

Donor 7628

Genetic Testing Summary

Fairfax Cryobank recommends reviewing this genetic testing summary with your healthcare provider to determine suitability.

Last Updated: 11/25/2025

Donor Reported Ancestry: English, German, Swedish, Irish Jewish Ancestry: No

Genetic Test*	Result	Comments/Donor's Residual Risk**
Chromosome analysis (karyotype)	Normal male karyotype	No evidence of clinically significant chromosome abnormalities
Hemoglobin evaluation	Normal hemoglobin fractionation and MCV/MCH results	Reduced risk to be a carrier for sickle cell anemia, beta thalassemia, alpha thalassemia trait (aa/ and a-/a-) and other hemoglobinopathies
Expanded Genetic Disease Carrier Screening Panel attached- 549 diseases by gene sequencing.	Negative for genes sequenced	
Special Testing		
Gene: SH3TC2	No carrier mutations identified via gene sequencing with deletion and duplication analysis	
Genes: MFN2 and VWF	No diagnostic sequence or copy- number variants were identified via gene sequencing with deletion and duplication analysis	

^{*}No single test can screen for all genetic disorders. A negative screening result significantly reduces, but cannot eliminate, the risk for these conditions in a pregnancy.

^{**}Donor residual risk is the chance the donor is still a carrier after testing negative.

Patient Name: Donor 7628

Date Of Birth:

Gender: Male

Ethnicity:

Northern European

N/A

Caucasian

Patient ID:

Medical Record #: N/A

Collection Kit:

Accession ID:

Case File ID:

Test Information

Ordering Physician:

Clinic Information: Fairfax Cryobank

Phone:

Report Date: 07/22/2024 07/10/2024 Sample Collected: Sample Received: 07/11/2024

Sample Type: Blood



CARRIER SCREENING REPORT

ABOUT THIS SCREEN: Horizon[™] is a carrier screen for specific autosomal recessive and Xlinked diseases. This information can help patients learn their risk of having a child with specific genetic conditions.

ORDER SELECTED: The Horizon Custom

panel was ordered for this patient. Males are not

screened for X-linked diseases

FINAL RESULTS SUMMARY:



Negative for 549 out of 549 diseases

No Pathogenic variants were detected in the genes that were screened. The patient's remaining carrier risk after the negative screening results is listed for each disease/gene on the Horizon website at https://www.natera.com/panel-option/h-all/. Please see the following pages of this report for a comprehensive list of all conditions included on this individual's screen.

Carrier screening is not diagnostic and may not detect all possible pathogenic variants in a given gene.

RECOMMENDATIONS

Individuals who would like to review their Horizon report with a Natera Laboratory Genetic Counselor may schedule a telephone genetic information session by calling 650-249-9090 or visiting naterasession.com. Clinicians with questions may contact Natera at 650-249-9090 or email support@natera.com.

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ne Keen-Kim, Ph.D., FACMGO Laboratory Director, Natora



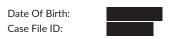
Patient Name: Donor 7628 **Test Information**

Ordering Physician:



Clinic Information:

Fairfax Cryobank



Report Date: 07/22/2024

DISEASES SCREENED

Below is a list of all diseases screened and the result. Certain conditions have unique patient-specific numerical values, therefore, results for those conditions are formatted differently.

Autosomal Recessive

17-BETA HYDROXYSTEROID DEHYDROGENASE 3 DEFICIENCY (HSD17B3) negative

3-BETA-HYDROXYSTEROID DEHYDROGENASE TYPE II DEFICIENCY (HSD3B2) negative 3-HYDROXY-3-METHYLGLUTARYL-COENZYME A LYASE DEFICIENCY (HMGCL) negative 3-HYDROXYACYL-COA DEHYDROGENASE DEFICIENCY (HADH) negative 3-METHYLCROTONYL-CoA CARBOXYLASE 2 DEFICIENCY (MCCC2) negative 3-PHOSPHOGLYCERATE DEHYDROGENASE DEFICIENCY (PHGDH) negative

5-ALPHA-REDUCTASE DEFICIENCY (SRD5A2) negative

6-PYRUVOYL-TETRAHYDROPTERIN SYNTHASE (PTPS) DEFICIENCY (PTS) negative

ABCA4-RELATED CONDITIONS (ABCA4) negative ABETALIPOPROTEINEMIA (MTTP) negative ACHONDROGENESIS, TYPE 1B (SLC26A2) negative ACHROMATOPSIA, CNGB3-RELATED (CNGB3) negative ACRODERMATITIS ENTEROPATHICA (SLC39A4) negative ACTION MYOCLONUS-RENAL FAILURE (AMRF) SYNDROME (SCARB2) negative

ACUTE INFANTILE LIVER FAILURE, TRMU-RELATED (TRMU) negative ACYL-COA OXIDASE I DEFICIENCY (ACOX1) negative AICARDI-GOUTIÈRES SYNDROME (SAMHD1) negative

AICARDI-GOUTIERES SYNDROME, RNASEH2A-RELATED (RNASEH2A) negative AICARDI-GOUTIERES SYNDROME, RNASEH2B-RELATED (RNASEH2B) negative AICARDI-GOUTIERES SYNDROME, RNASEH2C-RELATED (RNASEH2C) negative

AICARDI-GOUTIÈRES SYNDROME, TREX1-RELATED (TREX1) negative

ALPHA-MANNOSIDOSIS (MAN2B1) negative ALPHA-THALASSEMIA (HBA1/HBA2) negative ALPORT SYNDROME, COL4A3-RELATED (COL4A3) negative

ALPORT SYNDROME, COL4A4-RELATED (COL4A4) negative ALSTROM SYNDROME (ALMS1) negative AMISH INFANTILE EPILEPSY SYNDROME (ST3GAL5) negative

ANDERMANN SYNDROME (SLC12A6) negative

ARGININE:GLYCINE AMIDINOTRANSFERASE DEFICIENCY (AGAT DEFICIENCY)

ARGINING ARGIN Regative
ARGININEMIA (ARG1) negative
ARGININOSUCCINATE LYASE DEFICIENCY (ASL) negative

AROMATASE DEFICIENCY (CYP19A1) negative
ASPARAGINE SYNTHETASE DEFICIENCY (ASNS) negative

ASPARTYLGLYCOSAMINURIA (AGA) negative

ATAXIA WITH VITAMIN E DEFICIENCY (TTPA) negative
ATAXIA-TELANGIECTASIA (ATM) negative
ATAXIA-TELANGIECTASIA-LIKE DISORDER 1 (MRE11) negative

ATRANSFERRINEMIA (TF) negative
AUTISM SPECTRUM, EPILEPSY AND ARTHROGRYPOSIS (SLC35A3) negative
AUTOIMMUNE POLYGLANDULAR SYNDROME, TYPE 1 (AIRE) negative

AUTOSOMAL RECESSIVE CONGENITAL ICHTHYOSIS (ARCI), SLC27A4-RELATED

(SLC27A4) negative AUTOSOMAL RECESSIVE SPASTIC ATAXIA OF CHARLEVOIX-SAGUENAY (SACS) negative

BARDET-BIEDL SYNDROME, ARL6-RELATED (ARL6) negative BARDET-BIEDL SYNDROME, BBS10-RELATED (BBS10) negative BARDET-BIEDL SYNDROME, BBS12-RELATED (BBS12) negative BARDET-BIEDL SYNDROME, BBS1-RELATED (BBS1) negative BARDET-BIEDL SYNDROME, BBS2-RELATED (BBS2) negative BARDET-BIEDL SYNDROME, BBS2-RELATED (BBS4) negative BARDET-BIEDL SYNDROME, BBS5-RELATED (BBS4) negative BARDET-BIEDL SYNDROME, BBS7-RELATED (BBS7) negative BARDET-BIEDL SYNDROME, BBS7-RELATED (BBS7) negative BARDET-BIEDL SYNDROME, BBS9-RELATED (BBS9) negative BARDET-BIEDL SYNDROME, TTC8-RELATED (TTC8) negative BARE LYMPHOCYTE SYNDROME, CIITA-RELATED (CIITA) negative BARTTER SYNDROME, BSND-RELATED (BSND) negative BARTTER SYNDROME, KCNJ1-RELATED (KCNJ1) negative BARTTER SYNDROME, SLC12A1-RELATED (SLC12A1) negative BATTEN DISEASE, CLN3-RELATED (CLN3) negative BETA-HEMOGLOBINOPATHIES (HBB) negative BETA-KETOTHIOLASE DEFICIENCY (ACAT1) negative BETA-MANNOSIDOSIS (MANBA) negative BETA-UREIDOPROPIONASE DEFICIENCY (UPB1) negative

BILATERAL FRONTOPARIETAL POLYMICROGYRIA (GPR56) negative

BIOTINIDASE DEFICIENCY (BTD) negative BIOTIN-THIAMINE-RESPONSIVE BASAL GANGLIA DISEASE (BTBGD) (SLC19A3) negative

BLOOM SYNDROME (BLM) negative
BRITTLE CORNEA SYNDROME 1 (ZNF469) negative

BRITTLE CORNEA SYNDROME 2 (PRDM5) negative

CANAVAN DISEASE (ASPA) negative

CARBAMOYL PHOSPHATE SYNTHETASE I DEFICIENCY (CPS1) negative

CARNITINE DEFICIENCY (SLC22A5) negative
CARNITINE PALMITOYLTRANSFERASE IA DEFICIENCY (CPT1A) negative
CARNITINE PALMITOYLTRANSFERASE II DEFICIENCY (CPT2) negative

