) negative BARDET-BIEDL SYNDROME, TTC8-RELATED (TTC8) negative BART LYMPHOCYTE SYNDROME, CIITA-RELATED (CIITA) negative BARTTER SYNDROME, BSND-RELATED (BSND) negative BARTTER SYNDROME, KCNJ1-RELATED (KCNJ1) negative BARTTER SYNDROME, SLC12A1-RELATED (SLC12A1) negative BATTEN DISEASE, CLN3-RELATED (CLN3) negative BETA-HEMOGLOBINOPATHIES (HBB) negative BETA-KETOTHIOLASE DEFICIENCY (ACAT1) negative BETA-MANNOSIDOSIS (MANBA) negative
BETA-UREIDOPROPIONASE DEFICIENCY (UPB1) negative
BILATERAL FRONTOPARIETAL POLYMICROGYRIA (GPR56) negative BIOTINIDASE DEFICIENCY (BTD) negative BIOTIN-THIAMINE-RESPONSIVE BASAL GANGLIA DISEASE (BTBGD) (SLC19A3) negative BLOOM SYNDROME (BLM) negative BRITTLE CORNEA SYNDROME 1 (ZNF469) negative BRITTLE CORNEA SYNDROME 2 (PRDM5) negative

CANAVAN DISEASE (ASPA) negative CARBAMOYL PHOSPHATE SYNTHETASE I DEFICIENCY (CPS1) negative CARNITINE DEFICIENCY (SLC22A5) negative CARNITINE PALMITOYLTRANSFERASE IA DEFICIENCY (CPT1A) negative CARNITINE PALMITOYLTRANSFERASE II DEFICIENCY (CPT2) negative

CARNITINE-ACYLCARNITINE TRANSLOCASE DEFICIENCY (SLC25A20) negative CARPENTER SYNDROME (RAB23) negative
CARTILAGE-HAIR HYPOPLASIA (RMRP) negative
CATECHOLAMINERGIC POLYMORPHIC VENTRICULAR TACHYCARDIA (CASQ2) negative

CD59-MEDIATED HEMOLYTIC ANEMIA (CD59) negative

CEP152-RELATED MICROCEPHALY (CEP152) negative CEREBRAL DYSGENESIS, NEUROPATHY, ICHTHYOSIS, AND PALMOPLANTAR KERATODERMA (CEDNIK) SYNDROME (SNAP29) negative

CEREBROTENDINOUS XANTHOMATOSIS (CYP27A1) negative CHARCOT-MARIE-TOOTH DISEASE, RECESSIVE INTERMEDIATE C (PLEKHG5) negative CHARCOT-MARIE-TOOTH-DISEASE, TYPE 4D (NDRG1) negative

CHEDIAK-HIGASHI SYNDROME (LYST) negative

CHOREOACANTHOCYTOSIS (VP513A) negative CHRONIC GRANULOMATOUS DISEASE, CYBA-RELATED (CYBA) negative

CHRONIC GRANULOMATOUS DISEASE, NCF2-RELATED (NCF2) negative

CILIOPATHIES, RPGRIP1L-RELATED (RPGRIP1L) negative CITRIN DEFICIENCY (SLC25A13) negative CITRULLINEMIA, TYPE 1 (ASS1) negative

CLN10 DISEASE (CTSD) negative

COHEN SYNDROME (VPS13B) negative COL11A2-RELATED CONDITIONS (COL11A2) negative COMBINED MALONIC AND METHYLMALONIC ACIDURIA (ACSF3) negative

COMBINED OXIDATIVE PHOSPHORYLATION DEFICIENCY 1 (GFM1) negative COMBINED OXIDATIVE PHOSPHORYLATION DEFICIENCY 3 (TSFM) negative COMBINED PITUITARY HORMONE DEFICIENCY 1 (POU1F1) negative

COMBINED PITUITARY HORMONE DEFICIENCY-2 (PROP1) negative

CONGENITAL ADRENAL HYPERPLASIA, 11-BETA-HYDROXYLASE DEFICIENCY

CONGENITAL ADRENAL HYPERPLASIA, 17-ALPHA-HYDROXYLASE DEFICIENCY (CYP17A1) negative
CONGENITAL ADRENAL HYPERPLASIA, 21-HYDROXYLASE DEFICIENCY

(CYP21A2) negative

CONGENITAL ADRENAL INSUFFICIENCY, CYP11A1-RELATED (CYP11A1) negative CONGENITAL AMEGAKARYOCYTIC THROMBOCYTOPENIA (MPL) negative
CONGENITAL CHRONIC DIARRHEA (DGAT1) negative
CONGENITAL DISORDER OF GLYCOSYLATION TYPE 1, ALG1-RELATED (ALG1) negative

CONGENITAL DISORDER OF GLYCOSYLATION, TYPE 1A, PMM2-Related (PMM2) negative CONGENITAL DISORDER OF GLYCOSYLATION, TYPE 1B (MPI) negative CONGENITAL DISORDER OF GLYCOSYLATION, TYPE 1C (ALG6) negative

CONGENITAL DYSERYTHROPOIETIC ANEMIA TYPE 2 (SEC23B) negative

CONGENITAL FINNISH NEPHROSIS (NPHS1) negative
CONGENITAL HYDROCEPHALUS 1 (CCDC88C) negative
CONGENITAL HYPERINSULINISM, KCNJ11-Related (KCNJ11) negative

CONGENITAL HYPERINSULINISM, RCNJ11-Related (RCNJ11) negative CONGENITAL INSENSITIVITY TO PAIN WITH ANHIDROSIS (CIPA) (NTRK1) negative CONGENITAL MYASTHENIC SYNDROME, CHAT-RELATED (CHAT) negative CONGENITAL MYASTHENIC SYNDROME, CHRNE-RELATED (CHRNE) negative CONGENITAL MYASTHENIC SYNDROME, COLQ-RELATED (COLQ) negative CONGENITAL MYASTHENIC SYNDROME, DOK7-RELATED (DOK7) negative CONGENITAL MYASTHENIC SYNDROME, RAPSN-RELATED (RAPSN) negative

CONGENITAL NEPHROTIC SYNDROME, PLCE1-RELATED (PLCE1) negative

CONGENITAL NEUTROPENIA, G6PC3-RELATED (G6PC3) negative CONGENITAL NEUTROPENIA, HAX1-RELATED (HAX1) negative CONGENITAL NEUTROPENIA, VPS45-RELATED (VPS45) negative

CONGENITAL SECRETORY CHLORIDE DIARRHEA 1 (SLC26A3) negative CORNEAL DYSTROPHY AND PERCEPTIVE DEAFNESS (SLC4A11) negative CORTICOSTERONE METHYLOXIDASE DEFICIENCY (CYP11B2) negative

COSTEFF SYNDROME (3-METHYLGLUTACONIC ACIDURIA, TYPE 3) (OPA3) negative

CRB1-RELATED RETINAL DYSTROPHIES (CRB1) negative

CYSTIC FIBROSIS (CFTR) negative

CYSTINOSIS (CTNS) negative

CYTOCHROME C OXIDASE DEFICIENCY, PET100-RELATED (PET100) negative

CYTOCHROME P450 OXIOREDUCTASE DEFICIENCY (POR) negative

D-BIFUNCTIONAL PROTEIN DEFICIENCY (HSD17B4) negative



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DEAFNESS, AUTOSOMAL RECESSIVE 77 (LOXHD1) negative DIHYDROPTERIDINE REDUCTASE (DHPR) DEFICIENCY (QDPR) negative DONNAI-BARROW SYNDROME (LRP2) negative

DUBIN-JOHNSON SYNDROME (ABCC2) negative DYSKERATOSIS CONGENITA SPECTRUM DISORDERS (TERT) negative DYSKERATOSIS CONGENITA, RTEL1-RELATED (RTEL1) negative

DYSTROPHIC EPIDERMOLYSIS BULLOSA, COL7A1-Related (COL7A1) negative

EARLY INFANTILE EPILEPTIC ENCEPHALOPATHY, CAD-RELATED (CAD) negative EHLERS-DANLOS SYNDROME TYPE VI (PLOD1) negative EHLERS-DANLOS SYNDROME, CLASSIC-LIKE, TNXB-RELATED (TNXB) negative EHLERS-DANLOS SYNDROME, TYPE VII C (ADAMTS2) negative ELLIS-VAN CREVELD SYNDROME, EVC2-RELATED (EVC2) negative
ELLIS-VAN CREVELD SYNDROME, EVC-RELATED (EVC) negative
ENHANCED S-CONE SYNDROME (NR2E3) negative

EPIMERASE DEFICIENCY (GALACTOSEMIA TYPE III) (GALE) negative
EPIPHYSEAL DYSPLASIA, MULTIPLE, 7/DESBUQUOIS DYSPLASIA 1 (CANT1) negative
ERCC6-RELATED DISORDERS (ERCC6) negative

ERCC8-RELATED DISORDERS (ERCC8) negative

ETHYLMALONIC ENCEPHALOPATHY (ETHE1) negative

FACTOR XI DEFICIENCY (F11) negative

FAMILIAL DYSAUTONOMIA (IKBKAP) negative
FAMILIAL DYSAUTONOMIA (IKBKAP) negative
FAMILIAL HEMOPHAGOCYTIC LYMPHOHISTIOCYTOSIS, PRF1-RELATED (PRF1) negative
FAMILIAL HEMOPHAGOCYTIC LYMPHOHISTIOCYTOSIS, STX11-RELATED (STX11) negative
FAMILIAL HEMOPHAGOCYTIC LYMPHOHISTIOCYTOSIS, STXBP2-RELATED
(STXBP2) negative

