# Fairfax <br> Cr\%obank 

The Trusted Choice for Donor Sperm

## Donor 6427

## Genetic Testing Summary

> Fairfax Cryobank recommends reviewing this genetic testing summary with your healthcare provider to determine suitability.

Last Updated: 01/04/22

Donor Reported Ancestry: English, Irish, Norwegian Jewish Ancestry: No

| Genetic Test* | Result | Comments/Donor's Residual <br> Risk** |
| :--- | :--- | :--- |


| Chromosome analysis (karyotype) | Normal male karyotype | No evidence of clinically significant <br> chromosome abnormalities |
| :--- | :--- | :--- |
| Hemoglobin evaluation | Normal hemoglobin fractionation and <br> MCV/MCH results | Reduced risk to be a carrier for sickle <br> cell anemia, beta thalassemia, alpha <br> thalassemia trait (aa/-- and a-/a-) and <br> other hemoglobinopathies |
| Cystic Fibrosis (CF) carrier screening | Negative by gene sequencing in the <br> CFTR gene | $1 / 440$ |
| Spinal Muscular Atrophy (SMA) carrier <br> screening | Negative for deletions of exon 7 in the <br> SMN1 gene | $1 / 894$ |
| Expanded Genetic Disease Carrier |  |  |
| Screening Panel attached- 283 diseases |  |  |
| by gene sequencing | Carrier: Abetalipoproteinemia (MTTP) | Partner testing recommended before <br> using this donor. |

[^0]Carrier screening report
Donor 6427
Date of Birth:
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## Patient Information

Name: Donor 6427


Sema4 ID:
Client ID


Indication: Carrier Testing

## Specimen Information

Specimen Type: Blood
Date Collected: 05/05/2021
Date Received: 05/06/2021
Final Report: 05/22/2021

## Expanded Carrier Screen (283) Minus TSE

Number of genes tested: 283

## SUMMARY OF RESULTS AND RECOMMENDATIONS

| $\oplus$ Positive | $\ominus$ Negative |
| :---: | :---: |
| Carrier of Abetalipoproteinemia (AR) | Negative for all other genes tested |
| Associated gene(s): MTTP | To view a full list of genes and diseases tested |
| please see Table 1 in this report |  |
| Variant(s) Detected: c.2636_2637delAA, p.K879SfsX9, Likely |  |
| Pathogenic, Heterozygous (one copy) |  |

$A R=$ Autosomal recessive; $X L=X$-linked

## Recommendations

- Testing the partner for the above positive disorder(s) and genetic counseling are recommended.
- Please note that for female carriers of X-linked diseases, follow-up testing of a male partner is not indicated.
- CGG repeat analysis of FMR1 for fragile $X$ syndrome is not performed on males as repeat expansion of premutation alleles is not expected in the male germline.
- Individuals of Asian, African, Hispanic and Mediterranean ancestry should also be screened for hemoglobinopathies by CBC and hemoglobin electrophoresis.
- Consideration of residual risk by ethnicity after a negative carrier screen is recommended for the other diseases on the panel, especially in the case of a positive family history for a specific disorder.


## Interpretation of positive results

## Abetalipoproteinemia (AR)

## Results and Interpretation

A heterozygous (one copy) likely pathogenic frameshift variant, c.2636_2637delAA, p.K879SfsX9, was detected in the MTTPgene (NM_000253.3). When this variant is present in trans with a pathogenic variant, it is considered to be causative for abetalipoproteinemia. Therefore, this individual is expected to be at least a carrier for abetalipoproteinemia. Heterozygous carriers are not expected to exhibit symptoms of this disease.

## What is Abetalipoproteinemia?

Abetalipoproteinemia is an autosomal recessive disease caused by pathogenic variants in the MTTPgene and has the highest prevalence in the Ashkenazi Jewish population. Abetalipoproteinemia results from impaired absorption of dietary fats, cholesterol and fat-soluble vitamins. Clinically, this disease can cause failure to thrive, diarrhea, fatty stools, and abnormally shaped red blood cells. The resulting vitamin deficiency can also cause ataxia and retinitis pigmentosa in adulthood. Without treatments, life expectancy is significantly reduced, but with medical management patients may have a near-normal lifespan. No genotype-phenotype correlation has been reported.

## Test description

This patient was tested for a panel of diseases using a combination of sequencing, targeted genotyping and copy number analysis. Please note that negative results reduce but do not eliminate the possibility that this individual is a carrier for one or more of the disorders tested. Please see Table 1 for a list of genes and diseases tested, and go.sema4.com/residualrisk for specific detection rates and residual risk by ethnicity. With individuals of mixed ethnicity, it is recommended to use the highest residual risk estimate. Only variants determined to be pathogenic or likely pathogenic are reported in this carrier screening test.