CARNITINE-ACYLCARNITINE TRANSLOCASE DEFICIENCY (SLC25A20) negative

CARPENTER SYNDROME (RAB23) negative
CARTILAGE-HAIR HYPOPLASIA (RMRP) negative
CATECHOLAMINERGIC POLYMORPHIC VENTRICULAR TACHYCARDIA (CASQ2) negative CATECHOLAMINERGIC POLYMORPHIC VENTRICULAR TACHYCARDIA (CASQ2) negat CD59-MEDIATED HEMOLYTIC ANEMIA (CD59) negative CEP152-RELATED MICROCEPHALY (CEP152) negative CEREBRAL DYSGENESIS, NEUROPATHY, ICHTHYOSIS, AND PALMOPLANTAR KERATODERMA (CEDNIK) SYNDROME (SNAP29) negative CEREBROTENDINOUS XANTHOMATOSIS (CYP27A1) negative CHARCOT-MARIE-TOOTH DISEASE, RECESSIVE INTERMEDIATE C (PLEKHG5) negative CHARCOT-MARIE-TOOTH-DISEASE, TYPE 4D (NDRG1) negative

CHEDIAK-HIGASHI SYNDROME (LYST) negative CHOREOACANTHOCYTOSIS (VPS13A) negative CHRONIC GRANULOMATOUS DISEASE, CYBA-RELATED (CYBA) negative

CHRONIC GRANULOMATOUS DISEASE, NCF2-RELATED (NCF2) negative CILIOPATHIES, RPGRIP1L-RELATED (RPGRIP1L) negative CITRIN DEFICIENCY (SLC25A13) negative

CITRULLINEMIA, TYPE 1 (ASS1) negative

CIN10 DISEASE (CTSD) negative
COHEN SYNDROME (VP513B) negative
COL11A2-RELATED CONDITIONS (COL11A2) negative
COMBINED MALONIC AND METHYLMALONIC ACIDURIA (ACSF3) negative

COMBINED OXIDATIVE PHOSPHORYLATION DEFICIENCY 1 (GFM1) negative COMBINED OXIDATIVE PHOSPHORYLATION DEFICIENCY 3 (TSFM) negative

COMBINED PITUITARY HORMONE DEFICIENCY 1 (POU1F1) negative

COMBINED PITUITARY HORMONE DEFICIENCY-2 (PROP1) negative CONGENITAL ADRENAL HYPERPLASIA, 11-BETA-HYDROXYLASE DEFICIENCY

(CYP11B1) negative

CONGENITAL ADRENAL HYPERPLASIA, 17-ALPHA-HYDROXYLASE DEFICIENCY

(CYP17A1) negative CONGENITAL ADRENAL HYPERPLASIA, 21-HYDROXYLASE DEFICIENCY (CYP21A2) negative

CONGENITAL ADRENAL INSUFFICIENCY, CYP11A1-RELATED (CYP11A1) negative CONGENITAL AMEGAKARYOCYTIC THROMBOCYTOPENIA (MPL) negative

CONGENITAL CHRONIC DIARRHEA (DGAT1) negative

CONGENITAL DISORDER OF GLYCOSYLATION TYPE 1, ALG1-RELATED (ALG1) negative CONGENITAL DISORDER OF GLYCOSYLATION, TYPE 1A, PMM2-Related (PMM2) negative CONGENITAL DISORDER OF GLYCOSYLATION, TYPE 1B (MPI) negative

CONGENITAL DISORDER OF GLYCOSYLATION, TYPE 1C (ALG6) negative CONGENITAL DISORDER OF GLYCOSYLATION, TYPE 1C (ALG6) negative CONGENITAL DYSERYTHROPOIETIC ANEMIA TYPE 2 (SEC23B) negative CONGENITAL FINNISH NEPHROSIS (NPHS1) negative CONGENITAL HYDROCEPHALUS 1 (CCDC88C) negative

CONGENITAL HYPERINSULINISM, KCNJ11-Related (KCNJ11) negative CONGENITAL INSENSITIVITY TO PAIN WITH ANHIDROSIS (CIPA) (NTRK1) negative CONGENITAL MYASTHENIC SYNDROME, CHAT-RELATED (CHAT) negative

CONGENITAL MYASTHENIC SYNDROME, CHRNE-RELATED (CHRNE) negative

CONGENITAL MYASTHENIC SYNDROME, COLQ-RELATED (COLQ) negative CONGENITAL MYASTHENIC SYNDROME, DOK7-RELATED (DOK7) negative

CONGENITAL MYASTHENIC SYNDROME, RAPSN-RELATED (RAPSN) negative

CONGENITAL NEPHROTIC SYNDROME, PICE1-RELATED (PLCE1) negative CONGENITAL NEUTROPENIA, G6PC3-RELATED (G6PC3) negative CONGENITAL NEUTROPENIA, HAX1-RELATED (HAX1) negative

CONGENITAL NEUTROPENIA, VPS45-RELATED (VPS45) negative
CONGENITAL SECRETORY CHLORIDE DIARRHEA 1 (SLC26A3) negative
CORNEAL DYSTROPHY AND PERCEPTIVE DEAFNESS (SLC4A11) negative

CORTICOSTERONE METHYLOXIDASE DEFICIENCY (CYP11B2) negative

COSTEFF SYNDROME (3-METHYLGLUTACONIC ACIDURIA, TYPE 3) (OPA3) negative CRB1-RELATED RETINAL DYSTROPHIES (CRB1) negative CYSTIC FIBROSIS (CFTR) negative

CYSTINOSIS (CTNS) negative

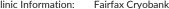
CYTOCHROME C OXIDASE DEFICIENCY, PET100-RELATED (PET100) negative CYTOCHROME P450 OXIOREDUCTASE DEFICIENCY (POR) negative

D-BIFUNCTIONAL PROTEIN DEFICIENCY (HSD17B4) negative



Patient Name: Donor 7628 **Test Information**

Ordering Physician:



Clinic Information: Fairfax Cryobank

Date Of Birth: Case File ID:

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DEAFNESS, AUTOSOMAL RECESSIVE 77 (LOXHD1) negative DIHYDROPTERIDINE REDUCTASE (DHPR) DEFICIENCY (QDPR) negative DONNAI-BARROW SYNDROME (LRP2) negative DUBIN-JOHNSON SYNDROME (ABCC2) negative
DYSKERATOSIS CONGENITA SPECTRUM DISORDERS (TERT) negative
DYSKERATOSIS CONGENITA, RTEL1-RELATED (RTEL1) negative DYSTROPHIC EPIDERMOLYSIS BULLOSA, COL7A1-Related (COL7A1) negative

EARLY INFANTILE EPILEPTIC ENCEPHALOPATHY, CAD-RELATED (CAD) negative EHLERS-DANLOS SYNDROME TYPE VI (PLOD1) negative EHLERS-DANLOS SYNDROME, CLASSIC-LIKE, TNXB-RELATED (TNXB) negative EHLERS-DANLOS SYNDROME, CLASSIC-LIRE, TRAB-RELATED (TAX EHLERS-DANLOS SYNDROME, TYPE VII C (ADAMTS2) negative ELLIS-VAN CREVELD SYNDROME, EVC2-RELATED (EVC2) negative ELLIS-VAN CREVELD SYNDROME, EVC-RELATED (EVC) negative ENHANCED S-CONE SYNDROME (NR2E3) negative EPIMERASE DEFICIENCY (GALACTOSEMIA TYPE III) (GALE) negative EPIPHYSEAL DYSPLASIA, MULTIPLE, 7/DESBUQUOIS DYSPLASIA 1 (CANT1) negative ERCC6-RELATED DISORDERS (ERCC6) negative ERCC8-RELATED DISORDERS (ERCC8) negative ETHYLMALONIC ENCEPHALOPATHY (ETHE1) negative

FACTOR XI DEFICIENCY (F11) negative

FAMILIAL DYSAUTONOMIA (IKBKAP) negative FAMILIAL HEMOPHAGOCYTIC LYMPHOHISTIOCYTOSIS, PRF1-RELATED (PRF1) negative FAMILIAL HEMOPHAGOCYTIC LYMPHOHISTIOCYTOSIS, STX11-RELATED (STX11) negative FAMILIAL HEMOPHAGOCYTIC LYMPHOHISTIOCYTOSIS, STXBP2-RELATED (STXBP2) negative FAMILIAL HEMOPHAGOCYTIC LYMPHOHISTIOCYTOSIS, UNC13D-RELATED (UNC13D) negative
FAMILIAL HYPERCHOLESTEROLEMIA, LDLRAP1-RELATED (LDLRAP1) negative
FAMILIAL HYPERCHOLESTEROLEMIA, LDLRAP1-RELATED (LDLRAP1) negative FAMILIAL HYPERINSULINISM, ABCC8-RELATED (ABCC8) negative FAMILIAL NEPHROGENIC DIABETES INSIPIDUS, AQP2-RELATED (AQP2) negative FANCONI ANEMIA, GROUP A (FANCA) negative FANCONI ANEMIA, GROUP C (FANCC) negative FANCONI ANEMIA, GROUP D2 (FANCD2) negative FANCONI ANEMIA, GROUP E (FANCE) negative FANCONI ANEMIA, GROUP F (FANCF) negative FANCONI ANEMIA, GROUP F (FANCF) negative FANCONI ANEMIA, GROUP I (FANCI) negative FANCONI ANEMIA, GROUP J (BRIP1) negative FANCONI ANEMIA, GROUP J (FANCI) negative FARBER LIPOGRANULOMATOSIS (ASAH1) negative FOVEAL HYPOPLASIA (SLC38A8) negative FRASER SYNDROME 3, GRIP1-RELATED (GRIP1) negative

FRASER SYNDROME, FREM2-RELATED (FREM2) negative FRIEDREICH ATAXIA (FXN) negative FRUCTOSE-1,6-BISPHOSPHATASE DEFICIENCY (FBP1) negative FUCOSIDOSIS, FUCA1-RELATED (FUCA1) negative FUMARASE DEFICIENCY (FH) negative

