



## Donor 6565-PRS

### Genetic Testing Summary

Fairfax Cryobank recommends reviewing this genetic testing summary with your healthcare provider to determine suitability.

Last Updated: 08/27/18

Donor Reported Ancestry: Austrian, Peruvian, Spanish

Jewish Ancestry: No

| Genetic Test*                                   | Result   | Comments/Donor's Residual Risk**  |
|---|--|---|
| Chromosome analysis (karyotype)                 | Normal male karyotype                                    | No evidence of clinically significant chromosome abnormalities  |
| Hemoglobin evaluation                           | Normal hemoglobin fractionation and MCV/MCH results      | Reduced risk to be a carrier for sickle cell anemia, beta thalassemia, alpha thalassemia trait (aa/-- and a-/a-) and other hemoglobinopathies |
| Cystic Fibrosis (CF) carrier screening          | Negative by genotyping of 97 mutations- in the CFTR gene | 1/343   |
| Spinal Muscular Atrophy (SMA) carrier screening | Negative for deletions of exon 7 in the SMN1 gene        | 1/632   |
| Fragile X, PCR DNA Analysis                     | Normal Male  |   |

\*No single test can screen for all genetic disorders. A negative screening result significantly reduces, but cannot eliminate, the risk for these conditions in a pregnancy.

\*\*Donor residual risk is the chance the donor is still a carrier after testing negative.



# Cystic Fibrosis Mutation Analysis

Patient Name: Donor 6565,  
 Referring Physician: Madelyn Kahn, MD  
 Specimen #: [REDACTED]  
 Patient ID: [REDACTED]

Client #: [REDACTED]  
 Case #: [REDACTED]

Pacific Reproductive Services  
 444 De Haro Street  
 Suite 222  
 San Francisco CA 94107

DOB: [REDACTED] Date Collected: 05/28/2010  
 Sex: M Date Received: 06/01/2010  
 SSN: Lab ID:  
 Hospital ID:  
 Specimen Type: BLDPER

Ethnicity: Caucasian

Indication: Carrier test / Gamete donor

RESULTS: Negative for the 97 mutations analyzed

*6/8/10*

## INTERPRETATION

This individual's risk to be a carrier is reduced from 1/25 (4%) to 1/343 (0.3%), based on these results and a negative family history.

## COMMENTS:

| Mutation Detection Rates among Ethnic Groups |   | Detection rates are based on mutation frequencies in patients affected with cystic fibrosis. Among individuals with an atypical or mild presentation (e.g. congenital absence of the vas deferens, pancreatitis) detection rates may vary from those provided here. |  |
|--|---|---|--|
| Ethnicity                                    | Carrier risk reduction when no family history | Detection rate  | References   |
| African American                             | 1/65 to 1/336                                 | 81%   | Genet in Med 3:168, 2001   |
| Ashkenazi Jewish                             | 1/26 to 1/634                                 | 97%   | Am J Hum Genet 51:951, 1994  |
| Asian  |   | Not Provided  | Insufficient data  |
| Caucasian                                    | 1/25 to 1/343                                 | 93%   | Genet in Med 3:168, 2001; Genet in Med 4:90, 2002                      |
| Hispanic                                     | 1/46 to 1/205                                 | 78%   | Genet in Med 3:168, 2001; www.dhs.ca.gov/pcfh/gdb/html/PDE/CFStudy.htm |
| Jewish, non-Ashkenazi                        |   | Varies by country of origin   | Genet Testing 5:47, 2001, Genet Testing, 1:35, 1997                    |
| Other or Mixed Ethnicity                     |   | Not Provided  | Detection rate not determined and varies with ethnicity                |

This interpretation is based on the clinical and family relationship information provided and the current understanding of the molecular genetics of this condition.

## METHOD / LIMITATIONS:

DNA is isolated from the sample and tested for the 97 CF mutations listed. Regions of the *CFTR* gene are amplified enzymatically and subjected to a solution-phase multiplex allele-specific primer extension with subsequent hybridization to a bead array and fluorescence detection. Some mutations are then specifically identified by bi-directional dideoxysequencing. The assay discriminates between  $\Delta F508$  and the following polymorphisms: F508C, I506V and I507V. False positive or negative results may occur for reasons that include genetic variants, blood transfusions, bone marrow transplantation, erroneous representation of family relationships or contamination of a fetal sample with maternal cells.