FAMILIAL HEMOPHAGOCYTIC LYMPHOHISTIOCYTOSIS, UNC13D-RELATED (UNC13D) see

FAMILIAL HYPERCHOLESTEROLEMIA, LDLRAP1-RELATED (LDLRAP1) negative FAMILIAL HYPERCHOLESTEROLEMIA, LDLR-RELATED (LDLR) negative FAMILIAL HYPERINSULINISM, ABCC8-RELATED (ABCC8) negative FAMILIAL HYPERINSULINISM NEGATIVE NEGAT

FAMILIAL NEPHROGENIC DIABETES INSIPIDUS, AQP2-RELATED (AQP2) negative FANCONI ANEMIA, GROUP A (FANCA) negative FANCONI ANEMIA, GROUP C (FANCC) negative

FANCONI ANEMIA, GROUP D2 (FANCD2) negative FANCONI ANEMIA, GROUP E (FANCE) negative FANCONI ANEMIA, GROUP F (FANCF) negative

FANCONI ANEMIA, GROUP F (FANCF) negative FANCONI ANEMIA, GROUP I (FANCG) negative FANCONI ANEMIA, GROUP J (BRIP1) negative FANCONI ANEMIA, GROUP L (FANCL) negative FANCONI ANEMIA, GROUP L (FANCL) negative

FARBER LIPOGRANULOMATOSIS (ASAH1) negative FOVEAL HYPOPLASIA (SLC38A8) negative FRASER SYNDROME 3, GRIP1-RELATED (GRIP1) negative FRASER SYNDROME, FRAS1-RELATED (FRAS1) negative

FRASER SYNDROME, FREM2-RELATED (FREM2) negative FRIEDREICH ATAXIA (FXN) negative FRUCTOSE-1,6-BISPHOSPHATASE DEFICIENCY (FBP1) negative

FUCOSIDOSIS, FUCA1-RELATED (FUCA1) negative FUMARASE DEFICIENCY (FH) negative

GABA-TRANSAMINASE DEFICIENCY (ABAT) negative GALACTOKINASE DEFICIENCY (GALACTOSEMIA, TYPE II) (GALK1) negative

GALACTOSEMIA (GALT) negative

GALACTOSIALIDOSIS (CTSA) negative GAUCHER DISEASE (GBA) negative GCH1-RELATED CONDITIONS (GCH1) negative

GDF5-RELATED CONDITIONS (GDF5) negative

GERODERMA OSTEODYSPLASTICA (GORAB) negative GITELMAN SYNDROME (SLC12A3) negative GLANZMANN THROMBASTHENIA (ITGB3) negative

GLUTARIC ACIDEMIA, TYPE 1 (GCDH) negative GLUTARIC ACIDEMIA, TYPE 2A (ETFA) negative GLUTARIC ACIDEMIA, TYPE 2B (ETFB) negative

GLUTARIC ACIDEMIA, TYPE 2C (ETFDH) negative

GLUTARIC ACIDEMIA, 17PE 2C (E1FDH) negative
GLUTATHIONE SYNTHETASE DEFICIENCY (GSS) negative
GLYCINE ENCEPHALOPATHY, AMT-RELATED (AMT) negative
GLYCINE ENCEPHALOPATHY, GLDC-RELATED (GLDC) negative
GLYCOGEN STORAGE DISEASE TYPE 5 (McArdle Disease) (PYGM) negative
GLYCOGEN STORAGE DISEASE TYPE IXB (PHKB) negative
GLYCOGEN STORAGE DISEASE TYPE IXC (PHKG2) negative

GLYCOGEN STORAGE DISEASE, TYPE 1a (G6PC) negative

GLYCOGEN STORAGE DISEASE, TYPE 1b (SLC37A4) negative GLYCOGEN STORAGE DISEASE, TYPE 2 (POMPE DISEASE) (GAA) negative

GLYCOGEN STORAGE DISEASE, TYPE 3 (AGL) negative

GLYCOGEN STORAGE DISEASE, TYPE 4 (GBE1) negative

GLYCOGEN STORAGE DISEASE, TYPE 7 (PFKM) negative GRACILE SYNDROME (BC511) negative
GUANIDINOACETATE METHYLTRANSFERASE DEFICIENCY (GAMT) negative

HARLEQUIN ICHTHYOSIS (ABCA12) negative HEME OXYGENASE 1 DEFICIENCY (HMOX1) negative

HEMOCHROMATOSIS TYPE 2A (HFE2) negative
HEMOCHROMATOSIS, TYPE 3, TFR2-Related (TFR2) negative
HEPATOCEREBRAL MITOCHONDRIAL DNA DEPLETION SYNDROME, MPV17-RELATED

(MPV17) negative

HEREDITARY FRUCTOSE INTOLERANCE (ALDOB) negative
HEREDITARY HEMOCHROMATOSIS TYPE 2B (HAMP) negative
HEREDITARY SPASTIC PARAPARESIS, TYPE 49 (TECPR2) negative

HEREDITARY SPASTIC PARAPLEGIA, CYP7B1-RELATED (CYP7B1) negative HERMANSKY-PUDLAK SYNDROME, AP3B1-RELATED (AP3B1) negative HERMANSKY-PUDLAK SYNDROME, BLOC1S3-RELATED (BLOC1S3) negative

HERMANSKY-PUDLAK SYNDROME, BLOC156-RELATED (BLOC156) negative HERMANSKY-PUDLAK SYNDROME, HPS1-RELATED (HPS1) negative HERMANSKY-PUDLAK SYNDROME, HPS3-RELATED (HPS3) negative

HERMANSKY-PUDLAK SYNDROME, HPS4-RELATED (HPS4) negative HERMANSKY-PUDLAK SYNDROME, HPS5-RELATED (HPS5) negative HERMANSKY-PUDLAK SYNDROME, HPS6-RELATED (HPS6) negative

HOLOCARBOXYLASE SYNTHETASE DEFICIENCY (HLCS) negative

HOMOCYSTINURIA AND MEGALOBLASTIC ANEMIA TYPE CBLG (MTR) negative HOMOCYSTINURIA DUE TO DEFICIENCY OF MTHFR (MTHFR) negative HOMOCYSTINURIA, CBS-RELATED (CBS) negative

HOMOCYSTINURIA, CBS-RELATED (CBS) negative
HOMOCYSTINURIA, Type cblE (MTRR) negative
HYDROLETHALUS SYNDROME (HYLS1) negative
HYPER-IGM IMMUNODEFICIENCY (CD40) negative
HYPERORNITHINEMIA-HYPERAMMONEMIA-HOMOCITRULLINURIA (HHH SYNDROME) (SLC25A15) negative HYPERPHOSPHATEMIC FAMILIAL TUMORAL CALCINOSIS, GALNT3-RELATED

(GALNT3) negative

HYPOMYELINATING LEUKODYSTROPHY 12 (VPS11) negative HYPOPHOSPHATASIA, ALPL-RELATED (ALPL) negative

IMERSLUND-GRÄSBECK SYNDROME 2 (AMN) negative

IMMUNODEFICIENCY-CENTROMERIC INSTABILITY-FACIAL ANOMALIES (ICF) SYNDROME, DNMT3B-RELATED (DNMT3B) negative

SYNDROME, DININI 3B-RELATED (DININI 3B) negative IMMUNODEFICIENCY-CENTROMERIC INSTABILITY-FACIAL ANOMALIES (ICF) SYNDROME, ZBTB24-RELATED (ZBTB24) negative INCLUSION BODY MYOPATHY 2 (GNE) negative INFANTILE CEREBRAL AND CEREBELLAR ATROPHY (MED17) negative

INFANTILE NEPHRONOPHTHISIS (INVS) negative INFANTILE NEUROAXONAL DYSTROPHY (PLA2G6) negative ISOLATED ECTOPIA LENTIS (ADAMTSL4) negative

ISOLATED SULFITE OXIDASE DEFICIENCY (SUOX) negative

ISOLATED THYROID-STIMULATING HORMONE DEFICIENCY (TSHB) negative ISOVALERIC ACIDEMIA (IVD) negative

JOHANSON-BLIZZARD SYNDROME (*UBR1*) negative JOUBERT SYNDROME 2 / MECKEL SYNDROME 2 (*TMEM216*) negative

JOUBERT SYNDROME AND RELATED DISORDERS (JSRD), TMEM67-RELATED

(TMEM67) negative
JOUBERT SYNDROME, AHI1-RELATED (AHI1) negative

JOUBERT SYNDROME, ARL13B-RELATED (ARL13B) negative

JOUBERT SYNDROME, B9D1-RELATED (B9D1) negative
JOUBERT SYNDROME, B9D2-RELATED (B9D2) negative
JOUBERT SYNDROME, C2CD3-RELATED/OROFACIODIGITAL SYNDROME 14