## Hlice KTamer

Alice Tanner, Ph.D., M.S., CGC, FACMG, Laboratory Director
Laboratory Medical Consultant: George A. Diaz, M.D., Ph.D

## Genes and diseases tested

For specific detection rates and residual risk by ethnicity, please visit go.sema4.com/residualrisk

Table 1: List of genes and diseases tested with detailed results

| Disease | Gene | Inheritance Pattern | Status | Detailed Summary |
| :---: | :---: | :---: | :---: | :---: |
| $\oplus$ Positive |  |  |  |  |
| Abetalipoproteinemia | MTTP | AR | Carrier | c.2636_2637delAA, p.K879SfsX9, Likely Pathogenic, Heterozygous (one copy) |
| $\ominus$ Negative |  |  |  |  |
| 3-Beta-Hydroxysteroid Dehydrogenase Type II Deficiency | HSD3B2 | AR | Reduced Risk |  |
| 3-Methylcrotonyl-CoA Carboxylase Deficiency (MCCC1-Related) | MCCC1 | AR | Reduced Risk |  |
| 3-Methylcrotonyl-CoA Carboxylase Deficiency <br> (MCCC2-Related) | MCCC2 | AR | Reduced Risk |  |
| 3-MethyIglutaconic Aciduria, Type III | OPA3 | AR | Reduced Risk |  |
| 3-Phosphoglycerate Dehydrogenase Deficiency | PHGDH | AR | Reduced Risk |  |
| 6-Pyruvoyl-Tetrahydropterin Synthase Deficiency | PTS | AR | Reduced Risk |  |
| Achromatopsia (CNGB3-related) | CNGB3 | AR | Reduced Risk |  |
| Acrodermatitis Enteropathica | SLC39A4 | AR | Reduced Risk |  |
| Acute Infantile Liver Failure | TRMU | AR | Reduced Risk |  |
| Acyl-CoA Oxidase I Deficiency | ACOX1 | AR | Reduced Risk |  |
| Adenosine Deaminase Deficiency | ADA | AR | Reduced Risk |  |
| Adrenoleukodystrophy, X-Linked | ABCD1 | XL | Reduced Risk |  |
| Aicardi-Goutieres Syndrome (SAMHD1-Related) | SAMHD1 | AR | Reduced Risk |  |
| Alpha-Mannosidosis | MAN2B1 | AR | Reduced Risk |  |
| Alpha-Thalassemia | HBA1/HBA2 | AR | Reduced Risk | HBA1 Copy Number: 2 <br> HBA2 Copy Number: 2 <br> No pathogenic copy number variants detected HBA1/HBA2 Sequencing: Negative |
| Alpha-Thalassemia Mental Retardation Syndrome | ATRX | XL | Reduced Risk |  |
| Alport Syndrome (COL4A3-Related) | COL4A3 | AR | Reduced Risk |  |
| Alport Syndrome (COL4A4-Related) | COL4A4 | AR | Reduced Risk |  |
| Alport Syndrome (COL4A5-Related) | COL4A5 | XL | Reduced Risk |  |
| Alstrom Syndrome | ALMS1 | AR | Reduced Risk |  |
| Andermann Syndrome | SLC12A6 | AR | Reduced Risk |  |
| Argininosuccinic Aciduria | ASL | AR | Reduced Risk |  |
| Aromatase Deficiency | CYP19A1 | AR | Reduced Risk |  |
| Arthrogryposis, Mental Retardation, and Seizures | SLC35A3 | AR | Reduced Risk |  |
| Asparagine Synthetase Deficiency | ASNS | AR | Reduced Risk |  |
| Aspartylglycosaminuria | AGA | AR | Reduced Risk |  |
| Ataxia With Isolated Vitamin E Deficiency | TTPA | AR | Reduced Risk |  |
| Ataxia-Telangiectasia | ATM | AR | Reduced Risk |  |
| Autosomal Recessive Spastic Ataxia of CharlevoixSaguenay | SACS | AR | Reduced Risk |  |
| Bardet-Biedl Syndrome (BBS10-Related) | BBS10 | AR | Reduced Risk |  |
| Bardet-Biedl Syndrome (BBS12-Related) | BBS12 | AR | Reduced Risk |  |
| Bardet-Biedl Syndrome (BBS1-Related) | BBS1 | AR | Reduced Risk |  |
| Bardet-Biedl Syndrome (BBS2-Related) | BBS2 | AR | Reduced Risk |  |
| Bare Lymphocyte Syndrome, Type II | CIITA | AR | Reduced Risk |  |
| Bartter Syndrome, Type 4A | BSND | AR | Reduced Risk |  |
| Bernard-Soulier Syndrome, Type A1 | GP1BA | AR | Reduced Risk |  |
| Bernard-Soulier Syndrome, Type C | GP9 | AR | Reduced Risk |  |
| Beta-Globin-Related Hemoglobinopathies | HBB | AR | Reduced Risk |  |