Under the direction of:

*Ruth A Heim, PhD, FACMG*

Date: 06/08/2010

Ruth A. Heim, Ph.D., FACMG

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## MUTATIONS ANALYZED

|               |              |             |             |            |
|---------------|--------------|-------------|-------------|------------|
| ΔF311         | 3120+1G>A    | 712-1G>T    | Q359K/T360K | S549N      |
| ΔF508         | 3120G>A      | 935delA     | Q493X       | S549R T>G  |
| ΔI507         | 3171delC     | 936delTA    | Q552X       | T338I      |
| 1078delT      | 3199del6     | A455E       | Q890X       | V520F      |
| 1288insTA     | 3659delC     | A559T       | R1066C      | W1089X     |
| 1677delTA     | 3667del4     | C524X       | R1158X      | W1204X     |
| 1717-1G>A     | 3791delC     | CFTRdele2,3 | R1162X      | W1282X     |
| 1812-1G>A     | 3849+10kbC>T | D1152H      | R117C       | Y1092X C>A |
| 1898+1G>A     | 3876delA     | E60X        | R117H       | Y1092X C>G |
| 1898+5G>T     | 3905insT     | E92X        | R334W       | Y122X      |
| 1949del84     | 394delTT     | G178R       | R347H       |            |
| 2043delG      | 4016insT     | G330X       | R347P       |            |
| 2055del9>A    | 405+1G>A     | G480C       | R352Q       |            |
| 2105del13ins5 | 405+3A>C     | G542X       | R553X       |            |
| 2108delA      | 406-1G>A     | G551D       | R560T       |            |
| 2143delT      | 444delA      | G85E        | R709X       |            |
| 2183delAA>G   | 457TAT>G     | K710X       | R75X        |            |
| 2184delA      | 574delA      | L206W       | R764X       |            |
| 2184insA      | 621+1G>T     | M1101K      | S1196X      |            |
| 2307insA      | 663delT      | N1303K      | S1251N      |            |
| 2789+5G>A     | 711+1G>T     | P574H       | S1255X      |            |
| 2869insG      | 711+5G>A     | Q1238X      | S364P       |            |

The test was developed and its performance characteristics have been determined by Genzyme. The laboratory is regulated under the Clinical Laboratory Improvement Amendments of 1988 (CLIA) as qualified to perform high complexity clinical testing. This test must be used in conjunction with clinical assessment, when available.



June 8, 2010

Pacific Reproductive Svcs SF  
444 De Haro St Ste 222  
San Francisco, CA 94107

Test Results of: DONOR 6565, X  
DOB: [REDACTED] Age: [REDACTED] Sex: M  
Collected on: 05/28/2010  
Received on: 05/29/2010  
Reported on: 06/08/2010

Branch Number: [REDACTED]  
Account Number: [REDACTED]  
Specimen Number: [REDACTED]  
Specimen Type: Blood

Patient ID#:

Physician:

**Test: Fragile X, PCR DNA Analysis**

**Result:**

**NORMAL, Male**  
31 CGG repeats identified / CP 6/25/10

**Interpretation:**

DNA studies by PCR analysis identified one allele. These results do not provide evidence of the common CGG repeat expansion observed in patients with fragile X syndrome. Routine chromosome analysis is recommended in the diagnostic work-up for other causes of mental retardation. Due to the nature of the assay, small variations in reported repeat number may exist within and between laboratories.

Fragile X syndrome is one of the most common causes of inherited mental retardation. Some individuals with fragile X have characteristic physical features and behaviors. There can be wide variability in phenotypic expression. Fragile X is most often caused by an expansion in the number of the CGG repeats in the fragile X gene (*FMR1*). People with fewer than 45 CGG repeats have alleles within the normal range. People with 45-54 repeats are considered normal but have alleles in the grey zone. Some increases and decreases in repeat number can occur in offspring of individuals with grey zone alleles, but the chance is small that grey zone alleles would expand to a full mutation in the next generation. Those with 55-200 repeats have alleles in the premutation range. These individuals are not expected to have fragile X, but are at increased risk to have children with fragile X syndrome. Individuals with more than 200 repeats have full mutations and are expected to be clinically affected. Exceptions can occur as there are rare forms of fragile X mental retardation protein (FMRP) deficiency not caused by CGG expansion, which may not be detected by this analysis.