(C2CD3) negative

JOUBERT SYNDROME, CC2D2A-RELATED/COACH SYNDROME (CC2D2A) negative
JOUBERT SYNDROME, CEP104-RELATED (CEP104) negative
JOUBERT SYNDROME, CEP120-RELATED/SHORT-RIB THORACIC DYSPLASIA 13 WITH OR

WITHOUT POLYDACTYLY (CEP120) negative
JOUBERT SYNDROME, CEP41-RELATED (CEP41) negative
JOUBERT SYNDROME, CPLANE1-RELATED / OROFACIODIGITAL SYNDROME 6

(CPLANE1) negative

JOUBERT SYNDROME, CSPP1-RELATED (CSPP1) negative

JOUBERT SYNDROME, INPPSE-RELATED (INPPSE) negative

JUNCTIONAL EPIDERMOLYSIS BULLOSA, COL17A1-RELATED (COL17A1) negative JUNCTIONAL EPIDERMOLYSIS BULLOSA, ITGA6-RELATED (ITGA6) negative
JUNCTIONAL EPIDERMOLYSIS BULLOSA, ITGB4-RELATED (ITGB4) negative
JUNCTIONAL EPIDERMOLYSIS BULLOSA, LAMB3-RELATED (LAMB3) negative

JUNCTIONAL EPIDERMOLYSIS BULLOSA, LAMC2-RELATED (LAMC2) negative
JUNCTIONAL EPIDERMOLYSIS BULLOSA/LARYNGOONYCHOCUTANEOUS SYNDROME,
LAMA3-RELATED (LAMA3) negative

KRABBE DISEASE (GALC) negative



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LAMELLAR ICHTHYOSIS, TYPE 1 (TGM1) negative LARON SYNDROME (GHR) negative LEBER CONGENITAL AMAUROSIS 2 (RPE65) negative LEBER CONGENITAL AMAUROSIS TYPE AIPL1 (AIPL1) negative LEBER CONGENITAL AMAUROSIS TYPE GUCY2D (GUCY2D) negative LEBER CONGENITAL AMAUROSIS TYPE TULP1 (TULP1) negative LEBER CONGENITAL AMAUROSIS, IQCB1-RELATED/SENIOR-LOKEN SYNDROME 5 (IQCB1) negative
LEBER CONGENITAL AMAUROSIS, TYPE CEP290 (CEP290) negative

LEBER CONGENITAL AMAUROSIS, TYPE LCAS (LCA5) negative LEBER CONGENITAL AMAUROSIS, TYPE RDH12 (RDH12) negative LEIGH SYNDROME, FRENCH-CANADIAN TYPE (LRPPRC) negative LETHAL CONGENITAL CONTRACTURE SYNDROME 1 (GLE1) negative LEUKOENCEPHALOPATHY WITH VANISHING WHITE MATTER (EIF2B5) negative LEUKOENCEPHALOPATHY WITH VANISHING WHITE MATTER, EIF2B1-RELATED

(EIF2B1) negative LEUKOENCEPHALOPATHY WITH VANISHING WHITE MATTER, EIF2B2-RELATED

(EIF2B2) negative LEUKOENCEPHALOPATHY WITH VANISHING WHITE MATTER, EIF2B3-RELATED (EIF2B3) negative

LEUKOENCEPHALOPATHY WITH VANISHING WHITE MATTER, EIF2B4-RELATED (EIF2B4) negative

LIG4 SYNDROME (LIG4) negative

LIMB-GIRDLE MUSCULAR DYSTROPHY TYPE 8 (TRIM32) negative LIMB-GIRDLE MUSCULAR DYSTROPHY TYPE 28 (TRIM32) negative
LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2A (CAPN3) negative
LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2B (DYSF) negative
LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2C (SGCG) negative
LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2D (SGCA) negative
LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2E (SGCB) negative
LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2F (SGCD) negative
LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2F (SGCD) negative
LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2I (FKRP) negative
LIPOAMIDE DEHYDROGENASE DEFICIENCY (DIHYDROLIPOAMIDE DEHYDROGENASE
DEFICIENCY) (DID) negative

DEFICIENCY) (DLD) negative
LIPOID ADRENAL HYPERPLASIA (STAR) negative

LIPOPROTEIN LIPASE DEFICIENCY (LPL) negative
LONG CHAIN 3-HYDROXYACYL-COA DEHYDROGENASE DEFICIENCY (HADHA) negative LRAT-RELATED CONDITIONS (LRAT) negative

LUNG DISEASE, IMMUNODEFICIENCY, AND CHROMOSOME BREAKAGE SYNDROME (LICS) (NSMCE3) negative LYSINURIC PROTEIN INTOLERANCE (SLC7A7) negative

MALONYL-COA DECARBOXYLASE DEFICIENCY (MLYCD) negative MAPLE SYRUP URINE DISEASE, TYPE 1A (BCKDHA) negative MAPLE SYRUP URINE DISEASE, TYPE 1B (BCKDHB) negative MAPLE SYRUP URINE DISEASE, TYPE 2 (DBT) negative MCKUSICK-KAUFMAN SYNDROME (MKKS) negative MECKEL SYNDROME 7/NEPHRONOPHTHISIS 3 (NPHP3) negative MECKEL-GRUBER SYNDROME, TYPE 1 (MKS1) negative
MECR-RELATED NEUROLOGIC DISORDER (MECR) negative
MEDIUM CHAIN ACYL-CoA DEHYDROGENASE DEFICIENCY (ACADM) negative MEDNIK SYNDROME (AP151) negative
MEGALENCEPHALIC LEUKOENCEPHALOPATHY WITH SUBCORTICAL CYSTS (MLC1) negative MEROSIN-DEFICIENT MUSCULAR DYSTROPHY (LAMA2) negative METABOLIC ENCEPHALOPATHY AND ARRHYTHMIAS, TANGO2-RELATED (TANGO2) negative METACHROMATIC LEUKODYSTROPHY, ARSA-RELATED (ARSA) negative METACHROMATIC LEUKODYSTROPHY, PSAP-RELATED (PSAP) negative METHYLMALONIC ACIDEMIA AND HOMOCYSTINURIA TYPE CBLF (LMBRD1) negative METHYLMALONIC ACIDEMIA, MCEE-RELATED (MCEE) negative METHYLMALONIC ACIDURIA, MICES-RELATED (MICE) negative METHYLMALONIC ACIDURIA AND HOMOCYSTINURIA, TYPE CBLC (MMACHC) negative METHYLMALONIC ACIDURIA AND HOMOCYSTINURIA, TYPE CbID (MMADHC) negative METHYLMALONIC ACIDURIA, MMAA-RELATED (MMAA) negative METHYLMALONIC ACIDURIA, MMAB-RELATED (MMAB) negative

METHYLMALONIC ACIDURIA, TYPE MUT(0) (MUT) negative
MEVALONIC KINASE DEFICIENCY (MVK) negative
MICROCEPHALIC OSTEODYSPLASTIC PRIMORDIAL DWARFISM TYPE II (PCNT) negative MICROPHTHALMIA / ANOPHTHALMIA, VSX2-RELATED (VSX2) negative

MITOCHONDRIAL COMPLEX 1 DEFICIENCY, ACAD9-RELATED (ACAD9) negative MITOCHONDRIAL COMPLEX 1 DEFICIENCY, NDUFAF5-RELATED (NDUFAF5) negative MITOCHONDRIAL COMPLEX 1 DEFICIENCY, NDUFS6-RELATED (NDUFS6) negative MITOCHONDRIAL COMPLEX I DEFICIENCY, NUCLEAR TYPE 1 (NDUFS4) negative MITOCHONDRIAL COMPLEX I DEFICIENCY, NUCLEAR TYPE 10 (NDUFAF2) negative MITOCHONDRIAL COMPLEX I DEFICIENCY, NUCLEAR TYPE 17 (NDUFAF6) negative MITOCHONDRIAL COMPLEX I DEFICIENCY, NUCLEAR TYPE 19 (FOXRED1) negative MITOCHONDRIAL COMPLEX I DEFICIENCY, NUCLEAR TYPE 3 (NDUFST) negative MITOCHONDRIAL COMPLEX I DEFICIENCY, NUCLEAR TYPE 4 (NDUFV1) negative MITOCHONDRIAL COMPLEX IV DEFICIENCY, NUCLEAR TYPE 2, SCO2-RELATED

(SCO2) negative
MITOCHONDRIAL COMPLEX IV DEFICIENCY, NUCLEAR TYPE 6 (COX15) negative

MITOCHONDRIAL DNA DEPLETION SYNDROME 2 (TK2) negative MITOCHONDRIAL DNA DEPLETION SYNDROME 3 (DGUOK) negative MITOCHONDRIAL MYOPATHY AND SIDEROBLASTIC ANEMIA (MLASA1) (PUS1) negative MITOCHONDRIAL TRIFUNCTIONAL PROTEIN DEFICIENCY, HADHB-RELATED (HADHB) negative MOLYBDENUM COFACTOR DEFICIENCY TYPE B (MOCS2) negative

MOLYBDENUM COFACTOR DEFICIENCY, TYPE A (MOCS1) negative

MUCOLIPIDOSIS II/III A (GNPTAB) negative MUCOLIPIDOSIS III GAMMA (GNPTG) negative MUCOLIPIDOSIS, TYPE IV (MCOLN1) negative