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| Beta-Ketothiolase Deficiency | ACAT1 | AR | Reduced Risk |  |
| :---: | :---: | :---: | :---: | :---: |
| Bilateral Frontoparietal Polymicrogyria | GPR56 | AR | Reduced Risk |  |
| Biotinidase Deficiency | BTD | AR | Reduced Risk |  |
| Bloom Syndrome | BLM | AR | Reduced Risk |  |
| Canavan Disease | ASPA | AR | Reduced Risk |  |
| Carbamoylphosphate Synthetase I Deficiency | CPS1 | AR | Reduced Risk |  |
| Carnitine Palmitoyltransferase IA Deficiency | CPT1A | AR | Reduced Risk |  |
| Carnitine Palmitoyltransferase II Deficiency | CPT2 | AR | Reduced Risk |  |
| Carpenter Syndrome | RAB23 | AR | Reduced Risk |  |
| Cartilage-Hair Hypoplasia | RMRP | AR | Reduced Risk |  |
| Cerebral Creatine Deficiency Syndrome 1 | SLC6A8 | XL | Reduced Risk |  |
| Cerebral Creatine Deficiency Syndrome 2 | GAMT | AR | Reduced Risk |  |
| Cerebrotendinous Xanthomatosis | CYP27A1 | AR | Reduced Risk |  |
| Charcot-Marie-Tooth Disease, Type 4D | NDRG1 | AR | Reduced Risk |  |
| Charcot-Marie-Tooth Disease, Type 5 / Arts Syndrome | PRPS1 | XL | Reduced Risk |  |
| Charcot-Marie-Tooth Disease, X-Linked | GJB1 | XL | Reduced Risk |  |
| Choreoacanthocytosis | VPS13A | AR | Reduced Risk |  |
| Choroideremia | CHM | XL | Reduced Risk |  |
| Chronic Granulomatous Disease (CYBA-Related) | CYBA | AR | Reduced Risk |  |
| Chronic Granulomatous Disease ( $C Y B B$-Related) | СУВВ | XL | Reduced Risk |  |
| Citrin Deficiency | SLC25A13 | AR | Reduced Risk |  |
| Citrullinemia, Type 1 | ASS1 | AR | Reduced Risk |  |
| Cohen Syndrome | VPS13B | AR | Reduced Risk |  |
| Combined Malonic and Methylmalonic Aciduria | ACSF3 | AR | Reduced Risk |  |
| Combined Oxidative Phosphorylation Deficiency 1 | GFM1 | AR | Reduced Risk |  |
| Combined Oxidative Phosphorylation Deficiency 3 | TSFM | AR | Reduced Risk |  |
| Combined Pituitary Hormone Deficiency 2 | PROP1 | AR | Reduced Risk |  |
| Combined Pituitary Hormone Deficiency 3 | LHX3 | AR | Reduced Risk |  |
| Combined SAP Deficiency | PSAP | AR | Reduced Risk |  |
| Congenital Adrenal Hyperplasia due to 17-AlphaHydroxylase Deficiency | CYP17A1 | AR | Reduced Risk |  |
| Congenital Adrenal Hyperplasia due to 21-Hydroxylase Deficiency | CYP21A2 | AR | Reduced Risk | CYP21A2 copy number: 2 CYP21A2 sequencing: Negative |
| Congenital Amegakaryocytic Thrombocytopenia | MPL | AR | Reduced Risk |  |
| Congenital Disorder of Glycosylation, Type la | PMM2 | AR | Reduced Risk |  |
| Congenital Disorder of Glycosylation, Type Ib | MPI | AR | Reduced Risk |  |
| Congenital Disorder of Glycosylation, Type Ic | ALG6 | AR | Reduced Risk |  |
| Congenital Insensitivity to Pain with Anhidrosis | NTRK1 | AR | Reduced Risk |  |
| Congenital Myasthenic Syndrome (CHRNE-Related) | CHRNE | AR | Reduced Risk |  |
| Congenital Myasthenic Syndrome (RAPSN-Related) | RAPSN | AR | Reduced Risk |  |
| Congenital Neutropenia (HAX1-Related) | HAX1 | AR | Reduced Risk |  |
| Congenital Neutropenia (VPS45-Related) | VPS45 | AR | Reduced Risk |  |
| Corneal Dystrophy and Perceptive Deafness | SLC4A11 | AR | Reduced Risk |  |
| Corticosterone Methyloxidase Deficiency | CYP11B2 | AR | Reduced Risk |  |
| Cystic Fibrosis | CFTR | AR | Reduced Risk |  |
| Cystinosis | CTNS | AR | Reduced Risk |  |
| D-Bifunctional Protein Deficiency | HSD17B4 | AR | Reduced Risk |  |
| Deafness, Autosomal Recessive 77 | LOXHD1 | AR | Reduced Risk |  |
| Duchenne Muscular Dystrophy / Becker Muscular Dystrophy | DMD | XL | Reduced Risk |  |
| Dyskeratosis Congenita (RTEL1-Related) | RTEL1 | AR | Reduced Risk |  |
| Dystrophic Epidermolysis Bullosa | COL7A1 | AR | Reduced Risk |  |
| Ehlers-Danlos Syndrome, Type VIIC | ADAMTS2 | AR | Reduced Risk |  |
| Ellis-van Creveld Syndrome (EVC-Related) | EVC | AR | Reduced Risk |  |
| Emery-Dreifuss Myopathy 1 | EMD | XL | Reduced Risk |  |
| Enhanced S-Cone Syndrome | NR2E3 | AR | Reduced Risk |  |
| Ethylmalonic Encephalopathy | ETHE1 | AR | Reduced Risk |  |
| Fabry Disease | GLA | XL | Reduced Risk |  |
| Factor IX Deficiency | F9 | XL | Reduced Risk |  |
| Factor XI Deficiency | F11 | AR | Reduced Risk |  |
| Familial Autosomal Recessive Hypercholesterolemia | LDLRAP1 | AR | Reduced Risk |  |
| Familial Dysautonomia | IKBKAP | AR | Reduced Risk |  |