**Methodology:**

DNA analysis of the *FMR1* gene was performed by PCR amplification followed by agarose gel, as well as capillary electrophoresis. Southern blot analysis was not indicated due to the presence of one normal allele by PCR. The detection rate of this test is >99% for the common fragile X expansion (*FRAXA*). This test does not examine the *FRAXE* expansion. Molecular-based testing is highly accurate, but as in any laboratory test, rare diagnostic errors may occur. All test results must be combined with clinical information for the most accurate interpretation.

This test was developed and its performance characteristics determined by Laboratory Corporation of America Holdings (LabCorp). It has not been cleared or approved by the U.S. Food and Drug Administration.

The FDA has determined that such clearance or approval is not necessary. This test is used for clinical purposes. It should not be regarded as investigational or research.

**References:**

1. Park V, Howard-Peebles P, Sherman S, Taylor A, and Wulfsberg E. (1994). Am J Med Genet 53:380-381.
2. Maddalena A, et al. (2001). Genet Med 3:200-205.
3. Jacquemont S, Hagerman RJ, Lechey MA, Hall DA, Levine RA, Brunberg JA, Zhang L, Jardini T, Gane LW, Harris SW, Herman K, Grigsby J, Greco CM, Berry-Kravis E, Tassone F, and Hagerman PJ. (2004) J Amer Med Assoc 291:460-469.
4. Hagerman PJ and Hagerman RJ. (2004) Am J Hum Genet 74:805-816.

**Results Released By:** Kenneth J. Friedman, Ph.D., Director  
**Report Released By:** Dagny M. Patton, MS, CGC, Genetic Counselor

Arundhati Chatterjee, M.D.  
Medical Director

LabCorp  
1912 Alexander Drive, RTP, NC, 27709 (800) 345-GENE

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# SMN1 Copy Number Analysis

Patient Name: X Donor 6565

DOB: [REDACTED]

Age: [REDACTED]

SSN #: [REDACTED]

Gender: Male

163934 / 163935

Lab Corp

13112 Evening Creek Drive - South

San Diego, CA 92128

Genzyme Specimen #: [REDACTED]

Case #: [REDACTED]

Patient ID #: [REDACTED]

Date Collected: 05/28/2010

Date Received: 06/01/2010

Referring Physician: Madelyn Kahn

Client Lab ID #:

Genetic Counselor:

Hospital ID #:

Specimen Type: Peripheral Blood

Specimen ID #:

Specimen(s) Received: 1 - Lavender 10 ml round bottom tube(s)

Clinical Data: Carrier Test/Screening

Ethnicity: Not Provided

**RESULTS: SMN1 copy number: 2 (Reduced Carrier Risk)**

### INTERPRETATION:

This individual has an SMN1 copy number of two. This result reduces but does not eliminate the risk to be a carrier of SMA. Ethnic specific risk reductions based on a negative family history and an SMN1 copy number of two are provided in the Comments section of this report.

### COMMENT:

Spinal muscular atrophy (SMA) is an autosomal recessive disease of variable age of onset and severity caused by mutations (most often deletions or gene conversions) in the survival motor neuron (SMN1) gene. Molecular testing assesses the number of copies of the SMN1 gene. Individuals with one copy of the SMN1 gene are predicted to be carriers of SMA. Individuals with two or more copies have a reduced risk to be carriers. (Affected individuals have 0 copies of the SMN1 gene.)

This copy number analysis cannot detect individuals who are carriers of SMA as a result of either 2 (or very rarely 3) copies of the SMN1 gene on one chromosome and the absence of the SMN1 gene on the other chromosome or small intragenic mutations within the SMN1 gene. This analysis also will not detect germline mosaicism or mutations in genes other than SMN1. Additionally, de novo mutations have been reported in approximately 2% of SMA patients.

**Carrier Frequency and Risk Reductions for Individuals with No Family History of SMA**

| Ethnicity         | Detection Rate <sup>1</sup>  | A priori Carrier Risk <sup>1</sup> | Reduced Carrier Risk for 2 copy result | Reduced Carrier Risk for 3 copy result |
|-------------------|--|------------------------------------|--|--|
| Caucasian         | 94.9%  | 1:35                               | 1:632                                  | 1:3,500                                |
| Ashkenazi Jewish  | 90.2%  | 1:41                               | 1:350                                  | 1:4,000                                |
| Asian             | 92.6%  | 1:53                               | 1:628                                  | 1:5,000                                |
| Hispanic          | 90.6%  | 1:117                              | 1:1061                                 | 1:11,000                               |
| African American  | 71.1%  | 1:66                               | 1:121                                  | 1:3,000                                |
| Mixed Ethnicities | For counseling purposes, consider using the ethnic background with the most conservative risk estimates. |                                    |  |  |