MUCOPOLYSACCHARIDOSIS, TYPE I (HURLER SYNDROME) (IDUA) negative MUCOPOLYSACCHARIDOSIS, TYPE III A (SANFILIPPO A) (SGSH) negative MUCOPOLYSACCHARIDOSIS, TYPE III B (SANFILIPPO B) (NAGLU) negative MUCOPOLYSACCHARIDOSIS, TYPE III B (SANFILIPPO B) (NAGLU) negative MUCOPOLYSACCHARIDOSIS, TYPE III C (SANFILIPPO C) (HGSNAT) negative MUCOPOLYSACCHARIDOSIS, TYPE III D (SANFILIPPO D) (GNS) negative MUCOPOLYSACCHARIDOSIS, TYPE IV A (MORQUIO SYNDROME) (GALNS) negative MUCOPOLYSACCHARIDOSIS, TYPE IV B/GM1 GANGLIOSIDOSIS (GLB1) negative

MUCOPOLYSACCHARIDOSIS, TYPE IX (HYAL1) negative
MUCOPOLYSACCHARIDOSIS, TYPE VI (MAROTEAUX-LAMY) (ARSB) negative
MUCOPOLYSACCHARIDOSIS, TYPE VII (GUSB) negative

MULIBREY NANISM (TRIM37) negative

MULTIPLE PTERYGIUM SYNDROME, CHRNG-RELATED/ESCOBAR SYNDROME (CHRNG) negative

MULTIPLE SULFATASE DEFICIENCY (SUMF1) negative MUSCLE-EYE-BRAIN DISEASE, POMGNT1-RELATED (POMGNT1) negative MUSCULAR DYSTROPHY-DYSTROGLYCANOPATHY (RXYLT1) negative MUSK-RELATED CONGENITAL MYASTHENIC SYNDROME (MUSK) negative MYONEUROGASTROINTESTINAL ENCEPHALOPATHY (MNGIE) (TYMP) negative MYOTONIA CONGENITA (CLCN1) negative

N-ACETYLGLUTAMATE SYNTHASE DEFICIENCY (NAGS) negative NEMALINE MYOPATHY, NEB-RELATED (NEB) negative NEPHRONOPHTHISIS 1 (NPHP1) negative NEURONAL CEROID LIPOFUSCINOSIS, CLN5-RELATED (CLN5) negative NEURONAL CEROID LIPOFUSCINOSIS, CLN6-RELATED (CLN6) negative NEURONAL CEROID LIPOFUSCINOSIS, CLN8-RELATED (CLN8) negative NEURONAL CEROID LIPOFUSCINOSIS, MFSD8-RELATED (MFSD8) negative NEURONAL CEROID LIPOFUSCINOSIS, PPT1-RELATED (PPT1) negative NEURONAL CEROID LIPOFUSCINOSIS, TPP1-RELATED (TPP1) negative NGLY1-CONGENITAL DISORDER OF GLYCOSYLATION (NGLY1) negative NGLY1-CONGENTIAL DISORDER OF GLYCOSYLATION NIEMANN-PICK DISEASE, TYPE C1 / D (NPC1) negative NIEMANN-PICK DISEASE, TYPE C2 (NPC2) negative NIEMANN-PICK DISEASE, TYPES A / B (SMPD1) negative NIJMEGEN BREAKAGE SYNDROME (NBN) negative NON-SYNDROMIC HEARING LOSS, GJB2-RELATED (GJB2) negative NON-SYNDROMIC HEARING LOSS, MYO15A-RELATED (MYO15A) negative NONSYNDROMIC HEARING LOSS, OTOA-RELATED (OTOA) negative NONSYNDROMIC HEARING LOSS, OTOF-RELATED (OTOF) negative NONSYNDROMIC HEARING LOSS, PJVK-RELATED (PJVK) negative NONSYNDROMIC HEARING LOSS, SYNE4-RELATED (SYNE4) negative NONSYNDROMIC HEARING LOSS, TMC1-RELATED (TMC1) negative NONSYNDROMIC HEARING LOSS, TMPRSS3-RELATED (TMPRSS3) negative NONSYNDROMIC INTELLECTUAL DISABILITY (CC2D1A) negative NORMOPHOSPHATEMIC TUMORAL CALCINOSIS (SAMD9) negative

OCULOCUTANEOUS ALBINISM TYPE IV (SLC45A2) negative OCULOCUTANEOUS ALBINISM TYPE, III (TYRP1) negative OCULOCUTANEOUS ALBINISM, OCA2-RELATED (OCA2) negative OCULOCUTANEOUS ALBINISM, TYPES 1A AND 1B (TYR) negative ODONTO-ONYCHO-DERMAL DYSPLASIA / SCHOPF-SCHULZ-PASSARGE SYNDROME (WNT10A) negative OMENN SYNDROME, RAG2-RELATED (RAG2) negative
ORNITHINE AMINOTRANSFERASE DEFICIENCY (OAT) negative OSTEOGENESIS IMPERFECTA TYPE VII (CRTAP) negative OSTEOGENESIS IMPERFECTA TYPE VIII (P3H1) negative OSTEOGENESIS IMPERFECTA TYPE XI (FKBP10) negative OSTEOGENESIS IMPERFECTA TYPE XII (BMP1) negative OSTEOPETROSIS, INFANTILE MALIGNANT, TCIRG1-RELATED (TCIRG1) negative OSTEOPETROSIS, OSTM1-RELATED (OSTM1) negative

PANTOTHENATE KINASE-ASSOCIATED NEURODEGENERATION (PANK2) negative PAPILLON LEFÈVRE SYNDROME (CTSC) negative PARKINSON DISEASE 15 (FBXO7) negative PENDRED SYNDROME (SLC26A4) negative
PERLMAN SYNDROME (DIS3L2) negative
PGM3-CONGENITAL DISORDER OF GLYCOSYLATION (PGM3) negative PHENYLKETONURIA (PAH) negative
PIGN-CONGENITAL DISORDER OF GLYCOSYLATION (PIGN) negative
PITUITARY HORMONE DEFICIENCY, COMBINED 3 (LHX3) negative



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POLG-RELATED DISORDERS (POLG) negative
POLYCYSTIC KIDNEY DISEASE, AUTOSOMAL RECESSIVE (PKHD1) negative PONTOCEREBELLAR HYPOPLASIA, EXOSC3-RELATED (EXOSC3) negative

PONTOCEREBELLAR HYPOPLASIA, RARS2-RELATED (RARS2) negative PONTOCEREBELLAR HYPOPLASIA, TSEN2-RELATED (TSEN2) negative PONTOCEREBELLAR HYPOPLASIA, TSEN54-RELATED (TSEN54) negative

PONTOCEREBELLAR HYPOPLASIA, TYPE 1A (VRK1) negative PONTOCEREBELLAR HYPOPLASIA, TYPE 2D (SEPSECS) negative PONTOCEREBELLAR HYPOPLASIA, VPS53-RELATED (VPS53) negative

PRIMARY CILIARY DYSKINESIA, CCDC103-RELATED (CCDC103) negative

PRIMARY CILIARY DYSKINESIA, CCDC39-RELATED (CCDC39) negative PRIMARY CILIARY DYSKINESIA, DNAH11-RELATED (DNAH11) negative

PRIMARY CILIARY DYSKINESIA, DNAH5-RELATED (DNAH5) negative

PRIMARY CILIARY DYSKINESIA, DNAI1-RELATED (DNAI1) negative PRIMARY CILIARY DYSKINESIA, DNAI2-RELATED (DNAI2) negative PRIMARY CONGENITAL GLAUCOMA/PETERS ANOMALY (CYP1B1) negative

PRIMARY CONGENITAL GLAUCUMA/PETERS ANOMALY (CYPIBI) negative PRIMARY HYPEROXALURIA, TYPE 1 (AGXT) negative PRIMARY HYPEROXALURIA, TYPE 3 (HOGA1) negative PRIMARY HYPEROXALURIA, TYPE 3 (HOGA1) negative PRIMARY MICROCEPHALY 1, AUTOSOMAL RECESSIVE (MCPH1) negative

PROGRESSIVE EARLY-ONSET ENCEPAHLOPATHY WITH BRAIN ATROPHY AND THIN CORPUS CALLOSUM (TBCD) negative PROGRESSIVE FAMILIAL INTRAHEPATIC CHOLESTASIS, ABCB4-RELATED (ABCB4) negative PROGRESSIVE FAMILIAL INTRAHEPATIC CHOLESTASIS, TYPE 1 (PFIC1) (AF881) negative PROGRESSIVE FAMILIAL INTRAHEPATIC CHOLESTASIS, TYPE 2 (ABCB11) negative PROGRESSIVE FAMILIAL INTRAHEPATIC CHOLESTASIS, TYPE 4 (PFIC4) (TJP2) negative

PROGRESSIVE PSEUDORHEUMATOID DYSPLASIA (CCN6) negative PROLIDASE DEFICIENCY (PEPD) negative PROPIONIC ACIDEMIA, PCCA-RELATED (PCCA) negative