GABA-TRANSAMINASE DEFICIENCY (ABAT) negative GALACTOKINASE DEFICIENCY (GALACTOSEMIA, TYPE II) (GALK1) negative GALACTOSEMIA (GALT) negative GALACTOSIALIDOSIS (CTSA) negative GAUCHER DISEASE (GBA) negative GCH1-RELATED CONDITIONS (GCH1) negative GCH1/RELATED CONDITIONS (GCH1) negative GDF5-RELATED CONDITIONS (GDF5) negative GERODERMA OSTEODYSPLASTICA (GORAB) negative GITELMAN SYNDROME (SLC12A3) negative GLANZMANN THROMBASTHENIA (ITGB3) negative GLANZMANN I HROMBASI HENIA (I/GB3) negative
GLUTARIC ACIDEMIA, TYPE 1 (GCDH) negative
GLUTARIC ACIDEMIA, TYPE 2A (ETFA) negative
GLUTARIC ACIDEMIA, TYPE 2B (ETFB) negative
GLUTARIC ACIDEMIA, TYPE 2C (ETFDH) negative
GLUTA GLYCOGEN STORAGE DISEASE TYPE 5 (McArdle Disease) (PYGM) negative GLYCOGEN STORAGE DISEASE TYPE IXB (PHKB) negative GLYCOGEN STORAGE DISEASE TYPE IXC (PHKG2) negative GLYCOGEN STORAGE DISEASE, TYPE 1a (G6PC) negative GLYCOGEN STORAGE DISEASE, TYPE 1b (SLC37A4) negative GLYCOGEN STORAGE DISEASE, TYPE 2 (POMPE DISEASE) (GAA) negative

FRASER SYNDROME, FRAS1-RELATED (FRAS1) negative

GRACILE SYNDROME (BCS1L) negative GUANIDINOACETATE METHYLTRANSFERASE DEFICIENCY (GAMT) negative HARLEQUIN ICHTHYOSIS (ABCA12) negative HEME OXYGENASE 1 DEFICIENCY (HMOX1) negative HEMOCHROMATOSIS TYPE 2A (HFE2) negative

HEMOCHROMATOSIS, TYPE 3, TFR2-Related (TFR2) negative
HEPATOCEREBRAL MITOCHONDRIAL DNA DEPLETION SYNDROME, MPV17-RELATED (MPV17) negative

HEREDITARY FRUCTOSE INTOLERANCE (ALDOB) negative HEREDITARY HEMOCHROMATOSIS TYPE 2B (HAMP) negative HEREDITARY SPASTIC PARAPARESIS, TYPE 49 (TECPR2) negative HEREDITARY SPASTIC PARAPARESIS, 17PE 49 (TECPK2) negative HERDITARY SPASTIC PARAPLEGIA, CYP7B1-RELATED (CYP7B1) negative HERMANSKY-PUDLAK SYNDROME, AP3B1-RELATED (AP3B1) negative HERMANSKY-PUDLAK SYNDROME, BLOC1S3-RELATED (BLOC1S3) negative HERMANSKY-PUDLAK SYNDROME, BLOC1S6-RELATED (BLOC1S6) negative HERMANSKY-PUDLAK SYNDROME, HP51-RELATED (HP51) negative HERMANSKY-PUDLAK SYNDROME, HPS3-RELATED (HP53) negative HERMANSKY-PUDLAK SYNDROME, HPS4-RELATED (HP54) negative HERMANSKY-PUDLAK SYNDROME, HPS5-RELATED (HPS5) negative HERMANSKY-PUDLAK SYNDROME, HPS6-RELATED (HPS6) negative HOLOCARBOXYLASE SYNTHETASE DEFICIENCY (HLCS) negative HOMOCYSTINURIA AND MEGALOBLASTIC ANEMIA TYPE CBLG (MTR) negative HOMOCYSTINURIA DUE TO DEFICIENCY OF MTHFR (MTHFR) negative HOMOCYSTINURIA, CBS-RELATED (CBS) negative HOMOCYSTINURIA, Type cblE (MTRR) negative HYDROLETHALUS SYNDROME (HYLS1) negative

HYPER-IGM IMMUNODEFICIENCY (CD40) negative HYPERORNITHINEMIA-HYPERAMMONEMIA-HOMOCITRULLINURIA (HHH SYNDROME) (SLC25A15) negative HYPERPHOSPHATEMIC FAMILIAL TUMORAL CALCINOSIS, GALNT3-RELATED

(GALNT3) negative HYPOMYELINATING LEUKODYSTROPHY 12 (VPS11) negative HYPOPHOSPHATASIA, ALPL-RELATED (ALPL) negative

IMERSLUND-GRÄSBECK SYNDROME 2 (AMN) negative IMMUNODEFICIENCY-CENTROMERIC INSTABILITY-FACIAL ANOMALIES (ICF) SYNDROME, DNMT3B-RELATED (DNMT3B) negative IMMUNODEFICIENCY-CENTROMERIC INSTABILITY-FACIAL ANOMALIES (ICF) SYNDROME, ZBTB24-RELATED (ZBTB24) negative INCLUSION BODY MYOPATHY 2 (GNE) negative
INFANTILE CEREBRAL AND CEREBELLAR ATROPHY (MED17) negative INFANTILE NEPHRONOPHTHISIS (INVS) negative INFANTILE NEPHRONOPHI HISIS (INVS) negative
INFANTILE NEUROAXONAL DYSTROPHY (PLA2G6) negative
ISOLATED ECTOPIA LENTIS (ADAMTSL4) negative
ISOLATED SULFITE OXIDASE DEFICIENCY (SUOX) negative
ISOLATED THYROID-STIMULATING HORMONE DEFICIENCY (TSHB) negative ISOVALERIC ACIDEMIA (IVD) negative

JOHANSON-BLIZZARD SYNDROME (UBR1) negative
JOUBERT SYNDROME 2 / MECKEL SYNDROME 2 (TMEM216) negative
JOUBERT SYNDROME AND RELATED DISORDERS (JSRD), TMEM67-RELATED JOUBERT SYNDROME, AHI1-RELATED (AHI1) negative
JOUBERT SYNDROME, ARL13B-RELATED (ARL13B) negative JOUBERT SYNDROME, B9D1-RELATED (B9D1) negative JOUBERT SYNDROME, B9D2-RELATED (B9D2) negative JOUBERT SYNDROME, C2CD3-RELATED/OROFACIODIGITAL SYNDROME 14 (C2CD3) negative JOUBERT SYNDROME, CC2D2A-RELATED/COACH SYNDROME (CC2D2A) negative JOUBERT SYNDROME, CEP104-RELATED/COACH 31NDROME (CED2A) Regative
JOUBERT SYNDROME, CEP120-RELATED/CF04) negative
JOUBERT SYNDROME, CEP120-RELATED/SHORT-RIB THORACIC DYSPLASIA 13 WITH OR
WITHOUT POLYDACTYLY (CEP120) negative
JOUBERT SYNDROME, CEP41-RELATED (CEP41) negative
JOUBERT SYNDROME, CPLANE1-RELATED / OROFACIODIGITAL SYNDROME 6 (CPLANE1) negative (CPLANEL) negative
JOUBERT SYNDROME, CSPP1-RELATED (CSPP1) negative
JOUBERT SYNDROME, INPP5E-RELATED (INPP5E) negative
JUNCTIONAL EPIDERMOLYSIS BULLOSA, COL17A1-RELATED (COL17A1) negative
JUNCTIONAL EPIDERMOLYSIS BULLOSA, ITGA6-RELATED (ITGA6) negative JUNCTIONAL EPIDERMOLYSIS BULLOSA, ITGB4-RELATED (ITGB4) negative JUNCTIONAL EPIDERMOLYSIS BULLOSA, LAMB3-RELATED (LAMB3) negative JUNCTIONAL EPIDERMOLYSIS BULLOSA, LAMC2-RELATED (LAMC2) negative JUNCTIONAL EPIDERMOLYSIS BULLOSA/LARYNGOONYCHOCUTANEOUS SYNDROME, LAMA3-RELATED (LAMA3) negative

KRABBE DISEASE (GALC) negative

LAMELLAR ICHTHYOSIS, TYPE 1 (TGM1) negative



GLYCOGEN STORAGE DISEASE, TYPE 3 (AGL) negative GLYCOGEN STORAGE DISEASE, TYPE 4 (GBE1) negative GLYCOGEN STORAGE DISEASE, TYPE 7 (PFKM) negative

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Ordering Physician:

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LARON SYNDROME (GHR) negative
LEBER CONGENITAL AMAUROSIS 2 (RPE65) negative

LEBER CONGENITAL AMAUROSIS TYPE AIPL1 (AIPL1) negative

LEBER CONGENITAL AMAUROSIS TYPE GUCYZD (GUCYZD) negative
LEBER CONGENITAL AMAUROSIS TYPE TULP1 (TULP1) negative
LEBER CONGENITAL AMAUROSIS, IQCB1-RELATED/SENIOR-LOKEN SYNDROME 5

LEBER CONGENITAL AMAUROSIS, TYPE CEP290 (CEP290) negative

LEBER CONGENITAL AMAUROSIS, TYPE LCA5 (LCA5) negative LEBER CONGENITAL AMAUROSIS, TYPE RDH12 (RDH12) negative

LEIGH SYNDROME, FRENCH-CANADIAN TYPE (LRPPRC) negative LETHAL CONGENITAL CONTRACTURE SYNDROME 1 (GLE1) negative

LEUKOENCEPHALOPATHY WITH VANISHING WHITE MATTER (EIF2B5) negative

LEUKOENCEPHALOPATHY WITH VANISHING WHITE MATTER, EIF2B1-RELATED (EIF2B1) negative LEUKOENCEPHALOPATHY WITH VANISHING WHITE MATTER, EIF2B2-RELATED

(EIF2B2) negative LEUKOENCEPHALOPATHY WITH VANISHING WHITE MATTER, EIF2B3-RELATED (EIF2B3) negative

LEUKOENCEPHALOPATHY WITH VANISHING WHITE MATTER, EIF2B4-RELATED

(EIF2B4) negative
LIG4 SYNDROME (LIG4) negative
LIMB-GIRDLE MUSCULAR DYSTROPHY TYPE 8 (TRIM32) negative

LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2A (CAPN3) negative

LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 28 (DYSF) negative LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2C (SGCG) negative

LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2D (SGCA) negative LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2D (SGCA) negative LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2E (SGCB) negative LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2F (SGCD) negative LIMB-GIRDLE MUSCULAR DYSTROPHY, TYPE 2I (FKRP) negative

LIPOAMIDE DEHYDROGENASE DEFICIENCY (DIHYDROLIPOAMIDE DEHYDROGENASE DEFICIENCY) (DLD) negative LIPOID ADRENAL HYPERPLASIA (STAR) negative

LIPOPROTEIN LIPASE DEFICIENCY (LPL) negative

LONG CHAIN 3-HYDROXYACYL-COA DEHYDROGENASE DEFICIENCY (HADHA) negative LRAT-RELATED CONDITIONS (LRAT) negative LUNG DISEASE, IMMUNODEFICIENCY, AND CHROMOSOME BREAKAGE SYNDROME

(LICS) (NSMCE3) negative
LYSINURIC PROTEIN INTOLERANCE (SLC7A7) negative

MALONYL-COA DECARBOXYLASE DEFICIENCY (MLYCD) negative MAPLE SYRUP URINE DISEASE, TYPE 1A (BCKDHA) negative MAPLE SYRUP URINE DISEASE, TYPE 1B (BCKDHB) negative

MAPLE SYRUP URINE DISEASE, TYPE 2 (DBT) negative MCKUSICK-KAUFMAN SYNDROME (MKKS) negative

MECKEL SYNDROME 7/NEPHRONOPHTHISIS 3 (NPHP3) negative

MECKEL-GRUBER SYNDROME, TYPE 1 (MKS1) negative

MECR-RELATED NEUROLOGIC DISORDER (MECR) negative MEDIUM CHAIN ACYL-CoA DEHYDROGENASE DEFICIENCY (ACADM) negative

MEDNIK SYNDROME (AP1S1) negative

MEGALENCEPHALIC LEUKOENCEPHALOPATHY WITH SUBCORTICAL CYSTS

(MLC1) negative
MEROSIN-DEFICIENT MUSCULAR DYSTROPHY (LAMA2) negative METABOLIC ENCEPHALOPATHY AND ARRHYTHMIAS, TANGO2-RELATED

(TANGO2) negative METACHROMATIC LEUKODYSTROPHY, ARSA-RELATED (ARSA) negative METACHROMATIC LEUKODYSTROPHY, PSAP-RELATED (PSAP) negative

METHYLMALONIC ACIDEMIA AND HOMOCYSTINURIA TYPE CBLF (LMBRD1) negative

METHYLMALONIC ACIDEMIA, MCEE-RELATED (MCEE) negative
METHYLMALONIC ACIDURIA AND HOMOCYSTINURIA, TYPE CBLC (MMACHC) negative
METHYLMALONIC ACIDURIA AND HOMOCYSTINURIA, TYPE CblD (MMADHC) negative

METHYLMALONIC ACIDURIA, MMAA-RELATED (MMAA) negative METHYLMALONIC ACIDURIA, MMAB-RELATED (MMAB) negative METHYLMALONIC ACIDURIA, TYPE MUT(0) (MUT) negative

MEVALONIC KINASE DEFICIENCY (MVK) negative
MICROCEPHALIC OSTEODYSPLASTIC PRIMORDIAL DWARFISM TYPE II (PCNT) negative
MICROPHTHALMIA / ANOPHTHALMIA, VSX2-RELATED (VSX2) negative

MITOCHONDRIAL COMPLEX 1 DEFICIENCY, ACAD9-RELATED (ACAD9) negative

MITOCHONDRIAL COMPLEX 1 DEFICIENCY, NDUFAF5-RELATED (NDUFAF5) negative MITOCHONDRIAL COMPLEX 1 DEFICIENCY, NDUFS6-RELATED (NDUFS6) negative MITOCHONDRIAL COMPLEX I DEFICIENCY, NUCLEAR TYPE 1 (NDUFS4) negative

MITOCHONDRIAL COMPLEX I DEFICIENCY, NUCLEAR TYPE 10 (NDUFAF2) negative MITOCHONDRIAL COMPLEX I DEFICIENCY, NUCLEAR TYPE 17 (NDUFAF6) negative MITOCHONDRIAL COMPLEX I DEFICIENCY, NUCLEAR TYPE 19 (FOXRED1) negative

MITOCHONDRIAL COMPLEX I DEFICIENCY, NUCLEAR TYPE 3 (NDUFS7) negative MITOCHONDRIAL COMPLEX I DEFICIENCY, NUCLEAR TYPE 4 (NDUFV1) negative MITOCHONDRIAL COMPLEX IV DEFICIENCY, NUCLEAR TYPE 2, SCO2-RELATED

(SCO2) negative

MITOCHONDRIAL COMPLEX IV DEFICIENCY, NUCLEAR TYPE 6 (COX15) negative MITOCHONDRIAL DNA DEPLETION SYNDROME 2 (TK2) negative

MITOCHONDRIAL DNA DEPLETION SYNDROME 3 (DGUOK) negative MITOCHONDRIAL MYOPATHY AND SIDEROBLASTIC ANEMIA (MLASA1) (PUS1) negative

MITOCHONDRIAL TRIFUNCTIONAL PROTEIN DEFICIENCY, HADHB-RELATED

MOLYBDENUM COFACTOR DEFICIENCY TYPE B (MOCS2) negative MOLYBDENUM COFACTOR DEFICIENCY, TYPE A (MOCS1) negative

MUCOLIPIDOSIS II/III A (GNPTAB) negative

MUCOLIPIDOSIS III GAMMA (GNPTG) negative
MUCOLIPIDOSIS, TYPE IV (MCOLN1) negative
MUCOPOLYSACCHARIDOSIS, TYPE I (HURLER SYNDROME) (IDUA) negative

MUCOPOLYSACCHARIDOSIS, TYPE III A (SANFILIPPO A) (SGSH) negative MUCOPOLYSACCHARIDOSIS, TYPE III B (SANFILIPPO B) (NAGLU) negative MUCOPOLYSACCHARIDOSIS, TYPE III C (SANFILIPPO C) (HGSNAT) negative

MUCOPOLYSACCHARIDOSIS, TYPE III D (SANFILIPPO D) (GNS) negative

MUCOPOLYSACCHARIDOSIS, TYPE IV A (MORQUIO SYNDROME) (GALNS) negative MUCOPOLYSACCHARIDOSIS, TYPE IV B/GM1 GANGLIOSIDOSIS (GLB1) negative MUCOPOLYSACCHARIDOSIS, TYPE IX (HYAL1) negative

MUCOPOLYSACCHARIDOSIS, TYPE VI (MAROTEAUX-LAMY) (ARSB) negative

MUCOPOLYSACCHARIDOSIS, TYPE VII (GUSB) negative
MULIBREY NANISM (TR/M37) negative
MULTIPLE PTERYGIUM SYNDROME, CHRNG-RELATED/ESCOBAR SYNDROME

(CHRNG) negative
MULTIPLE SULFATASE DEFICIENCY (SUMF1) negative
MUSCLE-EYE-BRAIN DISEASE, POMGNT1-RELATED (POMGNT1) negative MUSCULAR DYSTROPHY-DYSTROGLYCANOPATHY (RXYLT1) negative

MUSK-RELATED CONGENITAL MYASTHENIC SYNDROME (MUSK) negative MYONEUROGASTROINTESTINAL ENCEPHALOPATHY (MNGIE) (TYMP) negative

MYOTONIA CONGENITA (CLCN1) negative

N-ACETYLGLUTAMATE SYNTHASE DEFICIENCY (NAGS) negative

NEMALINE MYOPATHY, NEB-RELATED (NEB) negative
NEPHRONOPHTHISIS 1 (NPHP1) negative
NEURONAL CEROID LIPOFUSCINOSIS, CLN5-RELATED (CLN5) negative
NEURONAL CEROID LIPOFUSCINOSIS, CLN6-RELATED (CLN6) negative

NEURONAL CEROID LIPOFUSCINOSIS, CLN8-RELATED (CLN8) negative NEURONAL CEROID LIPOFUSCINOSIS, MFSD8-RELATED (MFSD8) negative NEURONAL CEROID LIPOFUSCINOSIS, PPT1-RELATED (PPT1) negative

NEURONAL CEROID LIPOFUSCINOSIS, PP11-RELATED (PP11) negative NEURONAL CEROID LIPOFUSCINOSIS, TPP1-RELATED (TPP1) negative NGLY1-CONGENITAL DISORDER OF GLYCOSYLATION (NGLY1) negative NIEMANN-PICK DISEASE, TYPE C1 / D (NPC1) negative NIEMANN-PICK DISEASE, TYPE C2 (NPC2) negative

NIEMANN-PICK DISEASE, TYPES A / B (SMPD1) negative
NIJMEGEN BREAKAGE SYNDROME (NBN) negative
NON-SYNDROMIC HEARING LOSS, GJB2-RELATED (GJB2) negative

NON-SYNDROMIC HEARING LOSS, MYO15A-RELATED (MYO15A) negative NONSYNDROMIC HEARING LOSS, OTOA-RELATED (OTOA) negative NONSYNDROMIC HEARING LOSS, OTOF-RELATED (OTOF) negative

NONSYNDROMIC HEARING LOSS, PJVK-RELATED (PJVK) negative

NONSYNDROMIC HEARING LOSS, SYNE4-RELATED (SYNE4) negative NONSYNDROMIC HEARING LOSS, TMC1-RELATED (TMC1) negative

NONSYNDROMIC HEARING LOSS, TMPRSS3-RELATED (TMPRSS3) negative

NONSYNDROMIC INTELLECTUAL DISABILITY (CC2D1A) negative

NORMOPHOSPHATEMIC TUMORAL CALCINOSIS (SAMD9) negative

OCULOCUTANEOUS ALBINISM TYPE IV (SLC45A2) negative OCULOCUTANEOUS ALBINISM TYPE, III (TYRP1) negative

OCULOCUTANEOUS ALBINISM, OCA2-RELATED (OCA2) negative

OCULOCUTANEOUS ALBINISM, TYPES 1A AND 1B (TYR) negative

ODONTO-ONYCHO-DERMAL DYSPLASIA / SCHOPF-SCHULZ-PASSARGE SYNDROME (WNT10A) negative

OMENN SYNDROME, RAG2-RELATED (RAG2) negative

ORNITHINE AMINOTRANSFERASE DEFICIENCY (OAT) negative OSTEOGENESIS IMPERFECTA TYPE VII (CRTAP) negative

OSTEOGENESIS IMPERFECTA TYPE VIII (P3H1) negative

OSTEOGENESIS IMPERFECTA TYPE XI (FKBP10) negative
OSTEOGENESIS IMPERFECTA TYPE XIII (BMP1) negative
OSTEOPETROSIS, INFANTILE MALIGNANT, TCIRG1-RELATED (TCIRG1) negative