### METHOD/LIMITATIONS:

Specimen DNA is isolated and amplified by real-time polymerase chain reaction (PCR) for exon 7 of the SMN1 gene and two reference genes. A mathematical algorithm is used to calculate the number of copies of SMN1. Sequencing of the primer and probe binding sites for the SMN1 real-time PCR reaction is performed on all fetal samples, and on samples from individuals with 1 copy of SMN1 on carrier testing, to rule out the presence of sequence variants which could interfere with analysis and interpretation. False positive or negative results may occur for reasons that include genetic variants, blood transfusions, bone marrow transplantation, erroneous representation of family relationships or contamination of a fetal sample with maternal cells.

### REFERENCES:

- Carrier frequency and detection rate are calculated based on analysis of allele frequencies among > 1000 individuals from each ethnic group noted (Genzyme Genetics, data submitted for publication).
- Online review of SMA: <http://www.genereviews.org/profiles/sma>

The test was developed and its performance characteristics have been determined by Genzyme. The laboratory is regulated under the Clinical Laboratory Improvement Amendments of 1988 (CLIA) as qualified to perform high complexity clinical testing. This test must be used in conjunction with clinical assessment, when available.

Electronically Signed by: Lynne S. Rosenblum, Ph.D., FACMG, on 06/04/2010

Reported by: /

Testing performed at Genzyme Genetics 3400 Computer Drive, Westborough, MA 01581 1-800-255-7357



Pacific Reproductive Svcs SF

444 De Haro St Ste 222  
 San Francisco, CA 94107  
 Ph: (415)487-2288  
 Fax: (415) 863-4358 CAB-60

**LCLS Specimen Number:** [REDACTED]  
**Patient Name:** DONOR 6565, X  
**Date of Birth:** [REDACTED]  
**Gender:** M  
**Patient ID:** [REDACTED]  
**Lab Number:** [REDACTED]  
**Indications:** DONOR

**Account Number:** [REDACTED]  
**Ordering Physician:**  
**Specimen Type:** BLOOD  
**Date Collected:** 05/28/2010  
**Date Received:** 05/29/2010  
**CoPath Number:**  
**Client Reference:**

**Test:** Chromosome, Blood

**Date Reported:** 06/15/2010

Cells Counted: 15  
 Cells Analyzed: 5

Cells Karyotyped: 2  
 Band Resolution: 650

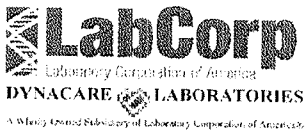
**CYTOGENETIC RESULT:** 46,XY

**INTERPRETATION: NORMAL MALE KARYOTYPE**

Cytogenetic analysis of PHA stimulated cultures has revealed a MALE karyotype with an apparently normal GTG banding pattern in all cells observed.

This result does not exclude the possibility of subtle rearrangements below the resolution of cytogenetics or congenital anomalies due to other etiologies.

Molecular results of fragile X testing will be reported separately.