PROPIONIC ACIDEMIA, PCCB-RELATED (PCCB) negative

PSEUDOXANTHOMA ELASTICUM (ABCC6) negative
PTERIN-4 ALPHA-CARBINOLAMINE DEHYDRATASE (PCD) DEFICIENCY (PCBD1) negative

PTERIN-4 ALPHA-CARBINOLAMINE DEHYDRATASE (PCD) DEFICIENCY (PCBD1 PYCNODYSOSTOSIS (CTSK) negative PYRIDOXAL 5'-PHOSPHATE-DEPENDENT EPILEPSY (PNPO) negative PYRIDOXINE-DEPENDENT EPILEPSY (ALDH7A1) negative PYRUVATE CARBOXYLASE DEFICIENCY (PC) negative PYRUVATE DEHYDROGENASE DEFICIENCY, PDHB-RELATED (PDHB) negative

REFSUM DISEASE, PHYH-RELATED (PHYH) negative
RENAL TUBULAR ACIDOSIS AND DEAFNESS, ATP6V1B1-RELATED (ATP6V1B1) negative
RENAL TUBULAR ACIDOSIS, PROXIMAL, WITH OCULAR ABNORMALITIES AND MENTAL
RETARDATION (SLC4A4) negative
RETINITIS PIGMENTOSA 25 (EYS) negative

RETINITIS PIGMENTOSA 26 (CERKL) negative RETINITIS PIGMENTOSA 28 (FAM161A) negative RETINITIS PIGMENTOSA 36 (PRCD) negative

RETINITIS PIGMENTOSA 59 (DHDDS) negative

RETINITIS PIGMENTOSA 62 (MAK) negative RHIZOMELIC CHONDRODYSPLASIA PUNCTATA, TYPE 1 (PEX7) negative RHIZOMELIC CHONDRODYSPLASIA PUNCTATA, TYPE 2 (GNPAT) negative

RHIZOMELIC CHONDRODYSPLASIA PUNCTATA, TYPE 3 (AGPS) negative RLBP1-RELATED RETINOPATHY (RLBP1) negative ROBERTS SYNDROME (ESCO2) negative

RYR1-RELATED CONDITIONS (RYR1) negative

SALLA DISEASE (SLC17A5) negative

SANDHOFF DISEASE (HEXB) negative SCHIMKE IMMUNOOSSEOUS DYSPLASIA (SMARCAL1) negative SCHINDLER DISEASE (NAGA) negative

SEGAWA SYNDROME, TH-RELATED (TH) negative
SENIOR-LOKEN SYNDROME 4/NEPHRONOPHTHISIS 4 (NPHP4) negative
SEPIAPTERIN REDUCTASE DEFICIENCY (SPR) negative
SEVERE COMBINED IMMUNODEFICIENCY (SCID), CD3D-RELATED (CD3D) negative

SEVERE COMBINED IMMUNODEFICIENCY (SCID), CD3E-RELATED (CD3E) negative SEVERE COMBINED IMMUNODEFICIENCY (SCID), FOXN1-RELATED (FOXN1) negative SEVERE COMBINED IMMUNODEFICIENCY (SCID), IKBKB-RELATED (IKBKB) negative

SEVERE COMBINED IMMUNODEFICIENCY (SCID), IL7R-RELATED (IL7R) negative SEVERE COMBINED IMMUNODEFICIENCY (SCID), JAK3-RELATED (JAK3) negative SEVERE COMBINED IMMUNODEFICIENCY (SCID), PTPRC-RELATED (PTPRC) negative

SEVERE COMBINED IMMUNODEFICIENCY (SCID), RAG1-RELATED (RAG1) negative

SEVERE COMBINED IMMUNODEFICIENCY, ADA-Related (ADA) negative SEVERE COMBINED IMMUNODEFICIENCY, TYPE ATHABASKAN (DCLRE1C) negative

SHORT-RIB THORACIC DYSPLASIA 3 WITH OR WITHOUT POLYDACTYLY

(DYNC2H1) negative

SHWACHMAN-DIAMOND SYNDROME, SBDS-RELATED (SBDS) negative

SIALIDOSIS (NEU1) negative SJÖGREN-LARSSON SYNDROME (ALDH3A2) negative SMITH-LEMLI-OPITZ SYNDROME (DHCR7) negativ

SPASTIC PARAPLEGIA, TYPE 15 (ZFYVE26) negative

SPASTIC TETRAPLEGIA, THIN CORPUS CALLOSUM, AND PROGRESSIVE MICROCEPHALY (SPATCCM) (SLC1A4) negative

SPG11-RELATED CONDITIONS (SPG11) negative

SPINAL MUSCULAR ATROPHY (SMN1) negative SMN1: Two copies; g.27134T>G: absent; the absence of the g.27134T>G variant decreases the chance to be a silent (2+0) carrier.

SPINAL MUSCULAR ATROPHY WITH RESPIRATORY DISTRESS TYPE 1 (IGHMBP2) negative

SPINOCEREBELLAR ATAXIA, AUTOSOMAL RECESSIVE 10 (ANO10) negative SPINOCEREBELLAR ATAXIA, AUTOSOMAL RECESSIVE 12 (WWOX) negative SPONDYLOCOSTAL DYSOSTOSIS 1 (DLL3) negative

SPONDYLOTHORACIC DYSOSTOSIS, MESP2-Related (MESP2) negative

STEEL SYNDROME (COL27A1) negative STEROID-RESISTANT NEPHROTIC SYNDROME (NPHS2) negative

STUVE-WIEDEMANN SYNDROME (LIFR) negative

SURF1-RELATED CONDITIONS (SURF1) negative SURFACTANT DYSFUNCTION, ABCA3-RELATED (ABCA3) negative

TAY-SACHS DISEASE (HEXA) negative
TBCE-RELATED CONDITIONS (TBCE) negative

THIAMINE-RESPONSIVE MEGALOBLASTIC ANEMIA SYNDROME (SLC19A2) negative

THYROID DYSHORMONOGENESIS 1 (SLC5A5) negative THYROID DYSHORMONOGENESIS 2A (TPO) negative

THYROID DYSHORMONOGENESIS 3 (TG) negative

THYROID DYSHORMONOGENESIS 3 (16) negative THYROID DYSHORMONOGENESIS 6 (DUOX2) negative TRANSCOBALAMIN II DEFICIENCY (TCN2) negative TRICHOHEPATOENTERIC SYNDROME, SKIC2-RELATED (SKIC2) negative TRICHOHEPATOENTERIC SYNDROME, TTC37-RELATED (TTC37) negative

TRICHOTHIODYSTROPHY 1/XERODERMA PIGMENTOSUM, GROUP D (ERCC2) negative TRIMETHYLAMINURIA (FMO3) negative TRIPLE A SYNDROME (AAAS) negative

TSHR-RELATED CONDITIONS (TSHR) negative TYROSINEMIA TYPE III (HPD) negative TYROSINEMIA, TYPE 1 (FAH) negative TYROSINEMIA, TYPE 2 (TAT) negative

USHER SYNDROME, TYPE 1B (MYO7A) negative

USHER SYNDROME, TYPE 1C (USH1C) negative

USHER SYNDROME, TYPE 1D (CDH23) negative USHER SYNDROME, TYPE 1F (PCDH15) negative

USHER SYNDROME, TYPE 1J/DEAFNESS, AUTOSOMAL RECESSIVE, 48 (CIB2) negative

USHER SYNDROME, TYPE 2A (USH2A) negative USHER SYNDROME, TYPE 2C (ADGRV1) negative USHER SYNDROME, TYPE 3 (CLRN1) negative

VERY LONG-CHAIN ACYL-CoA DEHYDROGENASE DEFICIENCY (ACADVL) negative

VICI SYNDROME (EPG5) negative

VITAMIN D-DEPENDENT RICKETS, TYPE 1A (CYP27B1) negative VITAMIN D-RESISTANT RICKETS TYPE 2A (VDR) negative VLDLR-ASSOCIATED CEREBELLAR HYPOPLASIA (VLDLR) negative

WALKER-WARBURG SYNDROME, CRPPA-RELATED (CRPPA) negative

WALKER-WARBURG SYNDROME, FKTN-RELATED (FKTN) negative WALKER-WARBURG SYNDROME, LARGE1-RELATED (LARGE1) negative WALKER-WARBURG SYNDROME, POMT1-RELATED (POMT1) negative

WALKER-WARBURG SYNDROME, POMT2-RELATED (POMT2) see first page

WARSAW BREAKAGE SYNDROME (DDX11) negative WERNER SYNDROME (WRN) negative WILSON DISEASE (ATP7B) negative

WOLCOTT-RALLISON SYNDROME (EIF2AK3) negative

WOLMAN DISEASE (LIPA) negative WOODHOUSE-SAKATI SYNDROME (DCAF17) negative

XERODERMA PIGMENTOSUM, GROUP C (XPC) negative

XERODERMA PIGMENTOSUM VARIANT TYPE (POLH) negative XERODERMA PIGMENTOSUM, GROUP A (XPA) negative

ZELLWEGER SPECTRUM DISORDER, PEX13-RELATED (PEX13) negative ZELLWEGER SPECTRUM DISORDER, PEX16-RELATED (PEX16) negative ZELLWEGER SPECTRUM DISORDER, PEX5-RELATED (PEX5) negative ZELLWEGER SPECTRUM DISORDERS, PEX10-RELATED (PEX10) negative

ZELLWEGER SPECTRUM DISORDERS, PEX12-RELATED (PEX12) negative ZELLWEGER SPECTRUM DISORDERS, PEX1-RELATED (PEX1) negative ZELLWEGER SPECTRUM DISORDERS, PEX26-RELATED (PEX26) negative



Patient Name:

Test InformationOrdering Physician:

ho

Clinic Information:

Date Of Birth: Case File ID:

Report Date:

Z ZELLWEGER SPECTRUM DISORDERS, PEX2-RELATED (PEX2) negative ZELLWEGER SPECTRUM DISORDERS, PEX6-RELATED (PEX6) negative

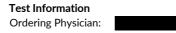


Patient	Information
D 11 1	N.I.