OSTEOPETROSIS, OSTM1-RELATED (OSTM1) negative

PANTOTHENATE KINASE-ASSOCIATED NEURODEGENERATION (PANK2) negative

PAPILLON LEFÈVRE SYNDROME (CTSC) negative PARKINSON DISEASE 15 (FBXO7) negative

PENDRED SYNDROME (SLC26A4) negative

PERLMAN SYNDROME (DIS3L2) negative PGM3-CONGENITAL DISORDER OF GLYCOSYLATION (PGM3) negative PHENYLKETONURIA (PAH) negative

PIGN-CONGENITAL DISORDER OF GLYCOSYLATION (PIGN) negative

PITUITARY HORMONE DEFICIENCY, COMBINED 3 (LHX3) negative POLG-RELATED DISORDERS (POLG) negative



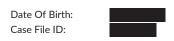
Patient Name: Donor 7628 **Test Information**

Ordering Physician:



Clinic Information:





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POLYCYSTIC KIDNEY DISEASE, AUTOSOMAL RECESSIVE (PKHD1) negative PONTOCEREBELLAR HYPOPLASIA, EXOSC3-RELATED (EXOSC3) negative PONTOCEREBELLAR HYPOPLASIA, RARS2-RELATED (RARS2) negative PONTOCEREBELLAR HYPOPLASIA, TSEN2-RELATED (TSEN2) negative PONTOCEREBELLAR HYPOPLASIA, TSEN54-RELATED (TSEN54) negative PONTOCEREBELLAR HYPOPLASIA, TYPE 1A (VRK1) negative PONTOCEREBELLAR HYPOPLASIA, TYPE 2D (SEPSECS) negative PONTOCEREBELLAR HYPOPLASIA, VPS53-RELATED (VPS53) negative PRIMARY CILIARY DYSKINESIA, CCDC103-RELATED (CCDC103) negative PRIMARY CILIARY DYSKINESIA, CCDC39-RELATED (CCDC39) negative PRIMARY CILIARY DYSKINESIA, DNAH11-RELATED (DNAH11) negative PRIMARY CILIARY DYSKINESIA, DNAH5-RELATED (DNAH5) negative PRIMARY CILIARY DYSKINESIA, DNAI1-RELATED (DNAI1) negative PRIMARY CILIARY DYSKINESIA, DIAI1-RELATED (DIAI2) negative
PRIMARY CONGENITAL GLAUCOMA/PETERS ANOMALY (CYP1B1) negative
PRIMARY HYPEROXALURIA, TYPE 1 (AGXT) negative
PRIMARY HYPEROXALURIA, TYPE 2 (GRHPR) negative PRIMARY HYPEROXALURIA, TYPE 3 (HOGA1) negative PRIMARY MICROCEPHALY 1, AUTOSOMAL RECESSIVE (MCPH1) negative PROGRESSIVE EARLY-ONSET ENCEPAHLOPATHY WITH BRAIN ATROPHY AND THIN CORPUS CALLOSUM (TBCD) negative
PROGRESSIVE FAMILIAL INTRAHEPATIC CHOLESTASIS, ABCB4-RELATED (ABCB4) negative
PROGRESSIVE FAMILIAL INTRAHEPATIC CHOLESTASIS, TYPE 1 (PFIC1) (ATP8B1) negative PROGRESSIVE FAMILIAL INTRAHEPATIC CHOLESTASIS, TYPE 2 (ABCB11) negative PROGRESSIVE FAMILIAL INTRAHEPATIC CHOLESTASIS, TYPE 4 (PFIC4) (TJP2) negative PROGRESSIVE PSEUDORHEUMATOID DYSPLASIA (CCN6) negative

PROLIDASE DEFICIENCY (PEPD) negative
PROPIONIC ACIDEMIA, PCCA-RELATED (PCCA) negative
PROPIONIC ACIDEMIA, PCCB-RELATED (PCCB) negative
PSEUDOXANTHOMA ELASTICUM (ABCC6) negative

PTERIN-4 ALPHA-CARBINOLAMINE DEHYDRATASE (PCD) DEFICIENCY (PCBD1) negative PYCNODYSOSTOSIS (CTSK) negative PYRIDOXAL 5'-PHOSPHATE-DEPENDENT EPILEPSY (PNPO) negative

PYRIDOXINE-DEPENDENT EPILEPSY (ALDH7A1) negative

PYRUVATE CARBOXYLASE DEFICIENCY (PC) negative PYRUVATE DEHYDROGENASE DEFICIENCY, PDHB-RELATED (PDHB) negative

REFSUM DISEASE, PHYH-RELATED (PHYH) negative RENAL TUBULAR ACIDOSIS AND DEAFNESS, ATP6V1B1-RELATED (ATP6V1B1) negative RENAL TUBULAR ACIDOSIS, PROXIMAL, WITH OCULAR ABNORMALITIES AND MENTAL RETARDATION (SLC4A4) negative RETINITIS PIGMENTOSA 25 (EYS) negative RETINITIS PIGMENTOSA 26 (CERKL) negative RETINITIS PIGMENTOSA 28 (FAM161A) negative RETINITIS PIGMENTOSA 36 (PRCD) negative RETINITIS PIGMENTOSA 59 (DHDDS) negative

RETINITIS PIGMENTOSA 62 (MAK) negative RHIZOMELIC CHONDRODYSPLASIA PUNCTATA, TYPE 1 (PEX7) negative RHIZOMELIC CHONDRODYSPLASIA PUNCTATA, TYPE 2 (GNPAT) negative RHIZOMELIC CHONDRODYSPLASIA PUNCTATA, TYPE 3 (AGPS) negative

RLBP1-RELATED RETINOPATHY (*RLBP1*) negative ROBERTS SYNDROME (*ESCO2*) negative RYR1-RELATED CONDITIONS (*RYR1*) negative

SALLA DISEASE (SLC17A5) negative SANDHOFF DISEASE (HEXB) negative SCHIMKE IMMUNOOSSEOUS DYSPLASIA (SMARCAL1) negative SCHINDLER DISEASE (NAGA) negative SEGAWA SYNDROME, TH-RELATED (TH) negative SENIOR-LOKEN SYNDROME, IH-RELATED (IH) negative
SENIOR-LOKEN SYNDROME 4/NEPHRONOPHTHISIS 4 (NPHP4) negative
SEPIAPTERIN REDUCTASE DEFICIENCY (SPR) negative
SEVERE COMBINED IMMUNODEFICIENCY (SCID), CD3D-RELATED (CD3D) negative
SEVERE COMBINED IMMUNODEFICIENCY (SCID), CD3E-RELATED (CD3E) negative SEVERE COMBINED IMMUNODEFICIENCY (SCID), FOXN1-RELATED (FOXN1) negative SEVERE COMBINED IMMUNODEFICIENCY (SCID), IKBKB-RELATED (IKBKB) negative SEVERE COMBINED IMMUNODEFICIENCY (SCID), IL7R-RELATED (IL7R) negative SEVERE COMBINED IMMUNODEFICIENCY (SCID), JAK3-RELATED (JAK3) negative SEVERE COMBINED IMMUNODEFICIENCY (SCID), PTPRC-RELATED (PTPRC) negative SEVERE COMBINED IMMUNODEFICIENCY (SCID), RAG1-RELATED (RAG1) negative SEVERE COMBINED IMMUNODEFICIENCY, ADA-Related (ADA) negative SEVERE COMBINED IMMUNODEFICIENCY, TYPE ATHABASKAN (DCLRE1C) negative SHORT-RIB THORACIC DYSPLASIA 3 WITH OR WITHOUT POLYDACTYLY (DYNC2H1) negative SHWACHMAN-DIAMOND SYNDROME, SBDS-RELATED (SBDS) negative SIALIDOSIS (NEU1) negative SJÖGREN-LARSSON SYNDROME (ALDH3A2) negative SMITH-LEMLI-OPITZ SYNDROME (DHCR7) negative SPASTIC PARAPLEGIA, TYPE 15 (ZFYVE26) negative

SPASTIC TETRAPLEGIA, THIN CORPUS CALLOSUM, AND PROGRESSIVE MICROCEPHALY (SPATCCM) (SLC1A4) negative SPG11-RELATED CONDITIONS (SPG11) negative SPINAL MUSCULAR ATROPHY (SMN1) negative SMN1: Two copies; g.27134T>G: absent; the absence of the g.27134T>G variant decreases the chance to be a silent (2+0) carrier. SPINAL MUSCULAR ATROPHY WITH RESPIRATORY DISTRESS TYPE 1 (IGHMBP2) negative SPINOCEREBELLAR ATAXIA, AUTOSOMAL RECESSIVE 10 (ANO10) negative SPINOCEREBELLAR ATAXIA, AUTOSOMAL RECESSIVE 12 (WWOX) negative SPONDYLOCOSTAL DYSOSTOSIS 1 (DLL3) negative SPONDYLOTHORACIC DYSOSTOSIS, MESP2-Related (MESP2) negative STEEL SYNDROME (COL27A1) negative
STEROID-RESISTANT NEPHROTIC SYNDROME (NPHS2) negative
STUVE-WIEDEMANN SYNDROME (LIFR) negative SURF1-RELATED CONDITIONS (SURF1) negative SURFACTANT DYSFUNCTION, ABCA3-RELATED (ABCA3) negative