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| Familial Hypercholesterolemia | LDLR | AR | Reduced Risk |  |
| :---: | :---: | :---: | :---: | :---: |
| Familial Hyperinsulinism (ABCC8-Related) | ABCC8 | AR | Reduced Risk |  |
| Familial Hyperinsulinism (KCNJ11-Related) | KCNJ11 | AR | Reduced Risk |  |
| Familial Mediterranean Fever | MEFV | AR | Reduced Risk |  |
| Fanconi Anemia, Group A | FANCA | AR | Reduced Risk |  |
| Fanconi Anemia, Group C | FANCC | AR | Reduced Risk |  |
| Fanconi Anemia, Group G | FANCG | AR | Reduced Risk |  |
| Fragile XSyndrome | FMR1 | XL | Reduced Risk | FMR1 CGG repeat sizes: Not Performed <br> FMR1 Sequencing: Negative <br> Fragile X CGG triplet repeat expansion testing was not performed at this time, as the patient has either been previously tested or is a male. |
| Fumarase Deficiency | FH | AR | Reduced Risk |  |
| GRACILE Syndrome and Other BCS1L-Related Disorders | BCS1L | AR | Reduced Risk |  |
| Galactokinase Deficiency | GALK1 | AR | Reduced Risk |  |
| Galactosemia | GALT | AR | Reduced Risk |  |
| Gaucher Disease | GBA | AR | Reduced Risk |  |
| Gitelman Syndrome | SLC12A3 | AR | Reduced Risk |  |
| Glutaric Acidemia, Type I | GCDH | AR | Reduced Risk |  |
| Glutaric Acidemia, Type Ila | ETFA | AR | Reduced Risk |  |
| Glutaric Acidemia, Type Ilc | ETFDH | AR | Reduced Risk |  |
| Glycine Encephalopathy (AMT-Related) | AMT | AR | Reduced Risk |  |
| Glycine Encephalopathy (GLDC-Related) | GLDC | AR | Reduced Risk |  |
| Glycogen Storage Disease, Type II | GAA | AR | Reduced Risk |  |
| Glycogen Storage Disease, Type III | AGL | AR | Reduced Risk |  |
| Glycogen Storage Disease, Type IV / Adult <br> Polyglucosan Body Disease | GBE1 | AR | Reduced Risk |  |
| Glycogen Storage Disease, Type la | G6PC | AR | Reduced Risk |  |
| Glycogen Storage Disease, Type lb | SLC37A4 | AR | Reduced Risk |  |
| Glycogen Storage Disease, Type V | PYGM | AR | Reduced Risk |  |
| Glycogen Storage Disease, Type VII | PFKM | AR | Reduced Risk |  |
| HMG-CoA Lyase Deficiency | HMGCL | AR | Reduced Risk |  |
| Hemochromatosis, Type 2A | HFE2 | AR | Reduced Risk |  |
| Hemochromatosis, Type 3 | TFR2 | AR | Reduced Risk |  |
| Hereditary Fructose Intolerance | ALDOB | AR | Reduced Risk |  |
| Hereditary Spastic Paraparesis 49 | TECPR2 | AR | Reduced Risk |  |
| Hermansky-Pudlak Syndrome, Type 1 | HPS1 | AR | Reduced Risk |  |
| Hermansky-Pudlak Syndrome, Type 3 | HPS3 | AR | Reduced Risk |  |
| Holocarboxylase Synthetase Deficiency | HLCS | AR | Reduced Risk |  |
| Homocystinuria (CBS-Related) | CBS | AR | Reduced Risk |  |
| Homocystinuria due to MTHFR Deficiency | MTHFR | AR | Reduced Risk |  |
| Homocystinuria, cblE Type | MTRR | AR | Reduced Risk |  |
| Hydrolethalus Syndrome | HYLS1 | AR | Reduced Risk |  |
| Hyperornithinemia-HyperammonemiaHomocitrullinuria Syndrome | SLC25A15 | AR | Reduced Risk |  |
| Hypohidrotic Ectodermal Dysplasia 1 | EDA | XL | Reduced Risk |  |
| Hypophosphatasia | ALPL | AR | Reduced Risk |  |
| Inclusion Body Myopathy 2 | GNE | AR | Reduced Risk |  |
| Infantile Cerebral and Cerebellar Atrophy | MED17 | AR | Reduced Risk |  |
| Isovaleric Acidemia | IVD | AR | Reduced Risk |  |
| Joubert Syndrome 2 | TMEM216 | AR | Reduced Risk |  |
| Joubert Syndrome 7 / Meckel Syndrome 5 / COACH Syndrome | RPGRIP1L | AR | Reduced Risk |  |
| Junctional Epidermolysis Bullosa (LAMA3-Related) | LAMA3 | AR | Reduced Risk |  |
| Junctional Epidermolysis Bullosa (LAMB3-Related) | LAMB3 | AR | Reduced Risk |  |
| Junctional Epidermolysis Bullosa (LAMC2-Related) | LAMC2 | AR | Reduced Risk |  |
| Krabbe Disease | GALC | AR | Reduced Risk |  |
| Lamellar Ichthyosis, Type 1 | TGM1 | AR | Reduced Risk |  |
| Leber Congenital Amaurosis 10 and Other CEP290Related Ciliopathies | CEP290 | AR | Reduced Risk |  |
| Leber Congenital Amaurosis 13 | RDH12 | AR | Reduced Risk |  |