Patient Name:

Date Of Birth:

Case File ID:



Clinic Information:



Report Date:

Testing Methodology, Limitations, and Comments:

Next-generation sequencing (NGS)

Sequencing library prepared from genomic DNA isolated from a patient sample is enriched for targets of interest using standard hybridization capture protocols and PCR amplification (for targets specified below). NGS is then performed to achieve the standards of quality control metrics, including a minimum coverage of 99% of targeted regions at 20X sequencing depth. Sequencing data is aligned to human reference sequence, followed by deduplication, metric collection and variant calling (coding region +/- 20bp). Variants are then classified according to ACMGG/AMP standards of interpretation using publicly available databases including but not limited to ENSEMBL, HGMD Pro, ClinGen, ClinVar, 1000G, ESP and gnomAD. Variants predicted to be pathogenic or likely pathogenic for the specified diseases are reported. It should be noted that the data interpretation is based on our current understanding of the genes and variants at the time of reporting. Putative positive sequencing variants that do not meet internal quality standards or are within highly homologous regions are confirmed by Sanger sequencing or gene-specific long-range PCR as needed prior to reporting.

Copy Number Variant (CNV) analysis is limited to deletions involving two or more exons for all genes on the panel, in addition to specific known recurrent single-exon deletions. CNVs of small size may have reduced detection rate. This method does not detect gene inversions, single-exonic and sub-exonic deletions (unless otherwise specified), and duplications of all sizes (unless otherwise specified). Additionally, this method does not define the exact breakpoints of detected CNV events. Confirmation testing for copy number variation is performed by specific PCR, Multiplex Ligation-dependent Probe Amplification (MLPA), next generation sequencing, or other methodology.

This test may not detect certain variants due to local sequence characteristics, high/low genomic complexity, homologous sequence, or allele dropout (PCR-based assays). Variants within noncoding regions (promoter, 5'UTR, 3'UTR, deep intronic regions, unless otherwise specified), small deletions or insertions larger than 25bp, low-level mosaic variants, structural variants such as inversions, and/or balanced translocations may not be detected with this technology.

SPECIAL NOTES

For ABCC6, variants in exons 1-9 are not detected due to the presence of regions of high homology.

For CFTR, when the CFTR R117H variant is detected, reflex analysis of the polythymidine variations (5T, 7T and 9T) at the intron 9 branch/acceptor site of the CFTR gene will be performed.

For CYP21A2, targets were enriched using long-range PCR amplification, followed by next generation sequencing. Duplication analysis will only be performed and reported when c.955C>T (p.Q319*) is detected. Sequencing and CNV analysis may have reduced sensitivity, if variants result from complex rearrangements, in trans with a gene deletion, or CYP21A2 gene duplication on one chromosome and deletion on the other chromosome. This analysis cannot detect sequencing variants located on the CYP21A2 duplicated copy.

For DDX11, only NM_030653.3:c.1763 - 1G > C variant will be analyzed and reported.

For GJB2, CNV analysis of upstream deletions of GJB6-D13S1830 (309kb deletion) and GJB6-D13S1854 (232kb deletion) is included.

For HBA1/HBA2, CNV analysis is offered to detect common deletions of -alpha3.7, -alpha4.2, --MED, --SEA, --FIL, --THAI, --alpha20.5, and/or HS-40.

For OTOA, variants in exons 20 - 28 are not analyzed due to high sequence homology.

For RPGRIP1L, variants in exon 23 are not detected due to assay limitation.

For SAMD9, only p.K1495E variant will be analyzed and reported.

Friedreich Ataxia (FXN)

The GAA repeat region of the FXN gene is assessed by trinucleotide PCR assay and capillary electrophoresis. Variances of +/-1 repeat for normal alleles and up to +/-3 repeats for premutation alleles may occur. For fully penetrant expanded alleles, the precise repeat size cannot be determined, therefore the approximate allele size is reported. Sequencing and copy number variants are analyzed by next-generation sequencing analysis.

Friedreich Ataxia Repeat Categories

Categories	GAA Repeat Sizes
Normal	<34
Premutation	34 - 65
Full	>65



Patient Information Patient Name:	Test Information Ordering Physician:
Date Of Birth:	Clinic Information:
Case File ID:	Report Date:



Spinal Muscular Atrophy (SMN1)

The total combined copy number of SMN1 and SMN2 exon 7 is quantified based on NGS read depth. The ratio of SMN1 to SMN2 is calculated based on the read depth of a single nucleotide that distinguishes these two genes in exon 7. In addition to copy number analysis, testing for the presence or absence of a single nucleotide polymorphism (g.27134T>G in intron 7 of SMN1) associated with the presence of a SMN1 duplication allele is performed using NGS.

Ethnicity	Two SMN1 copies carrier risk before g.27134T>G testing	Carrier risk after g.27134T	Carrier risk after g.27134T>G testing	
		g.27134T>G ABSENT	g.27134T>G PRESENT	
Caucasian	1 in 632	1 in 769	1 in 29	
Ashkenazi Jewish	1 in 350	1 in 580	LIKELY CARRIER	
Asian	1 in 628	1 in 702	LIKELY CARRIER	
African-American	1 in 121	1 in 396	1 in 34	
Hispanic	1 in 1061	1 in 1762	1 in 140	

Variant Classification

Only pathogenic or likely pathogenic variants are reported. Other variants including benign variants, likely benign variants, variants of uncertain significance, or inconclusive variants identified during this analysis may be reported in certain circumstances. Our laboratory's variant classification criteria are based on the ACMG and internal guidelines and our current understanding of the specific genes. This interpretation may change over time as more information about a gene and/or variant becomes available. Natera and its lab partner(s) may reclassify variants at certain intervals but may not release updated reports without a specific request made to Natera by the ordering provider. Natera may disclose incidental findings if deemed clinically pertinent to the test performed.

Negative Results

A negative carrier screening result reduces the risk for a patient to be a carrier of a specific disease but does not completely rule out carrier status. Please visit https://www.natera.com/panel-option/h-all/ for a table of carrier rates, detection rates, residual risks and promised variants/exons per gene. Carrier rates before and after testing vary by ethnicity and assume a negative family history for each disease screened and the absence of clinical symptoms in the patient. Any patient with a family history for a specific genetic disease will have a higher carrier risk prior to testing and, if the disease-causing mutation in their family is not included on the test, their carrier risk would remain unchanged. Genetic counseling is recommended for patients with a family history of genetic disease so that risk figures based on actual family history can be determined and discussed along with potential implications for reproduction. Horizon carrier screening has been developed to identify the reproductive risks for monogenic inherited conditions. Even when one or both members of a couple screen negative for pathogenic variants in a specific gene, the disease risk for their offspring is not zero. There is still a low risk for the condition in their offspring due to a number of different mechanisms that are not detected by Horizon including, but not limited to, pathogenic variant(s) in the tested gene or in a different gene not included on Horizon, pathogenic variant(s) in an upstream regulator, uniparental disomy, de novo mutation(s), or digenic or polygenic inheritance.

Additional Comments

These analyses generally provide highly accurate information regarding the patient's carrier status. Despite this high level of accuracy, it should be kept in mind that there are many potential sources of diagnostic error, including misidentification of samples, polymorphisms, or other rare genetic variants that interfere with analysis. Families should understand that rare diagnostic errors may occur for these reasons.