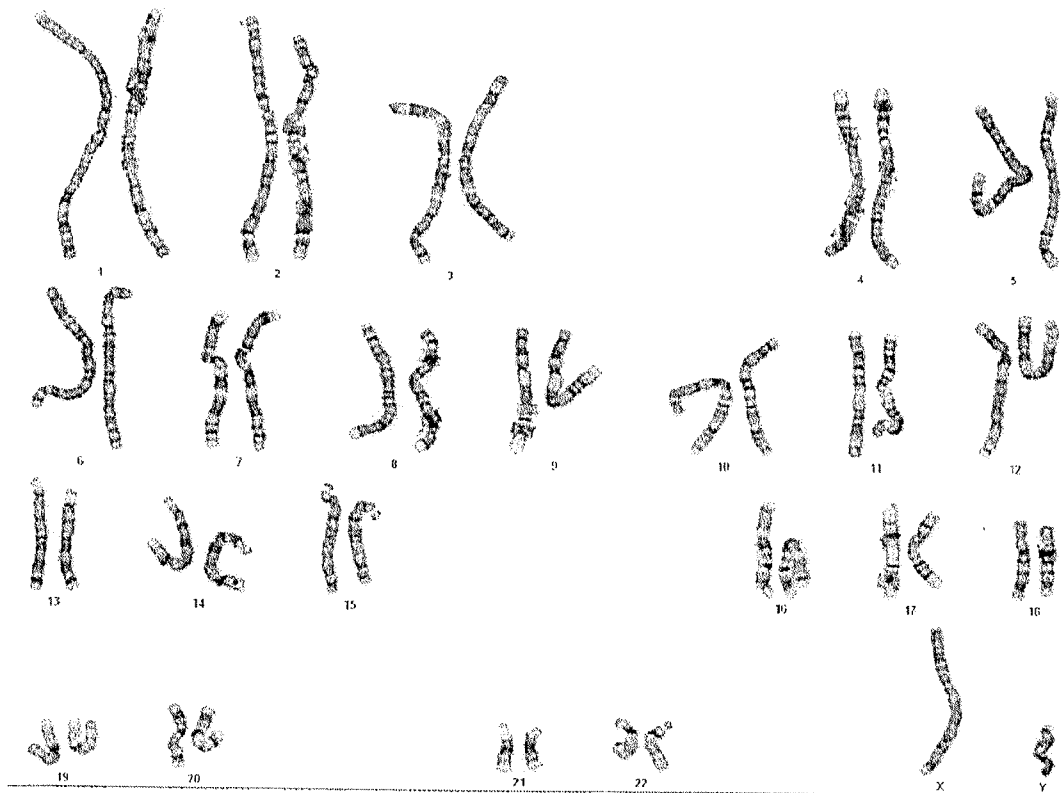


Pacific Reproductive Svcs SF

444 De Haro St Ste 222  
San Francisco, CA 94107  
Ph: (415)487-2288  
Fax: (415) 863-4358 CAB-60

LCLS Specimen Number: [REDACTED]  
Patient Name: DONOR 6565, X  
Date of Birth: [REDACTED]  
Gender: M  
Patient ID: [REDACTED]  
Lab Number: [REDACTED]  
Indications: DONOR

Account Number: [REDACTED]  
Ordering Physician:  
Specimen Type: BLOOD  
Date Collected: 05/28/2010  
Date Received: 05/29/2010  
CoPath Number:  
Client Reference:



Frederick W. Luthardt PhD, FACMG  
Board Certified Cytogeneticist

David Corwin, M.D.  
Medical Director  
Peter Papenhausen, PhD  
National Director of Cytogenetics

Test Site: Dynacare Laboratories  
550 17th Ave. Suite 200, SEATTLE, WA, 98122-5789 (800) 676-8033

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LabCorp Kansas City  
1706 N Corrington Avenue  
Kansas City, MO 64120

Phone: 800-457-1177

|                         |  |                     |  |                              |                |                      |       |
|-------------------------|--|---------------------|--|------------------------------|----------------|----------------------|-------|
| Specimen Number         |  | Patient ID          |  | Control Number               | Account Number | Account Phone Number | Route |
| DONOR 6565              |  | Patient Last Name   |  | Pacific Reproductive Svcs SF |                |                      |       |
| Patient First Name      |  | Patient Middle Name |  | 444 De Haro St Ste 222       |                |                      |       |
| DONOR 6565              |  | Patient SS#         |  | Patient Phone                |                | Total Volume         |       |
| Age (Y/M/D)             |  | Date of Birth       |  | Sex                          | Fasting        |                      |       |
|                         |  |                     |  | M                            |                |                      |       |
| Patient Address         |  |                     |  | Additional Information       |                |                      |       |
| Date and Time Collected |  | Date Entered        |  | Date and Time Reported       |                | Physician Name       |       |
| 04/22/10 15:30          |  | 04/23/10            |  | 04/26/10 15:14ET             |                | NPI                  |       |
|                         |  |                     |  |                              |                | Physician ID         |       |