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| Leber Congenital Amaurosis 2 / Retinitis Pigmentosa 20 | RPE65 | AR | Reduced Risk |
| :---: | :---: | :---: | :---: |
| Leber Congenital Amaurosis 5 | LCA5 | AR | Reduced Risk |
| Leber Congenital Amaurosis 8 / Retinitis Pigmentosa 12 / Pigmented Paravenous Chorioretinal Atrophy | CRB1 | AR | Reduced Risk |
| Leigh Syndrome, French-Canadian Type | LRPPRC | AR | Reduced Risk |
| Lethal Congenital Contracture Syndrome 1 / Lethal Arthrogryposis with Anterior Horn Cell Disease | GLE1 | AR | Reduced Risk |
| Leukoencephalopathy with Vanishing White Matter | EIF2B5 | AR | Reduced Risk |
| Limb-Girdle Muscular Dystrophy, Type 2A | CAPN3 | AR | Reduced Risk |
| Limb-Girdle Muscular Dystrophy, Type 2B | DYSF | AR | Reduced Risk |
| Limb-Girdle Muscular Dystrophy, Type 2C | SGCG | AR | Reduced Risk |
| Limb-Girdle Muscular Dystrophy, Type 2D | SGCA | AR | Reduced Risk |
| Limb-Girdle Muscular Dystrophy, Type 2E | SGCB | AR | Reduced Risk |
| Limb-Girdle Muscular Dystrophy, Type 21 | FKRP | AR | Reduced Risk |
| Lipoamide Dehydrogenase Deficiency | DLD | AR | Reduced Risk |
| Lipoid Adrenal Hyperplasia | STAR | AR | Reduced Risk |
| Lipoprotein Lipase Deficiency | LPL | AR | Reduced Risk |
| Long-Chain 3-Hydroxyacyl-CoA Dehydrogenase Deficiency | HADHA | AR | Reduced Risk |
| Lysinuric Protein Intolerance | SLC7A7 | AR | Reduced Risk |
| Maple Syrup Urine Disease, Type 1a | BCKDHA | AR | Reduced Risk |
| Maple Syrup Urine Disease, Type 1b | BCKDHB | AR | Reduced Risk |
| Meckel Syndrome 1 / Bardet-Biedl Syndrome 13 | MKS1 | AR | Reduced Risk |
| Medium Chain Acyl-CoA Dehydrogenase Deficiency | ACADM | AR | Reduced Risk |
| Megalencephalic Leukoencephalopathy with Subcortical Cysts | MLC1 | AR | Reduced Risk |
| Menkes Disease | ATP7A | XL | Reduced Risk |
| Metachromatic Leukodystrophy | ARSA | AR | Reduced Risk |
| MethyImalonic Acidemia (MMAA-Related) | MMAA | AR | Reduced Risk |
| Methylmalonic Acidemia (MMAB-Related) | MMAB | AR | Reduced Risk |
| Methylmalonic Acidemia (MUT-Related) | MUT | AR | Reduced Risk |
| Methylmalonic Aciduria and Homocystinuria, Cobalamin C Type | MMACHC | AR | Reduced Risk |
| Methylmalonic Aciduria and Homocystinuria, Cobalamin D Type | MMADHC | AR | Reduced Risk |
| Microphthalmia / Anophthalmia | VSX2 | AR | Reduced Risk |
| Mitochondrial Complex I Deficiency (ACAD9-Related) | ACAD9 | AR | Reduced Risk |
| Mitochondrial Complex I Deficiency (NDUFAF5Related) | NDUFAF5 | AR | Reduced Risk |
| Mitochondrial Complex I Deficiency (NDUFS6-Related) | NDUFS6 | AR | Reduced Risk |
| Mitochondrial DNA Depletion Syndrome 6 / Navajo Neurohepatopathy | MPV17 | AR | Reduced Risk |
| Mitochondrial Myopathy and Sideroblastic Anemia 1 | PUS1 | AR | Reduced Risk |
| Mucolipidosis II / IIIA | GNPTAB | AR | Reduced Risk |
| Mucolipidosis III Gamma | GNPTG | AR | Reduced Risk |
| Mucolipidosis IV | MCOLN1 | AR | Reduced Risk |
| Mucopolysaccharidosis Type I | IDUA | AR | Reduced Risk |
| Mucopolysaccharidosis Type II | IDS | XL | Reduced Risk |
| Mucopolysaccharidosis Type IIIA | SGSH | AR | Reduced Risk |
| Mucopolysaccharidosis Type IIIB | NAGLU | AR | Reduced Risk |
| Mucopolysaccharidosis Type IIIC | HGSNAT | AR | Reduced Risk |
| Mucopolysaccharidosis Type IIID | GNS | AR | Reduced Risk |
| Mucopolysaccharidosis Type IVb / GM1 Gangliosidosis | GLB1 | AR | Reduced Risk |
| Mucopolysaccharidosis type IX | HYAL1 | AR | Reduced Risk |
| Mucopolysaccharidosis type VI | ARSB | AR | Reduced Risk |
| Multiple Sulfatase Deficiency | SUMF1 | AR | Reduced Risk |
| Muscle-Eye-Brain Disease and Other POMGNT1Related Congenital Muscular DystrophyDystroglycanopathies | POMGNT1 | AR | Reduced Risk |
| Myoneurogastrointestinal Encephalopathy | TYMP | AR | Reduced Risk |
| Myotubular Myopathy 1 | MTM1 | XL | Reduced Risk |
| N-Acetylglutamate Synthase Deficiency | NAGS | AR | Reduced Risk |
| Nemaline Myopathy 2 | NEB | AR | Reduced Risk |