7703,DONOR 🛦

7703,DONOR (





Specimen: Requisition Lab Reference ID Report Status: FINAL / SEE REPORT Collected: 07/05/2024 10:00 Received: 07/06/2024 14:55 Reported: 07/15/2024 17:55



CBC (includes Differential and Platelets)	(FINAL) Value		
Analyte		Reference Range: 3.8-10.8 Thous/uL	FINAL
White Blood Cell Count		Reference Range: 4.20-5.80 Mill/uL	(FINAL)
Red Blood Cell Count		Reference Range: 13.2-17.1 g/dL	FINAL
HEMOGLOBIN		Reference Range: 38.5-50.0 %	FINAL
Hematocrit	3.70.5	Reference Range: 80.0-100.0 fL	FINAL
MCV		Reference Range: 27.0-33.0 pg	FINAL
мсн	28.4	Reference Range: 32.0-36.0 g/dL	(FINAL)
▲ MCHC	30.8 L	Reference Range: 11.0-15.0 %	FINAL
RDW	12.4	Reference Range: 140-400 Thous/uL	FINAL
PLATELET COUNT	236	Reference Range: 7.5-12.5 fl	FINAL
MPV	10.2	Reference Range: 1500-7800 cells/uL	FINAL
Absolute Neutrophils	2246	Reference Range: 850-3900 cells/uL	FINAL
Absolute Lymphocytes	1182	Reference Range: 200-950 cells/uL	FINAL
Absolute Monocytes	371	Reference Range: 15-500 cells/uL	FINAL
Absolute Eosinophils	90	Reference Range: 0-200 cells/uL	(FINAL.)
Absolute Basophils	12	%	FINAL
Neutrophils	57.6	%	FINAL
Lymphocytes	30.3	%	FINAL
Monocytes	9.50	%	FINAL
Eosinophils	2.3	%	FINAL
Basophils	0.30	% Reference Range: 0 /100 WBC	FINAL
Nucleated RBC	0.00	Natalanda Handara 11.22	
			Lab: AMD
Hemoglobinopathy Evaluation (FINAL)	Value		
Analyte	yaluc		FINAL
Hemoglobinopathy Evaluation	5.24	Reference Range: 4.20-5.80 Mill/uL	FINAL
Red Blood Cell Count	14.9		FINA
HEMOGLOBIN	14.3	,	FINA
Hematocrit	48.	3 Reference Range: 38.5-50.0 %	FINA
Hematocrit	92.	200010001	FINA
MCV	28.	07.0.22.0.22	FINA
MCH 7703,DONOR (1/3	4 Individue IIII.	7/24/

	12.4	Reference Range: 11.0-15.0 %	FINAL
RDW	12.7		FINAL
Hemoglobinopathy Evaluation		00.0.0	(FINAL)
Hemoglobin A	97.2	Reference Range: >96.0 %	(FINAL)
Hemoglobin F	0.0	Reference Range: <2.0 %	
Hemoglobin A2 (Quant)	2.8	Reference Range: 2.0-3.2 %	(FINAL)
Interpretation			(FEVAL)

Interpretation

NORMAL PATTERN

There is a normal pattern of hemoglobins and normal levels of Hb A2 and Hb F are present. No variant hemoglobins are observed. This is consistent with A/A phenotype. If iron deficiency coexists with a mild/silent beta thalassemia trait Hb A2 may be in the normal range. Rare variant hemoglobins have no separation from hemoglobin A by capillary zone electrophoresis (CZE) or high-performance liquid chromatography (HPLC). If clinically indicated, Thalassemia and Hemoglobinopathy Comprehensive (TC 17365) should be considered.

Chromosome Analysis, Blood

Order ID:

Specimen Type:

Blood

Clinical Indication:

Donor screening

NORMAL MALE KARYOTYPE

Chromosome analysis revealed normal G-band patterns within the limits of standard cytogenetic analysis.

Please expect the results of any other concurrent study in a separate report.

NOMENCLATURE:

46, XY

ASSAY INFORMATION:

Method:

G-Band (Digital Analysis:

MetaSystems/Ikaros)

Cells Counted:

20

Band Level:

550

Cells Analyzed: Cells Karyotyped: 5 3

This test does not address genetic disorders that cannot be detected by standard cytogenetic methods or rare events such as low level mosaicism or subtle rearrangements.

Haiying Meng, M.D., Ph.D., FACMG, Technical Director, Cytogenetics and Genomics, 703-802-7156

Electronic Signature:

7/15/2024 5:13 PM

For additional information, please refer to http://education.questdiagnostics.com/faq/chromsblood (This link is being provided for informational/ educational purposes only).

Performing Sites

AMD Quest Diagnostics Nichols Institute, 14225 Newbrook Drive, Chantilly, VA 20151 Laboratory Director: Patrick W Mason, MD PhD

Key

Priority Out of Range A Out of Range PEND Pending Result PRE Preliminary Result FINAL Final Result RE Reissued Result

(FINAL)

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7703,DONOR





Patient Information:
7703, Donor
DOB:
Sex: M
MR#: 7703
Patient#:



Specimen Type: DNA Collected: Not provided Received Date: Mar 13,2025 Authorized Date: Mar 17,2025 Physician:
Wieloch, Shannon
GC: Wieloch, Shannon
Fairfax Cryobank
3015 Williams Drive #110
Fairfax, VA 22031
Phone: 7038763869

Fax:

Laboratory:

Fulgent Therapeutics LLC CAP#: 8042697 CLIA#: 05D2043189 Laboratory Director: Dr. Amar Jariwala Report Date: Apr 03,2025

Final Report

TEST PERFORMED

Single Gene - Gene

(1 Gene Panel: IL36RN; gene sequencing with deletion and duplication analysis)

RESULTS:

No clinically significant sequence or copy-number variants were identified in the submitted specimen.

A negative result does not rule out the possibility of a genetic predisposition nor does it rule out any pathogenic mutations of the sort not queried by this test or in areas not reliably assessed by this test.

INTERPRETATION:

Notes and Recommendations:

- As requested, this report only includes variants classified as Pathogenic, Likely Pathogenic, or Risk Allele at the time of analysis. If detected, this report does not include variants classified as of uncertain significance.
- Gene specific notes and limitations may be present. See below.
- These results should be interpreted in the context of this individual's clinical findings, biochemical profile, and family history.
- Genetic counseling is recommended. Available genetic counselors and additional resources can be found at the National Society of Genetic Counselors (NSGC; https://www.nsgc.org)
- Guide to Interpreting Genomic Reports: A Genomics Toolkit (CSER Consortium; February 2017)
 (https://www.genome.gov/For-Health-Professionals/Provider-Genomics-Education-Resources#hep)

GENES TESTED:

Single Gene - Gene

1 genes tested (100.00% at >20x).

IL36RN

Gene Specific Notes and Limitations

No gene specific limitations apply to the genes on the tested panel.

Patient: 7703, Donor; Sex: M; DOB: ; MR#: 7703

PAGE 1 of 3





METHODS:

Genomic DNA was isolated from the submitted specimen indicated above (if cellular material was submitted). DNA was barcoded, and enriched for the coding exons of targeted genes using hybrid capture technology. Prepared DNA libraries were then sequenced using a Next Generation Sequencing technology. Following alignment to the human genome reference sequence (assembly GRCh37), variants were detected in regions of at least 10x coverage. For this specimen, 100.00% and 100.00% of coding regions and splicing junctions of genes listed had been sequenced with coverage of at least 10x and 20x, respectively, by NGS or by Sanger sequencing. The remaining regions did not have 10x coverage, and were not evaluated. Variants were interpreted manually using locus specific databases, literature searches, and other molecular biological principles. To minimize false positive results, any variants that do not meet internal quality standards are confirmed by Sanger sequencing. Variants classified as pathogenic, likely pathogenic, or risk allele which are located in the coding regions and nearby intronic regions (+/- 20bp) of the genes listed above are reported. Variants outside these intervals may be reported but are typically not guaranteed. When a single pathogenic or likely pathogenic variant is identified in a clinically relevant gene with autosomal recessive inheritance, the laboratory will attempt to ensure 100% coverage of coding sequences either through NGS or Sanger sequencing technologies ("fill-in"). All genes listed were evaluated for large deletions and/or duplications. However, single exon deletions or duplications will not be detected in this assay, nor will copy number alterations in regions of genes with significant pseudogenes. Putative deletions or duplications identified by NGS are confirmed by an orthogonal method (qPCR or MLPA), unless exceeding an internally specified and validated quality score, beyond which deletions and duplications are considered real without further confirmation. New York patients: diagnostic findings are confirmed by Sanger, MLPA, or qPCR; exception SNV variants in genes for which confirmation of NGS results has been performed >=10 times may not be confirmed if identified with high quality by NGS. Bioinformatics: The FPLMv2.0 pipeline was used to analyze this specimen.

LIMITATIONS:

These test results and variant interpretation are based on the proper identification of the submitted specimen, accuracy of any stated familial relationships, and use of the correct human reference sequences at the queried loci. In very rare instances, errors may result due to mix-up or co-mingling of specimens. Positive results do not imply that there are no other contributors, genetic or otherwise, to this individual's phenotype, and negative results do not rule out a genetic cause for the indication for testing. Official gene names change over time. Fulgent uses the most up to date gene names based on HUGO Gene Nomenclature Committee (https://www.genenames.org) recommendations. If the gene name on report does not match that of ordered gene, please contact the laboratory and details can be provided. Result interpretation is based on the available clinical and family history information for this individual, collected published information, and Alamut annotation available at the time of reporting. This assay is designed and validated for detection of germline variants only. It is not designed or validated for the detection of low-level mosaicism or somatic mutations. This assay will not detect certain types of genomic aberrations such as translocations, inversions, or repeat expansions (eq. trinucleotide or hexanucleotide repeat expansion). DNA alterations in regulatory regions or deep intronic regions (greater than 20bp from an exon) may not be detected by this test. Unless otherwise indicated, no additional assays have been performed to evaluate genetic changes in this specimen. There are technical limitations on the ability of DNA sequencing to detect small insertions and deletions. Our laboratory uses a sensitive detection algorithm for copy number variants, however these types of alterations are not detected as reliably as single nucleotide variants. Rarely, due to systematic chemical, computational, or human error, DNA variants may be missed. Although next generation sequencing technologies and our bioinformatics analysis significantly reduce the confounding contribution of pseudogene sequences or other highly-homologous sequences, sometimes these may still interfere with the technical ability of the assay to identify pathogenic alterations in both sequencing and deletion/duplication analyses. Deletion/duplication analysis can identify alterations of genomic regions which are two or more contiguous exons in size; single exon deletions or duplications may occasionally be identified, but are not routinely detected by this test. When novel DNA duplications are identified, it is not possible to discern the genomic location or orientation of the duplicated segment, hence the effect of the duplication cannot be predicted. Where deletions are detected, it is not always possible to determine whether the predicted product will remain in-frame or not. Unless otherwise indicated, deletion/duplication analysis has not been performed in regions that have been sequenced by Sanger.