TAY-SACHS DISEASE (HEXA) negative
TBCE-RELATED CONDITIONS (TBCE) negative
THIAMINE-RESPONSIVE MEGALOBLASTIC ANEMIA SYNDROME (SLC19A2) negative THYROID DYSHORMONOGENESIS 1 (SLC5A5) negative THYROID DYSHORMONOGENESIS 2A (TPO) negative THYROID DYSHORMONOGENESIS 3 (TG) negative THYROID DYSHORMONOGENESIS 6 (DUOX2) negative TRANSCOBALAMIN II DEFICIENCY (TCN2) negative TRICHOHEPATOENTERIC SYNDROME, SKIC2-RELATED (SKIC2) negative TRICHOHEPATOENTERIC SYNDROME, TTC37-RELATED (TTC37) negative TRICHOTHIODYSTROPHY 1/XERODERMA PIGMENTOSUM, GROUP D (ERCC2) negative TRIMETHYLAMINURIA (FMO3) negative TRIPLE A SYNDROME (AAAS) negative TSHR-RELATED CONDITIONS (TSHR) negative TYROSINEMIA TYPE III (*IPD*) negative TYROSINEMIA, TYPE 1 (*FAH*) negative TYROSINEMIA, TYPE 2 (*TAT*) negative

USHER SYNDROME, TYPE 1B (MYO7A) negative USHER SYNDROME, TYPE 1C (USH1C) negative USHER SYNDROME, TYPE 1D (CDH23) negative USHER SYNDROME, TYPE 1F (PCDH15) negative USHER SYNDROME, TYPE 1J/DEAFNESS, AUTOSOMAL RECESSIVE, 48 (CIB2) negative USHER SYNDROME, TYPE 2A (USH2A) negative USHER SYNDROME, TYPE 2C (ADGRV1) negative USHER SYNDROME, TYPE 3 (CLRN1) negative

VERY LONG-CHAIN ACYL-CoA DEHYDROGENASE DEFICIENCY (ACADVL) negative VICI SYNDROME (EPG5) negative VITAMIN D-DEPENDENT RICKETS, TYPE 1A (CYP27B1) negative VITAMIN D-RESISTANT RICKETS TYPE 2A (VDR) negative VLDLR-ASSOCIATED CEREBELLAR HYPOPLASIA (VLDLR) negative

WALKER-WARBURG SYNDROME, CRPPA-RELATED (CRPPA) negative WALKER-WARBURG SYNDROME, FKTN-RELATED (FKTN) negative WALKER-WARBURG SYNDROME, LARGE1-RELATED (LARGE1) negative WALKER-WARBURG SYNDROME, POMT1-RELATED (POMT1) negative WALKER-WARBURG SYNDROME, POMT2-RELATED (POMT2) negative WARSAW BREAKAGE SYNDROME (DDX11) negative WERNER SYNDROME (WRN) negative
WILSON DISEASE (ATP7B) negative
WOLCOTT-RALLISON SYNDROME (EIF2AK3) negative WOLMAN DISEASE (LIPA) negative
WOODHOUSE-SAKATI SYNDROME (DCAF17) negative

XERODERMA PIGMENTOSUM VARIANT TYPE (POLH) negative XERODERMA PIGMENTOSUM, GROUP A (XPA) negative XERODERMA PIGMENTOSUM, GROUP C (XPC) negative

ZELLWEGER SPECTRUM DISORDER, PEX13-RELATED (PEX13) negative ZELLWEGER SPECTRUM DISORDER, PEX16-RELATED (PEX16) negative ZELLWEGER SPECTRUM DISORDER, PEX5-RELATED (PEX5) negative ZELLWEGER SPECTRUM DISORDERS, PEX10-RELATED (PEX10) negative ZELLWEGER SPECTRUM DISORDERS, PEX12-RELATED (PEX12) negative ZELLWEGER SPECTRUM DISORDERS, PEX1-RELATED (PEX1) negative ZELLWEGER SPECTRUM DISORDERS, PEX26-RELATED (PEX26) negative ZELLWEGER SPECTRUM DISORDERS, PEX26-RELATED (PEX26) negative ZELLWEGER SPECTRUM DISORDERS, PEX26-RELATED (PEX27) negative



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 ${\bf Z}$ ZELLWEGER SPECTRUM DISORDERS, PEX6-RELATED (PEX6) ${\bf negative}$



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Testing Methodology, Limitations, and Comments:

Next-generation sequencing (NGS)

Sequencing library prepared from genomic DNA isolated from a patient sample is enriched for targets of interest using standard hybridization capture protocols and PCR amplification (for targets specified below). NGS is then performed to achieve the standards of quality control metrics, including a minimum coverage of 99% of targeted regions at 20X sequencing depth. Sequencing data is aligned to human reference sequence, followed by deduplication, metric collection and variant calling (coding region +/- 20bp). Variants are then classified according to ACMGG/AMP standards of interpretation using publicly available databases including but not limited to ENSEMBL, HGMD Pro, ClinGen, ClinVar, 1000G, ESP and gnomAD. Variants predicted to be pathogenic or likely pathogenic for the specified diseases are reported. It should be noted that the data interpretation is based on our current understanding of the genes and variants at the time of reporting. Putative positive sequencing variants that do not meet internal quality standards or are within highly homologous regions are confirmed by Sanger sequencing or genespecific long-range PCR as needed prior to reporting.

Copy Number Variant (CNV) analysis is limited to deletions involving two or more exons for all genes on the panel, in addition to specific known recurrent singleexon deletions. CNVs of small size may have reduced detection rate. This method does not detect gene inversions, single-exonic and sub-exonic deletions (unless otherwise specified), and duplications of all sizes (unless otherwise specified). Additionally, this method does not define the exact breakpoints of detected CNV events. Confirmation testing for copy number variation is performed by specific PCR, Multiplex Ligation-dependent Probe Amplification (MLPA), next generation sequencing, or other methodology.

This test may not detect certain variants due to local sequence characteristics, high/low genomic complexity, homologous sequence, or allele dropout (PCR-based assays). Variants within noncoding regions (promoter, 5'UTR, 3'UTR, deep intronic regions, unless otherwise specified), small deletions or insertions larger than 25bp, low-level mosaic variants, structural variants such as inversions, and/or balanced translocations may not be detected with this technology.

SPECIAL NOTES

For ABCC6, variants in exons 1-9 are not detected due to the presence of regions of high homology.

For CFTR, when the CFTR R117H variant is detected, reflex analysis of the polythymidine variations (5T, 7T and 9T) at the intron 9 branch/acceptor site of the CFTR gene will be performed.

For CYP21A2, targets were enriched using long-range PCR amplification, followed by next generation sequencing. Duplication analysis will only be performed and reported when c.955C>T (p.Q319*) is detected. Sequencing and CNV analysis may have reduced sensitivity, if variants result from complex rearrangements, in trans with a gene deletion, or CYP21A2 gene duplication on one chromosome and deletion on the other chromosome. This analysis cannot detect sequencing variants located on the CYP21A2 duplicated copy.

For DDX11, only NM_030653.3:c.1763 - 1G > C variant will be analyzed and reported.

For GJB2, CNV analysis of upstream deletions of GJB6-D13S1830 (309kb deletion) and GJB6-D13S1854 (232kb deletion) is included.

For HBA1/HBA2, CNV analysis is offered to detect common deletions of -alpha3.7, -alpha4.2, --MED, --SEA, --FIL, --THAI, --alpha20.5, and/or HS-40.

For OTOA, variants in exons 20 - 28 are not analyzed due to high sequence homology.

For RPGRIP1L, variants in exon 23 are not detected due to assay limitation.

For SAMD9, only p.K1495E variant will be analyzed and reported.

Friedreich Ataxia (FXN)

The GAA repeat region of the FXN gene is assessed by trinucleotide PCR assay and capillary electrophoresis. Variances of +/-1 repeat for normal alleles and up to +/-3 repeats for premutation alleles may occur. For fully penetrant expanded alleles, the precise repeat size cannot be determined, therefore the approximate allele size is reported. Sequencing and copy number variants are analyzed by next-generation sequencing analysis.

Friedreich Ataxia Repeat Categories

Categories	GAA Repeat Sizes
Normal	<34
Premutation	34 - 65
Full	>65



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Ordering Physician:

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Spinal Muscular Atrophy (SMN1)

The total combined copy number of SMN1 and SMN2 exon 7 is quantified based on NGS read depth. The ratio of SMN1 to SMN2 is calculated based on the read depth of a single nucleotide that distinguishes these two genes in exon 7. In addition to copy number analysis, testing for the presence of a single nucleotide polymorphism (g.27134T>G in intron 7 of SMN1) associated with the presence of a SMN1 duplication allele is performed using NGS.

Ethnicity	Two SMN1 copies carrier risk before g.27134T>G testing	Carrier risk after g.27134T	Carrier risk after g.27134T>G testing	
		g.27134T>G ABSENT	g.27134T>G PRESENT	
Caucasian	1 in 632	1 in 769	1 in 29	
Ashkenazi Jewish	1 in 350	1 in 580	LIKELY CARRIER	
Asian	1 in 628	1 in 702	LIKELY CARRIER	
African-American	1 in 121	1 in 396	1 in 34	
Hispanic	1 in 1061	1 in 1762	1 in 140	

Variant Classification

Only pathogenic or likely pathogenic variants are reported. Other variants including benign variants, likely benign variants, variants of uncertain significance, or inconclusive variants identified during this analysis may be reported in certain circumstances. Our laboratory's variant classification criteria are based on the ACMG and internal guidelines and our current understanding of the specific genes. This interpretation may change over time as more information about a gene and/or variant becomes available. Natera and its lab partner(s) may reclassify variants at certain intervals but may not release updated reports without a specific request made to Natera by the ordering provider. Natera may disclose incidental findings if deemed clinically pertinent to the test performed.