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| Steel Syndrome | COL27A1 | AR | Reduced Risk |
| :---: | :---: | :---: | :---: |
| Stuve-Wiedemann Syndrome | LIFR | AR | Reduced Risk |
| Sulfate Transporter-Related Osteochondrodysplasia | SLC26A2 | AR | Reduced Risk |
| Tay-Sachs Disease | HEXA | AR | Reduced Risk |
| Tyrosinemia, Type I | FAH | AR | Reduced Risk |
| Usher Syndrome, Type IB | MYO7A | AR | Reduced Risk |
| Usher Syndrome, Type IC | USH1C | AR | Reduced Risk |
| Usher Syndrome, Type ID | CDH23 | AR | Reduced Risk |
| Usher Syndrome, Type IF | PCDH15 | AR | Reduced Risk |
| Usher Syndrome, Type IIA | USH2A | AR | Reduced Risk |
| Usher Syndrome, Type III | CLRN1 | AR | Reduced Risk |
| Very Long Chain Acyl-CoA Dehydrogenase Deficiency | ACADVL | AR | Reduced Risk |
| Walker-Warburg Syndrome and Other FKTN-Related Dystrophies | FKTN | AR | Reduced Risk |
| Wilson Disease | ATP7B | AR | Reduced Risk |
| Wolman Disease / Cholesteryl Ester Storage Disease | LIPA | AR | Reduced Risk |
| X-Linked Juvenile Retinoschisis | RS1 | XL | Reduced Risk |
| X-Linked Severe Combined Immunodeficiency | LL2RG | XL | Reduced Risk |
| Zellweger Syndrome Spectrum (PEX10-Related) | PEX10 | AR | Reduced Risk |
| Zellweger Syndrome Spectrum (PEX1-Related) | PEX1 | AR | Reduced Risk |
| Zellweger Syndrome Spectrum (PEX2-Related) | PEX2 | AR | Reduced Risk |
| Zellweger Syndrome Spectrum (PEX6-Related) | PEX6 | AR | Reduced Risk |

$A R=$ Autosomal recessive; $X L=X$-linked

## Test methods and comments

Genomic DNA isolated from this patient was analyzed by one or more of the following methodologies, as applicable:

## Fragile X CGG Repeat Analysis (Analytical Detection Rate >99\%)

PCR amplification using Asuragen, Inc. AmplideX ${ }^{\circledR} F M R 1$ PCR reagents followed by capillary electrophoresis for allele sizing was performed. Samples positive for FMR1 CGG repeats in the premutation and full mutation size range were further analyzed by Southern blot analysis to assess the size and methylation status of the FMR1 CGG repeat.

## Genotyping (Analytical Detection Rate >99\%)

Multiplex PCR amplification and allele specific primer extension analyses using the MassARRAY ${ }^{\circledR}$ System were used to identify certain recurrent variants that are complex in nature or are present in low copy repeats. Rare sequence variants may interfere with assay performance.

## Multiplex Ligation-Dependent Probe Amplification (MLPA) (Analytical Detection Rate >99\%)

MLPA ${ }^{\circledR}$ probe sets and reagents from MRC-Holland were used for copy number analysis of specific targets versus known control samples. False positive or negative results may occur due to rare sequence variants in target regions detected by MLPA probes. Analytical sensitivity and specificity of the MLPA method are both $99 \%$.

For alpha thalassemia, the copy numbers of the HBA1 and HBA2 genes were analyzed. Alpha-globin gene deletions, triplications, and the Constant Spring (CS) mutation are assessed. This test is expected to detect approximately $90 \%$ of all alpha-thalassemia mutations, varying by ethnicity. carriers of alpha-thalassemia with three or more HBA copies on one chromosome, and one or no copies on the other chromosome, may not be detected. With the exception of triplications, other benign alpha-globin gene polymorphisms will not be reported. Analyses of HBA1 and HBA2 are performed in association with long-range PCR of the coding regions followed by short-read sequencing.
For Duchenne muscular dystrophy, the copy numbers of all DMD exons were analyzed. Potentially pathogenic single exon deletions and duplications are confirmed by a second method. Analysis of $D M D$ is performed in association with sequencing of the coding regions.
For congenital adrenal hyperplasia, the copy number of the CYP21A2 gene was analyzed. This analysis can detect large deletions typically due to unequal meiotic crossing-over between CYP21A2 and the pseudogene CYP21A1P. Classic 30-kb deletions make up approximately $20 \%$ of CYP21A2 pathogenic alleles. This test may also identify certain point mutations in CYP21A2 caused by gene conversion events between CYP21A2 and CYP21A1P. Some carriers may not be identified by dosage sensitive methods as this testing cannot detect individuals with two copies (duplication) of the CYP21A2 gene on one chromosome and loss of CYP21A2 (deletion) on the other chromosome. Analysis of CYP21A2 is performed in association with long-range PCR of the coding regions followed by short-read sequencing.
For spinal muscular atrophy (SMA), the copy numbers of the SMN1 and SMN2 genes were analyzed. The individual dosage of exons 7 and 8 as well as the combined dosage of exons 1, 4, 6 and 8 of SMN1 and SMN2 were assessed. Copy number gains and losses can be detected with
this assay. Depending on ethnicity, 6-29\% of carriers will not be identified by dosage sensitive methods as this testing cannot detect individuals with two copies (duplication) of the SMN1 gene on one chromosome and loss of SMN1 (deletion) on the other chromosome (silent $2+0$ carrier) or individuals that carry an intragenic mutation in SMN1. Please also note that $2 \%$ of individuals diagnosed with SMA have a causative SMN1 variant that occurred de novo, and therefore cannot be picked up by carrier screening in the parents. Analysis of SMN1 is performed in association with short-read sequencing of exons $2 a-7$, followed by confirmation using long-range PCR (described below). The presence of the $c . * 3+80 T>G(c h r 5: 70,247,901 T>G)$ variant allele in an individual with Ashkenazi Jewish or Asian ancestry is typically indicative of a duplication of SMN1. When present in an Ashkenazi Jewish or Asian individual with two copies of SMN1, c. ${ }^{*} 3+80 T>G$ is likely indicative of a silent ( $2+0$ ) carrier. In individuals with two copies of SMN1 with African American, Hispanic or Caucasian ancestry, the presence or absence of $c . * 3+80 T>G$ significantly increases or decreases, respectively, the likelihood of being a silent $2+0$ silent carrier.