SIGNATURE:

Yan Meng, Ph.D., CGMB, FACMG Laboratory Director, Fulgent

Dr. Harry Gao, DABMG, FACMG on 4/3/2025

Laboratory Director, Fulgent

Patient: 7703, Donor; Sex: M; DOB: ; MR#: 7703





DISCLAIMER:

This test was developed, performed and its performance characteristics determined by **Fulgent Therapeutics LLC**. It has not been cleared or approved by the FDA. The laboratory is regulated under CLIA as qualified to perform high-complexity testing. This test is used for clinical purposes. It should not be regarded as investigational or for research. Since genetic variation, as well as systematic and technical factors, can affect the accuracy of testing, the results of testing should always be interpreted in the context of clinical and familial data. For assistance with interpretation of these results, healthcare professionals may contact us directly at (626) 350-0537 or info@fulgentgenetics.com. It is recommended that patients receive appropriate genetic counseling to explain the implications of the test result, including its residual risks, uncertainties and reproductive or medical options.

Patient: 7703, Donor; Sex: M;
DOB: RM#: 7703
PAGE 3 of 3





Patient Information: 7703, Donor DOB:

Sex: M MR#: 7703 Patient#: Partner Information: Not Tested

Accession: N/A Wieloch, Shannon GC: Wieloch, Shannon Fairfax Cryobank 3015 Williams Drive #110 Fairfax, VA 22031

Phone:

Physician:

Laboratory:

Fulgent Therapeutics LLC CAP#: 8042697 CLIA#: 05D2043189 Laboratory Director: Dr. Amar Jariwala

Report Date: May 20,2025

Accession:

Specimen Type: DNA Collected: Not Provided

FINAL RESULTS

No carrier mutations identified

TEST PERFORMED

Single Gene Carrier Screening: TCTN1

(1 Gene Panel: *TCTN1*; gene sequencing with deletion and duplication analysis)

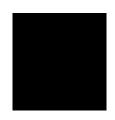
INTERPRETATION:

Notes and Recommendations:

- No carrier mutations were identified in the submitted specimen. A negative result does not rule out the possibility of a genetic
 predisposition nor does it rule out any pathogenic mutations in areas not assessed by this test or in regions that were covered
 at a level too low to reliably assess. Also, it does not rule out mutations that are of the sort not queried by this test; see
 Methods and Limitations for more information. A negative result reduces, but does not eliminate, the chance to be a carrier for
 any condition included in this screen. Please see the supplemental table for details.
- This carrier screening test does not screen for all possible genetic conditions, nor for all possible mutations in every gene tested. This report does not include variants of uncertain significance; only variants classified as pathogenic or likely pathogenic at the time of testing, and considered relevant for reproductive carrier screening, are reported. Please see the gene specific notes for details. Please note that the classification of variants can change over time.
- Patients may wish to discuss any carrier results with blood relatives, as there is an increased chance that they are also carriers. These results should be interpreted in the context of this individual's clinical findings, biochemical profile, and family history.
- Gene specific notes and limitations may be present. See below.
- Genetic counseling is recommended. Available genetic counselors and additional resources can be found at the National Society of Genetic Counselors (NSGC; https://www.nsgc.org)

Patient: 7703, Donor; Sex: M;
DOB: RM#: 7703
PAGE 1 of 4





GENES TESTED:

Custom Beacon Carrier Screening Panel - Gene

This analysis was run using the Custom Beacon Carrier Screening Panel gene list. 1 genes were tested with 100.0% of targets sequenced at >20x coverage. For more gene-specific information and assistance with residual risk calculation, see the SUPPLEMENTAL TABLE.

TCTN1

METHODS:

Genomic DNA was isolated from the submitted specimen indicated above (if cellular material was submitted). DNA was barcoded, and enriched for the coding exons of targeted genes using hybrid capture technology. Prepared DNA libraries were then sequenced using a Next Generation Sequencing technology. Following alignment to the human genome reference sequence (assembly GRCh37), variants were detected in regions of at least 10x coverage. For this specimen, 100.00% and 100.00% of coding regions and splicing junctions of genes listed had been sequenced with coverage of at least 10x and 20x, respectively, by NGS or by Sanger sequencing. The remaining regions did not have 10x coverage, and were not evaluated. Variants were interpreted manually using locus specific databases, literature searches, and other molecular biological principles. To minimize false positive results, any variants that do not meet internal quality standards are confirmed by Sanger sequencing. Variants classified as pathogenic, likely pathogenic, or risk allele which are located in the coding regions and nearby intronic regions (+/- 20bp) of the genes listed above are reported. Variants outside these intervals may be reported but are typically not guaranteed. When a single pathogenic or likely pathogenic variant is identified in a clinically relevant gene with autosomal recessive inheritance, the laboratory will attempt to ensure 100% coverage of coding sequences either through NGS or Sanger sequencing technologies ("fill-in"). All genes listed were evaluated for large deletions and/or duplications. However, single exon deletions or duplications will not be detected in this assay, nor will copy number alterations in regions of genes with significant pseudogenes. Putative deletions or duplications are analyzed using Fulgent Germline proprietary pipeline for this specimen. Bioinformatics: The FPLMv2.0 pipeline was used to analyze this specimen.

LIMITATIONS:

General Limitations

These test results and variant interpretation are based on the proper identification of the submitted specimen, accuracy of any stated familial relationships, and use of the correct human reference sequences at the queried loci. In very rare instances, errors may result due to mix-up or co-mingling of specimens. Positive results do not imply that there are no other contributors, genetic or otherwise, to future pregnancies, and negative results do not rule out the genetic risk to a pregnancy. Official gene names change over time. Fulgent uses the most up to date gene names based on HUGO Gene Nomenclature Committee (https://www.genenames.org) recommendations. If the gene name on report does not match that of ordered gene, please contact the laboratory and details can be provided. Result interpretation is based on the available clinical and family history information for this individual, collected published information, and Alamut annotation available at the time of reporting. This assay is not designed or validated for the detection of low-level mosaicism or somatic mutations. This assay will not detect certain types of genomic aberrations such as translocations, inversions, or repeat expansions other than specified genes. DNA alterations in regulatory regions or deep intronic regions (greater than 20bp from an exon) may not be detected by this test. Unless otherwise indicated, no additional assays have been performed to evaluate genetic changes in this specimen. There are technical limitations on the ability of DNA sequencing to detect small insertions and deletions. Our laboratory uses a sensitive detection algorithm, however these types of alterations are not detected as reliably as single nucleotide variants. Rarely, due to systematic chemical, computational, or human error, DNA variants may be missed. Although next generation sequencing technologies and our bioinformatics analysis significantly reduce the confounding contribution of pseudogene sequences or other highly-homologous sequences, sometimes these may still interfere with the technical ability of the assay to identify pathogenic alterations in both sequencing and deletion/duplication analyses. Deletion/duplication analysis can identify alterations of genomic regions which include one whole gene (buccal swab specimens and whole blood specimens) and are two or more contiguous exons in size (whole blood specimens only); single exon deletions or duplications may occasionally be identified, but are not routinely detected by this test. When novel DNA duplications are identified, it is not possible to discern the genomic location or orientation of the duplicated segment, hence the effect of the duplication cannot be predicted. Where deletions are detected, it is not always possible to determine whether the predicted product will remain in-frame or not. Unless otherwise indicated, deletion/duplication analysis has not been performed in regions that have been sequenced by Sanger.

Patient: 7703, Donor; Sex: M; DOB: MR#: 7703





Gene Specific Notes and Limitations

No gene specific limitations apply to the genes on the tested panel.

SIGNATURE:

Dr. Harry Gao, DABMG, FACMG on 5/20/2025

= Gao

Laboratory Director, Fulgent

DISCLAIMER:

This test was developed, performed, and its performance characteristics determined by Fulgent Therapeutics LLC (CAP# 8042697, CLIA# 05D2043189), 4399 Santa Anita Ave., El Monte, CA 91731. It has not been cleared or approved by the FDA. The laboratory is regulated under CLIA as qualified to perform high-complexity testing. This test is used for clinical purposes. It should not be regarded as investigational or for research. Since genetic variation, as well as systematic and technical factors, can affect the accuracy of testing, the results of testing should always be interpreted in the context of clinical and familial data. For assistance with interpretation of these results, healthcare professionals may contact us directly at (626) 350-0537 or info@fulgentgenetics.com. It is recommended that patients receive appropriate genetic counseling to explain the implications of the test result, including its residual risks, uncertainties and reproductive or medical options.

Patient: 7703, Donor; Sex: M;
DOB: Representation of the second of the s





To view the supplemental table describing the carrier frequencies, detection rates, and residual risks associated with the genes tested on any Beacon panel, please visit the following link:

Beacon Expanded Carrier Screening Supplemental Table



Patient: 7703, Donor; Sex: M; DOB: ; MR#: 7703