Tests Ordered  
CBC With Differential/Platelet; Hgb Frac. w/o Solubility

| TESTS                                 | RESULT   | FLAG       | UNITS       | REFERENCE INTERVAL | LAB |
|---------------------------------------|----------|------------|-------------|--------------------|-----|
| <b>CBC With Differential/Platelet</b> |          |            |             |                    |     |
| WBC                                   | 5.7      |            | x10E3/uL    | 4.0 - 10.5         | 01  |
| RBC                                   | 4.69     |            | x10E6/uL    | 4.10 - 5.60        | 01  |
| Hemoglobin                            | 15.0     |            | g/dL        | 12.5 - 17.0        | 01  |
| Hematocrit                            | 43.0     |            | %           | 36.0 - 50.0        | 01  |
| MCV                                   | 92       |            | fL          | 80 - 98            | 01  |
| MCH                                   | 32.0     |            | pg          | 27.0 - 34.0        | 01  |
| MCHC                                  | 34.9     |            | g/dL        | 32.0 - 36.0        | 01  |
| RDW                                   | 13.1     |            | %           | 11.7 - 15.0        | 01  |
| Platelets                             | 197      |            | x10E3/uL    | 140 - 415          | 01  |
| Neutrophils                           | 66       |            | %           | 40 - 74            | 01  |
| Lymphs                                | 29       |            | %           | 14 - 46            | 01  |
| <b>Monocytes</b>                      | <b>3</b> | <b>Low</b> | %           | 4 - 13             | 01  |
| Eos                                   | 2        |            | %           | 0 - 7              | 01  |
| Basos                                 | 0        |            | %           | 0 - 3              | 01  |
| Neutrophils (Absolute)                | 3.8      |            | x10E3/uL    | 1.8 - 7.8          | 01  |
| Lymphs (Absolute)                     | 1.7      |            | x10E3/uL    | 0.7 - 4.5          | 01  |
| Monocytes (Absolute)                  | 0.2      |            | x10E3/uL    | 0.1 - 1.0          | 01  |
| Eos (Absolute)                        | 0.1      |            | x10E3/uL    | 0.0 - 0.4          | 01  |
| Baso (Absolute)                       | 0.0      |            | x10E3/uL    | 0.0 - 0.2          | 01  |
| <b>Hgb Frac. w/o Solubility</b>       |          |            |             |                    |     |
| Hgb A                                 | 97.7     |            | %           | 94.0 - 98.0        | 02  |
| Hgb S                                 | 0.0      |            | %           | 0.0                | 02  |
| Hgb C                                 | 0.0      |            | %           | 0.0                | 02  |
| Hgb A2                                | 2.3      |            | %           | 0.7 - 3.1          | 02  |
| Hgb F                                 | 0.0      |            | %           |                    | 02  |
|                                       |          |            | 1 day       | 69.7 - 84.3        |     |
|                                       |          |            | 5 days      | 71.0 - 82.6        |     |
|                                       |          |            | 3 weeks     | 62.7 - 77.3        |     |
|                                       |          |            | 6- 9 weeks  | 41.9 - 63.9        |     |
|                                       |          |            | 3- 4 months | 7.2 - 39.2         |     |
|                                       |          |            | 6 months    | 2.5 - 6.9          |     |
|                                       |          |            | 8-12 months | 0.6 - 2.6          |     |
|                                       |          |            | > 1 year    | 0.0 - 2.0          |     |

DONOR 6565, DONOR 6565 Seq # 2011

FINAL REPORT





LabCorp Kansas City  
1706 N Corrington Avenue  
Kansas City, MO 64120

Phone: 800-457-1177

|                        |            |                |                         |               |              |            |               |                 |  |
|------------------------|------------|----------------|-------------------------|---------------|--------------|------------|---------------|-----------------|--|
| DONOR 6565, DONOR 6565 |            |                |                         |               | Patient Name |            |               | Specimen Number |  |
| Account Number         | Patient ID | Control Number | Date and Time Collected | Date Reported | Sex          | Age(Y/M/D) | Date of Birth |                 |  |
|                        |            | 60111024737    | 04/22/10 15:30          | 04/26/10      | M            |            |               |                 |  |

| TESTS | RESULT | FLAG | UNITS | REFERENCE INTERVAL | LAB |
|-------|--------|------|-------|--------------------|-----|
|-------|--------|------|-------|--------------------|-----|

Interpretation

Normal adult hemoglobin present. *CP 4/29/10*

02

|  |    |  |                            |
|--|----|--|----------------------------|
| 01   | KC | LabCorp Kansas City<br>1706 N Corrington Avenue, Kansas City, MO 64120 | Dir: Dugald Taylor, MD     |
| 02   | BN | LabCorp Burlington<br>1447 York Court, Burlington, NC 27215-3361       | Dir: William F Hancock, MD |
| For inquiries, the physician may contact <b>Branch: 800-888-1113 Lab: 800-457-1177</b> |    |  |                            |

|                        |  |  |            |
|------------------------|--|--|------------|
| DONOR 6565, DONOR 6565 |  |  | Seq # 2011 |
|------------------------|--|--|------------|

FINAL REPORT