MLPA for Gaucher disease ( $G B A$ ), cystic fibrosis (CFTR), and non-syndromic hearing loss ( $G J B 2 / G J B 6$ ) will only be performed if indicated for confirmation of detected CNVs. If GBA analysis was performed, the copy numbers of exons $1,3,4$, and $6-10$ of the $G B A$ gene (of 11 exons total) were analyzed. If CFTR analysis was performed, the copy numbers of all 27 CFTR exons were analyzed. If GJB2/GJB6 analysis was performed, the copy number of the two GJB2 exons were analyzed, as well as the presence or absence of the two upstream deletions of the GJB2 regulatory region, del(GJB6-D13S1830) and del(GJB6-D13S1854).

## Next Generation Sequencing (NGS) (Analytical Detection Rate >95\%)

NGS was performed on a panel of genes for the purpose of identifying pathogenic or likely pathogenic variants.
Agilent SureSelect ${ }^{T M}$ XT Low Input technology was used with a custom capture library to target the exonic regions and intron/exon splice junctions of the relevant genes, as well as a number of UTR, intronic or promoter regions that contain previously reported mutations. Libraries were pooled and sequenced on the Illumina NovaSeq 9000 platform, using paired-end 100 bp reads. The sequencing data was analyzed using a custom bioinformatics algorithm designed and validated in house.
The coding exons and splice junctions of the known protein-coding RefSeq genes were assessed for the average depth of coverage (minimum of 20X) and data quality threshold values. Most exons not meeting a minimum of >20X read depth across the exon are further analyzed by Sanger sequencing. Please note that several genomic regions present difficulties in mapping or obtaining read depth >20X. These regions, which are described below, will not be reflexed to Sanger sequencing if the mapping quality or coverage is poor. Any variants identified during testing in these regions are confirmed by a second method and reported if determined to be pathogenic or likely pathogenic. However, as there is a possibility of false negative results within these regions, detection rates and residual risks for these genes have been calculated with the presumption that variants in these exons will not be detected, unless included in the MassARRAY ${ }^{\circledR}$ genotyping platform.
Exceptions: $A B C D 1$ (NM_000033.3) exons 8 and 9; $A D A\left(N M \_000022.2\right)$ exon 1; ADAMTS2 (NM_014244.4) exon 1; AGPS(NM_003659.3) chr2:178,257,512-178,257,649 (partial exon 1); ALMS1 (NM_015120.4) chr2:73,612,990-73,613,041 (partial exon 1); CEP290 (NM_025114.3) exon 5. exon 7, chr12:88,519,017-88,519,039 (partial exon 13), chr12:88,514,049-88,514,058 (partial exon 15), chr12:88,502,837-88,502,841 (partial exon 23), chr12:88,481,551-88,481,589 (partial exon 32), chr12:88,471,605-88,471,700 (partial exon 40); CFTR (NM_000492.3) exon 10; COL4A4 (NM_000092.4) chr2:227,942,604-227,942,619 (partial exon 25); CYP11B2 (NM_000498.3) exons 3-7; DNAl2 (NM_023036.4) chr17:72,308,13672,308,147 (partial exon 12); EVC(NM_153717.2) exon 1; FH (NM_000143.3) exon 1; GAMT(NM_000156.5 exon 1; GLDC(NM_000170.2) exon 1; GNPTAB (NM_024312.4) chr17:4,837,000-4,837,400 (partial exon 2); GNPTG (NM_032520.4) exon 1; HGSNAT(NM_152419.2) exon 1; IDS (NM_000202.6) exon 3; LIFR (NM_002310.5) exon 19; NEB (NM_001271208.1) exons $82-105$; NPC1 (NM_000271.4) chr18:21,123.519-21,123.538 (partial exon 14); PUS1 (NM_025215.5) ; chr12:132,414,446-132,414.532 (partial exon 2); RPGRIP1L (NM_015272.2) exon 23; SGSH (NM_000199.3) chr17:78,194,022-78,194,072 (partial exon 1); SLC6A8(NM_005629.3) exons 3 and 4.
This test will detect variants within the exons and the intron-exon boundaries of the target regions. Variants outside these regions may not be detected, including, but not limited to, UTRs, promoters, and deep intronic areas, or regions that fall into the Exceptions mentioned above. This technology may not detect all small insertion/deletions and is not diagnostic for repeat expansions and structural genomic variation. In addition, a mutation(s) in a gene not included on the panel could be present in this patient.
Variant interpretation and classification was performed based on the American College of Medical Genetics Standards and Guidelines for the Interpretation of Sequence Variants (Richards et al, 2015). All potentially pathogenic variants may be confirmed by either a specific genotyping assay or Sanger sequencing, if indicated. Any benign variants, likely benign variants or variants of uncertain significance identified during this analysis will not be reported.

## Next Generation Sequencing for SMN1

Exonic regions and intron/exon splice junctions of SMN1 and SMN2 were captured, sequenced, and analyzed as described above. Any variants located within exons 2a-7 and classified as pathogenic or likely pathogenic were confirmed to be in either SMN1 or SMN2 using gene-specific long-range PCR analysis followed by Sanger sequencing. Variants located in exon 1 cannot be accurately assigned to either SMN1 or SMN2 using our current methodology, and so these variants are considered to be of uncertain significance and are not reported.