Negative Results

A negative carrier screening result reduces the risk for a patient to be a carrier of a specific disease but does not completely rule out carrier status. Please visit https://www.natera.com/panel-option/h-all/ for a table of carrier rates, detection rates, residual risks and promised variants/exons per gene. Carrier rates before and after testing vary by ethnicity and assume a negative family history for each disease screened and the absence of clinical symptoms in the patient. Any patient with a family history for a specific genetic disease will have a higher carrier risk prior to testing and, if the disease-causing mutation in their family is not included on the test, their carrier risk would remain unchanged. Genetic counseling is recommended for patients with a family history of genetic disease so that risk figures based on actual family history can be determined and discussed along with potential implications for reproduction. Horizon carrier screening has been developed to identify the reproductive risks for monogenic inherited conditions. Even when one or both members of a couple screen negative for pathogenic variants in a specific gene, the disease risk for their offspring is not zero. There is still a low risk for the condition in their offspring due to a number of different mechanisms that are not detected by Horizon including, but not limited to, pathogenic variant(s) in the tested gene or in a different gene not included on Horizon, pathogenic variant(s) in an upstream regulator, uniparental disomy, de novo mutation(s), or digenic or polygenic inheritance.

Additional Comments

These analyses generally provide highly accurate information regarding the patient's carrier status. Despite this high level of accuracy, it should be kept in mind that there are many potential sources of diagnostic error, including misidentification of samples, polymorphisms, or other rare genetic variants that interfere with analysis. Families should understand that rare diagnostic errors may occur for these reasons.



Nichols Institute, Chantilly

SPECIMEN INFORMATION

SPECIMEN: REQUISITION: LAB REF NO:

COLLECTED: 07/10/2024 RECEIVED: 07/11/2024 REPORTED: 07/22/2024

07/10/2024 15:00 07/11/2024 23:52 07/22/2024 18:16 PATIENT INFORMATION DONOR, 7628

DOB: Age:

ID: 7628-PHONE: REPORT STATUS Final

ORDERING PHYSICIAN

CLIENT INFORMATION

Test Name	In Range	Out of Range	Reference Range	Lab
Hemoglobinopathy Evaluation				AMD
Red Blood Cell Count HEMOGLOBIN Hematocrit		6.05 Н 17.3 Н	4.20-5.80 Mill/uL 13.2-17.1 g/dL	
Hematocrit		56.0 H	38.5-50.0 %	
MCV	92.6		80.0-100.0 fL	
MCH	28.6		27.0-33.0 pg	
RDW	12.0		11.0-15.0 %	
Hemoglobin A	97.1		>96.0 %	
Hemoglobin F	0.0		<2.0 %	
Hemoglobin A2 (Quant) Interpretation	2.9		2.0-3.2 %	

NORMAL PHENOTYPE

There are no variant peaks identified. The patient's hemogram shows elevated hemoglobin/hematocrit. It should be pointed out that elevated hemoglobin/hematocrit can be caused by a variety of conditions including high oxygen affinity hemoglobins which may not be detected by high-performance liquid chromatography (HPLC) and/or capillary zone electrophoresis (CZE). Rare variants hemoglobins have no separation from hemoglobin A by capillary zone electrophoresis (CZE) or high-performance liquid chromatography (HPLC).

If clinically indicated, Thalassemia and Hemoglobinopathy Comprehensive (TC 17365) should be

Results were reviewed and interpreted by

Edward C.C. Wong. M.D.

If physician or health provider needs additional information, please call 1-703-802-7256.





PATIENT INFORMATION DONOR, 7628

REPORT STATUS Final

Nichols Institute, Chantilly

REPORTED:

DOB: SEX: M Age:

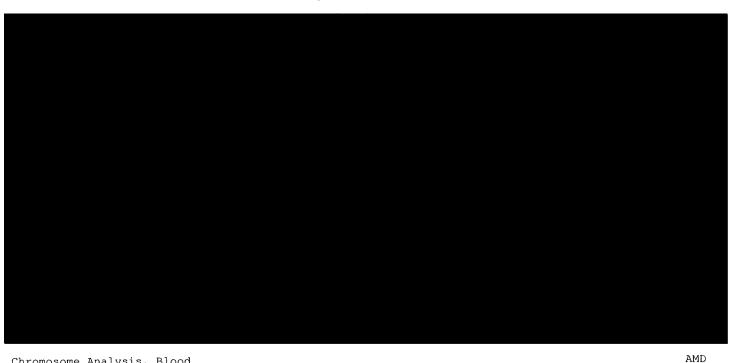
ORDERING PHYSICIAN

COLLECTED: 07/10/2024 15:00

07/10/2024 15:00 07/22/2024 18:16

ID: 7628-

Test Name In Range Out of Range Reference Range Lab



Chromosome Analysis, Blood Chromosome Analysis, Blood Chromosome Analysis, Blood

Order ID:

Specimen Type:

Blood

Clinical Indication:

Gamete donor

RESULT:

NORMAL MALE KARYOTYPE

INTERPRETATION:

Chromosome analysis revealed normal G-band patterns within the limits of standard cytogenetic analysis.

Please expect the results of any other concurrent study in a separate report.

NOMENCLATURE:

46,XY

ASSAY INFORMATION:

Method:

G-Band (Digital Analysis:

MetaSystems/Ikaros)

Cells Counted:
Band Level:
Cells Analyzed:

20 550 PATIENT INFORMATION DONOR, 7628

Final REPORT STATUS

Nichols Institute, Chantilly

Test Name

ORDERING PHYSICIAN

DOB:

COLLECTED: 07/10/2024 15:00 18:16 REPORTED: 07/22/2024

SEX: M

In Range

ID: 7628-

Age:

Out of Range

Reference Range

Lab

Chromosome Analysis, Blood (Continued) Chromosome Analysis, Blood (Continued)

Cells Karyotyped:

This test does not address genetic disorders that cannot be detected by standard cytogenetic methods or rare events such as low level mosaicism or subtle rearrangements.

Christine A. Curtis, Ph.D., FACMG, Technical Director, Cytogenetics and Genomics, 703-802-7156, [site AMD10]

Electronic Signature:

7/22/2024 5:31 PM

For additional information, please refer to http://education.questdiagnostics.com/faq/chromsblood (This link is being provided for informational/ educational purposes only).

Performing Laboratory Information:

Quest Diagnostics Nichols Institute 14225 Newbrook Drive Chantilly VA 20151 Laboratory Director: Patrick W Mason, MD PhD





Patient Information: 7628, Donor

DOB: Sex: M MR#: 7628 Patient#:

Partner Information:

Not Tested

Accession: N/A

Physician: Wieloch, Shannon GC: Wieloch, Shannon Fairfax Cryobank 3015 Williams Drive #110 Fairfax, VA 22031

Phone:

Laboratory:

Fulgent Therapeutics LLC CAP#: 8042697 CLIA#: 05D2043189 Laboratory Director: Dr. Amar Jariwala

Report Date: Nov 16,2025

Accession

Specimen Type: DNA Collected: Not Provided

FINAL RESULTS

No carrier mutations identified

TEST PERFORMED

Single Gene Carrier Screening: SH3TC2

(1 Gene Panel: SH3TC2; gene sequencing with deletion and duplication analysis)

INTERPRETATION:

Notes and Recommendations:

- No carrier mutations were identified in the submitted specimen. A negative result does not rule out the possibility of a genetic predisposition nor does it rule out any pathogenic mutations in areas not assessed by this test or in regions that were covered at a level too low to reliably assess. Also, it does not rule out mutations that are of the sort not queried by this test; see Methods and Limitations for more information. A negative result reduces, but does not eliminate, the chance to be a carrier for any condition included in this screen. Please see the supplemental table for details.
- This carrier screening test does not screen for all possible genetic conditions, nor for all possible mutations in every gene tested. This report does not include variants of uncertain significance; only variants classified as pathogenic or likely pathogenic at the time of testing, and considered relevant for reproductive carrier screening, are reported. Please see the gene specific notes for details. Please note that the classification of variants can change over time.
- Patients may wish to discuss any carrier results with blood relatives, as there is an increased chance that they are also carriers. These results should be interpreted in the context of this individual's clinical findings, biochemical profile, and family history.
- Gene specific notes and limitations may be present. See below.
- Genetic counseling is recommended. Available genetic counselors and additional resources can be found at the National Society of Genetic Counselors (NSGC; https://www.nsgc.org)





GENES TESTED:

Custom Beacon Carrier Screening Panel - Gene

This analysis was run using the Custom Beacon Carrier Screening Panel gene list. 1 genes were tested with 100.0% of targets sequenced at >20x coverage. For more gene-specific information and assistance with residual risk calculation, see the SUPPLEMENTAL TABLE.

SH3TC2

METHODS:

Genomic DNA was isolated from the submitted specimen indicated above (if cellular material was submitted). DNA was barcoded, and enriched for the coding exons of targeted genes using hybrid capture technology. Prepared DNA libraries were then sequenced using a Next Generation Sequencing technology. Following alignment to the human genome reference sequence (assembly GRCh37), variants were detected in regions of at least 10x coverage. For this specimen, 100.00% and 100.00% of coding regions and splicing junctions of genes listed had been sequenced with coverage of at least 10x and 20x, respectively, by NGS or by Sanger sequencing. The remaining regions did not have 10x coverage, and were not evaluated. Variants were interpreted manually using locus specific databases, literature searches, and other molecular biological principles. To minimize false positive results, any variants that do not meet internal quality standards are confirmed by Sanger sequencing. Variants classified as pathogenic, likely pathogenic, or risk allele which are located in the coding regions and nearby intronic regions (+/- 20bp) of the genes listed above are reported. Variants outside these intervals may be reported but are typically not guaranteed. When a single pathogenic or likely pathogenic variant is identified in a clinically relevant gene with autosomal recessive inheritance, the laboratory will attempt to ensure 100% coverage of coding sequences either through NGS or Sanger sequencing technologies ("fill-in"). All genes listed were evaluated for large deletions and/or duplications. However, single exon deletions or duplications will not be detected in this assay, nor will copy number alterations in regions of genes with significant pseudogenes. Putative deletions or duplications are analyzed using Fulgent Germline proprietary pipeline for this specimen. Bioinformatics: The FPLMv2.0 pipeline was used to analyze this specimen.