## Copy Number Variant Analysis (Analytical Detection Rate >95\%)

Large duplications and deletions were called from the relative read depths on an exon-by-exon basis using a custom exome hidden Markov model (XHMM) algorithm. Deletions or duplications determined to be pathogenic or likely pathogenic were confirmed by either a custom arrayCGH platform, quantitative PCR, or MLPA (depending on CNV size and gene content). While this algorithm is designed to pick up deletions and duplications of 2 or more exons in length, potentially pathogenic single-exon CNVs will be confirmed and reported, if detected.

## Exon Array (Confirmation method) (Accuracy >99\%)

The customized oligonucleotide microarray (Oxford Gene Technology) is a highly-targeted exon-focused array capable of detecting medically relevant microdeletions and microduplications at a much higher resolution than traditional aCGH methods. Each array matrix has approximately $180,00060-m e r$ oligonucleotide probes that cover the entire genome. This platform is designed based on human genome NCBI Build 37 (hg1g) and the CGH probes are enriched to target the exonic regions of the genes in this panel.

## Quantitative PCR (Confirmation method) (Accuracy >99\%)

The relative quantification PCR is utilized on a Roche Universal Library Probe (UPL) system, which relates the PCR signal of the target region in one group to another. To test for genomic imbalances, both sample DNA and reference DNA is amplified with primer/probe sets that specific to the target region and a control region with known genomic copy number. Relative genomic copy numbers are calculated based on the standard $\triangle \triangle C t$ formula.

## Long-Range PCR (Analytical Detection Rate >99\%)

Long-range PCR was performed to generate locus-specific amplicons for CYP21A2, HBA1 and HBA2 and GBA. The PCR products were then prepared for short-read NGS sequencing and sequenced. Sequenced reads were mapped back to the original genomic locus and run through the bioinformatics pipeline. If indicated, copy number from MLPA was correlated with the sequencing output to analyze the results. For CYP21A2, a certain percentage of healthy individuals carry a duplication of the CYP21A2 gene, which has no clinical consequences. In cases where two copies of a gene are located on the same chromosome in tandem, only the second copy will be amplified and assessed for potentially pathogenic variants, due to size limitations of the PCR reaction. However, because these alleles contain at least two copies of the CYP21A2 gene in tandem, it is expected that this patient has at least one functional gene in the tandem allele and this patient is therefore less likely to be a carrier. When an individual carries both a duplication allele and a pathogenic variant, or multiple pathogenic variants, the current analysis may not be able to determine the phase (cis/trans configuration) of the CYP21A2 alleles identified. Family studies may be required in certain scenarios where phasing is required to determine the carrier status.

## Residual Risk Calculations

Carrier frequencies and detection rates for each ethnicity were calculated trough the combination of internal curations of $>30,000$ variants and genomic frequency data from $>138,000$ individuals across seven ethnic groups in the gnomAD database. Additional variants in HGMD and novel deleterious variants were also incorporated into the calculation. Residual risk values are calculated using a Bayesian analysis combining the a priori risk of being a pathogenic mutation carrier (carrier frequency) and the detection rate. They are provided only as a guide for assessing approximate risk given a negative result, and values will vary based on the exact ethnic background of an individual. This report does not represent medical advice but should be interpreted by a genetic counselor, medical geneticist or physician skilled in genetic result interpretation and the relevant medical literature.

## Personalized Residual Risk Calculations

Agilent SureSelect ${ }^{T M}$ XT Low-Input technology was utilized in order to create whole-genome libraries for each patient sample. Libraries were then pooled and sequenced on the Illumina NovaSeq platform. Each sequencing lane was multiplexed to achieve 0.4-2x genome coverage, using paired-end 100 bp reads. The sequencing data underwent ancestral analysis using a customized, licensed bioinformatics algorithm that was validated in house. Identified sub-ethnic groupings were binned into one of 7 continental-level groups (African, East Asian, South Asian, Non-Finnish European, Finnish, Native American, and Ashkenazi Jewish) or, for those ethnicities that matched poorly to the continental-level groups, an $8^{\text {th }}$ "unassigned" group, which were then used to select residual risk values for each gene. For individuals belonging to multiple highlevel ethnic groupings, a weighting strategy was used to select the most appropriate residual risk. For genes that had insufficient data to calculate ethnic-specific residual risk values, or for sub-ethnic groupings that fell into the "unassigned" group, a "worldwide" residual risk was used. This "worldwide" residual risk was calculated using data from all available continental-level groups.

## Sanger Sequencing (Confirmation method) (Accuracy >99\%)

Sanger sequencing, as indicated, was performed using BigDye Terminator chemistry with the ABI 3730 DNA analyzer with target specific amplicons. It also may be used to supplement specific guaranteed target regions that fail NGS sequencing due to poor quality or low depth of coverage (<20 reads) or as a confirmatory method for NGS positive results. False negative results may occur if rare variants interfere with amplification or annealing.

Carrier screening report
Donor 6427
Date of Birth:
Sema4 ID:

Please note these tests were developed and their performance characteristics were determined by Mount Sinai Genomics, Inc. They have not been cleared or approved by the FDA. These analyses generally provide highly accurate information regarding the patient's carrier or affected status. Despite this high level of accuracy, it should be kept in mind that there are many potential sources of diagnostic error, including misidentification of samples, polymorphisms, or other rare genetic variants that interfere with analysis. Families should understand that rare diagnostic errors may occur for these reasons.

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Additional disease-specific references available upon request.


[^0]:    *No single test can screen for all genetic disorders. A negative screening result significantly reduces, but cannot eliminate, the risk for these conditions in a pregnancy.
    **Donor residual risk is the chance the donor is still a carrier after testing negative.