LIMITATIONS:

General Limitations

These test results and variant interpretation are based on the proper identification of the submitted specimen, accuracy of any stated familial relationships, and use of the correct human reference sequences at the queried loci. In very rare instances, errors may result due to mix-up or co-mingling of specimens. Positive results do not imply that there are no other contributors, genetic or otherwise, to future pregnancies, and negative results do not rule out the genetic risk to a pregnancy. Official gene names change over time. Fulgent uses the most up to date gene names based on HUGO Gene Nomenclature Committee (https://www.genenames.org) recommendations. If the gene name on report does not match that of ordered gene, please contact the laboratory and details can be provided. Result interpretation is based on the available clinical and family history information for this individual, collected published information, and Alamut annotation available at the time of reporting. This assay is not designed or validated for the detection of low-level mosaicism or somatic mutations. This assay will not detect certain types of genomic aberrations such as translocations, inversions, or repeat expansions other than specified genes. DNA alterations in regulatory regions or deep intronic regions (greater than 20bp from an exon) may not be detected by this test. Unless otherwise indicated, no additional assays have been performed to evaluate genetic changes in this specimen. There are technical limitations on the ability of DNA sequencing to detect small insertions and deletions. Our laboratory uses a sensitive detection algorithm, however these types of alterations are not detected as reliably as single nucleotide variants. Rarely, due to systematic chemical, computational, or human error, DNA variants may be missed. Although next generation sequencing technologies and our bioinformatics analysis significantly reduce the confounding contribution of pseudogene sequences or other highly-homologous sequences, sometimes these may still interfere with the technical ability of the assay to identify pathogenic alterations in both sequencing and deletion/duplication analyses. Deletion/duplication analysis can identify alterations of genomic regions which include one whole gene (buccal swab specimens and whole blood specimens) and are two or more contiguous exons in size (whole blood specimens only); single exon deletions or duplications may occasionally be identified, but are not routinely detected by this test. When novel DNA duplications are identified, it is not possible to discern the genomic location or orientation of the duplicated segment, hence the effect of the duplication cannot be predicted. Where deletions are detected, it is not always possible to determine whether the predicted product will remain in-frame or not. Unless otherwise indicated, deletion/duplication analysis has not been performed in regions that have been sequenced by Sanger.

Patient: 7628, Donor; Sex: M;

; MR#: 7628 PAGE 2 of 4





Gene Specific Notes and Limitations

No gene specific limitations apply to the genes on the tested panel.

SIGNATURE:

Yan Meng, Ph.D., CGMB, FACMG on 11/16/2025

Laboratory Director, Fulgent

Janley

DISCLAIMER:

This test was developed, performed, and its performance characteristics determined by Fulgent Therapeutics LLC (CAP# 8042697, CLIA# 05D2043189), 4399 Santa Anita Ave., El Monte, CA 91731. It has not been cleared or approved by the FDA. The laboratory is regulated under CLIA as qualified to perform high-complexity testing. This test is used for clinical purposes. It should not be regarded as investigational or for research. Since genetic variation, as well as systematic and technical factors, can affect the accuracy of testing, the results of testing should always be interpreted in the context of clinical and familial data. For assistance with interpretation of these results, healthcare professionals may contact us directly at (626) 350-0537 or info@fulgentgenetics.com. It is recommended that patients receive appropriate genetic counseling to explain the implications of the test result, including its residual risks, uncertainties and reproductive or medical options.





To view the supplemental table describing the carrier frequencies, detection rates, and residual risks associated with the genes tested on any Beacon panel, please visit the following link:

Beacon Expanded Carrier Screening Supplemental Table







Patient Information:
7628, Donor
DOB:
Sex: M
MR#: 7628
Patient#:



Specimen Type: DNA Collected: Not provided Received Date: Nov 07,2025 Authorized Date: Nov 10,2025 Physician:
Wieloch, Shannon
GC: Wieloch, Shannon
Fairfax Cryobank
3015 Williams Drive #110
Fairfax, VA 22031
Phone:
Fax:

Laboratory:
Fulgent Therapeutics LLC
CAP#: 8042697
CLIA#: 05D2043189
Laboratory Director:
Dr. Amar Jariwala
Report Date: Nov 22,2025

Final Report

TEST PERFORMED

Custom NGS Panel - 2 Genes

(2 Gene Panel: MFN2 and VWF; gene sequencing with deletion and duplication analysis)

RESULTS:

No diagnostic sequence or copy-number variants were identified in the submitted specimen.

A negative result does not rule out the possibility of a genetic predisposition nor does it rule out any pathogenic mutations of the sort not queried by this test or in areas not reliably assessed by this test.

INTERPRETATION:

Notes and Recommendations:

- As requested, this report only includes variants classified as Pathogenic, Likely Pathogenic, or Risk Allele at the time of analysis. If detected, this report does not include variants classified as of uncertain significance.
- Gene specific notes and limitations may be present. See below.
- These results should be interpreted in the context of this individual's clinical findings, biochemical profile, and family history.
- Genetic counseling is recommended. Available genetic counselors and additional resources can be found at the National Society of Genetic Counselors (NSGC; https://www.nsgc.org)
- Guide to Interpreting Genomic Reports: A Genomics Toolkit (CSER Consortium; February 2017) (https://www.genome.gov/For-Health-Professionals/Provider-Genomics-Education-Resources#hep)

GENES TESTED:

Custom NGS Panel - 2 Genes

2 genes tested (98.52% at >20x).

MFN2, VWF

Gene Specific Notes and Limitations

<u>VWF:</u> Significant pseudogene interference in exons 23-34 of the VWF gene has been known to occur and may interfere with the technical ability of the assay to identify pathogenic alterations in both sequencing and deletion/duplication analyses.





METHODS:

Genomic DNA was isolated from the submitted specimen indicated above (if cellular material was submitted). DNA was barcoded, and enriched for the coding exons of targeted genes using hybrid capture technology. Prepared DNA libraries were then sequenced using a Next Generation Sequencing technology. Following alignment to the human genome reference sequence (assembly GRCh37), variants were detected in regions of at least 10x coverage. For this specimen, 99.62% and 98.52% of coding regions and splicing junctions of genes listed had been sequenced with coverage of at least 10x and 20x, respectively, by NGS or by Sanger sequencing. The remaining regions did not have 10x coverage, and were not evaluated. Variants were interpreted manually using locus specific databases, literature searches, and other molecular biological principles. To minimize false positive results, any variants that do not meet internal quality standards are confirmed by Sanger sequencing. Variants classified as pathogenic, likely pathogenic, or risk allele which are located in the coding regions and nearby intronic regions (+/- 20bp) of the genes listed above are reported. Variants outside these intervals may be reported but are typically not guaranteed. When a single pathogenic or likely pathogenic variant is identified in a clinically relevant gene with autosomal recessive inheritance, the laboratory will attempt to ensure 100% coverage of coding sequences either through NGS or Sanger sequencing technologies ("fill-in"). All genes listed were evaluated for large deletions and/or duplications. However, single exon deletions or duplications will not be detected in this assay, nor will copy number alterations in regions of genes with significant pseudogenes. Putative deletions or duplications identified by NGS are confirmed by an orthogonal method (gPCR or MLPA), unless exceeding an internally specified and validated quality score, beyond which deletions and duplications are considered real without further confirmation. Bioinformatics: The FPLMv2.0 pipeline was used to analyze this specimen.

LIMITATIONS:

These test results and variant interpretation are based on the proper identification of the submitted specimen, accuracy of any stated familial relationships, and use of the correct human reference sequences at the queried loci. In very rare instances, errors may result due to mix-up or co-mingling of specimens. Positive results do not imply that there are no other contributors, genetic or otherwise, to this individual's phenotype, and negative results do not rule out a genetic cause for the indication for testing. Official gene names change over time. Fulgent uses the most up to date gene names based on HUGO Gene Nomenclature Committee (https://www.genenames.org) recommendations. If the gene name on report does not match that of ordered gene, please contact the laboratory and details can be provided. Result interpretation is based on the available clinical and family history information for this individual, collected published information, and Alamut annotation available at the time of reporting. This assay is designed and validated for detection of germline variants only. It is not designed or validated for the detection of low-level mosaicism or somatic mutations. This assay will not detect certain types of genomic aberrations such as translocations, inversions, or repeat expansions (eg. trinucleotide or hexanucleotide repeat expansion). DNA alterations in regulatory regions or deep intronic regions (greater than 20bp from an exon) may not be detected by this test. Unless otherwise indicated, no additional assays have been performed to evaluate genetic changes in this specimen. There are technical limitations on the ability of DNA sequencing to detect small insertions and deletions. Our laboratory uses a sensitive detection algorithm for copy number variants, however these types of alterations are not detected as reliably as single nucleotide variants. Rarely, due to systematic chemical, computational, or human error, DNA variants may be missed. Although next generation sequencing technologies and our bioinformatics analysis significantly reduce the confounding contribution of pseudogene sequences or other highly-homologous sequences, sometimes these may still interfere with the technical ability of the assay to identify pathogenic alterations in both sequencing and deletion/duplication analyses. Deletion/duplication analysis can identify alterations of genomic regions which are two or more contiguous exons in size; single exon deletions or duplications may occasionally be identified, but are not routinely detected by this test. When novel DNA duplications are identified, it is not possible to discern the genomic location or orientation of the duplicated segment, hence the effect of the duplication cannot be predicted. Where deletions are detected, it is not always possible to determine whether the predicted product will remain in-frame or not. Unless otherwise indicated, deletion/duplication analysis has not been performed in regions that have been sequenced by Sanger.

SIGNATURE:

Zhenbin Chen, Ph.D., CGMB, FACMG on 11/22/2025

Laboratory Director, Fulgent





DISCLAIMER:

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Patient: 7628, Donor; Sex: M; ; MR#: 7628

PAGE 3 of 3